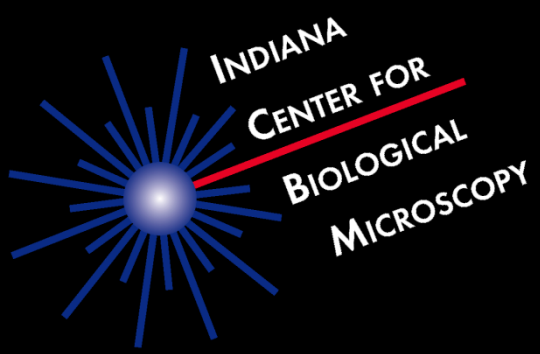
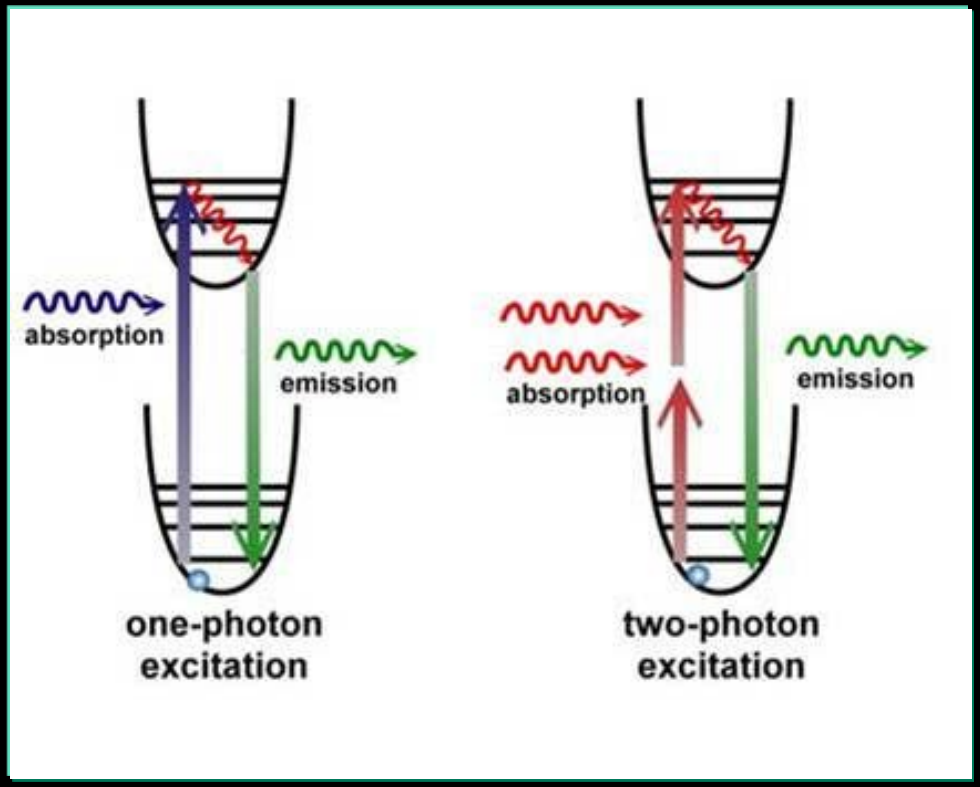


Intravital multiphoton microscopy
Principles and challenges

Ken Dunn, PhD
Scientific Director
Indiana Center for Biological Microscopy
Indiana University Medical Center

Multiphoton fluorescence excitation

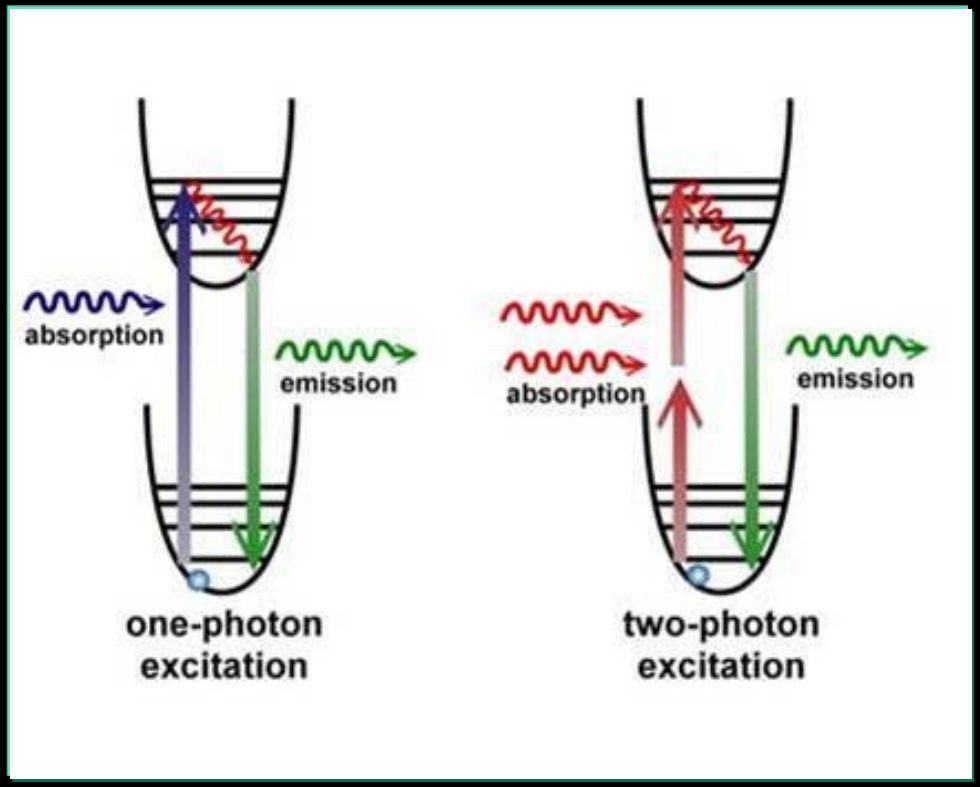


Biophotonics Imaging Lab, Univ. Illinois Urbana-Champaign

Fluorescence can be stimulated by the absorption of one photon of a particular energy level.

Fluorescence can also be stimulated by the simultaneous absorption of multiple, low-energy photons

Multiphoton fluorescence excitation
Requires an enormous density of photons



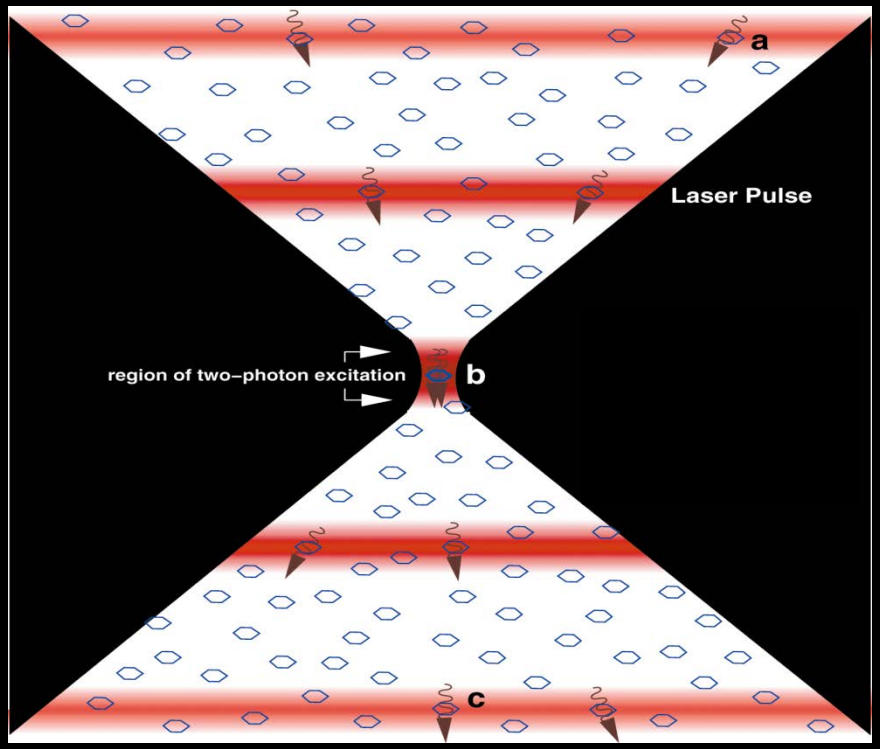
Biophotonics Imaging Lab, Univ. Illinois Urbana-Champaign

Fluorescence can be stimulated by the absorption of one photon of a particular energy level.

Fluorescence can also be stimulated by the simultaneous absorption of multiple, low-energy photons

"Simultaneous" means within ~ 10 attoseconds (10^{-17} seconds)
Two-photon fluorescence excitation requires an enormous flux of photons

Multiphoton fluorescence excitation Requires an enormous density of photons



Dave Piston

Detectible two-photon fluorescence excitation requires peak power on the order of 200 GW/cm^2

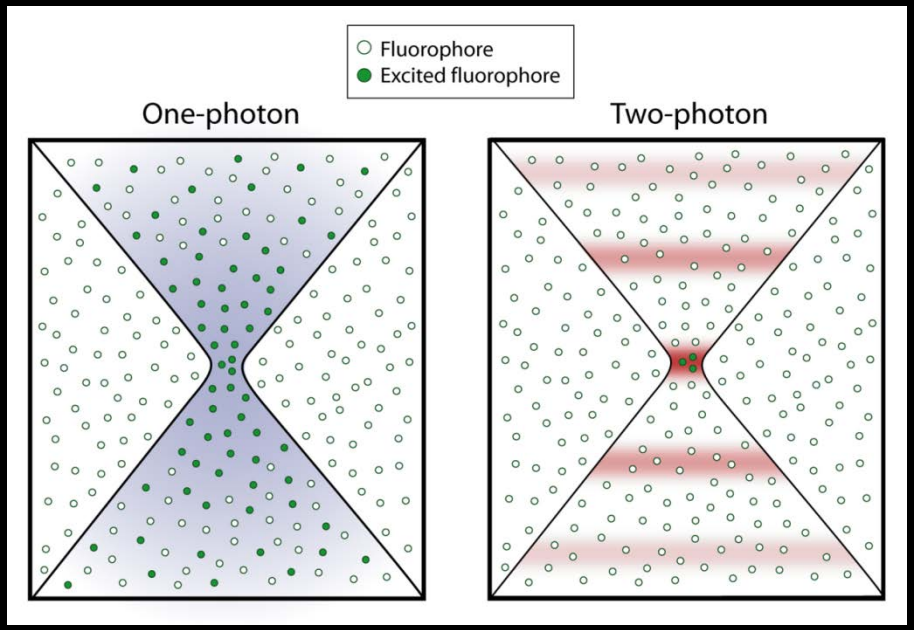
~ 300,000x the surface of the sun

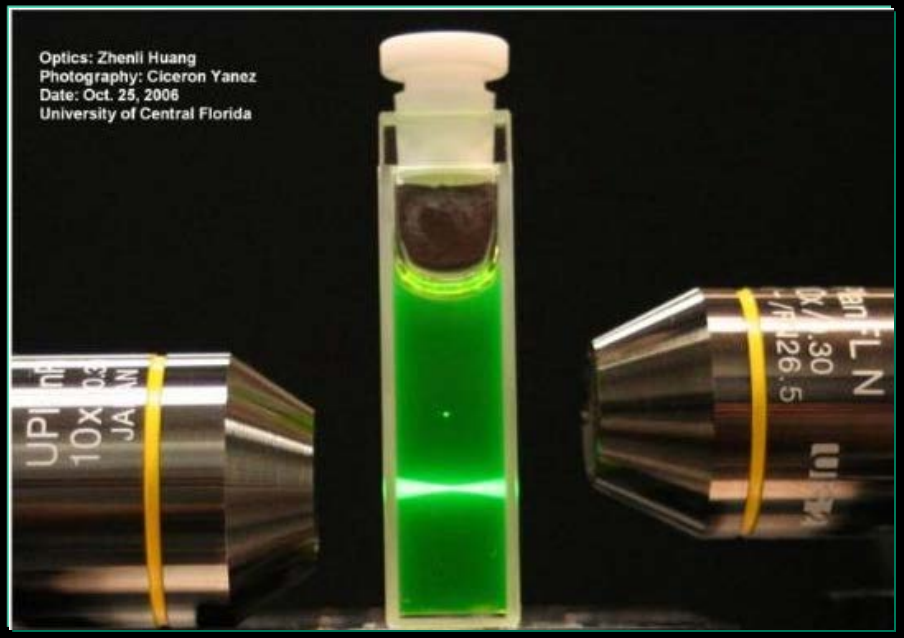
For multiphoton microscopy, this power is provided by focusing powerful pulsed lasers through high NA objective lenses.

Photon density decreases with the 4th power of distance from the lens focus

- Two-photon absorption occurs ONLY in the sub-femtoliter volume at the focus

Multiphoton fluorescence excitation is restricted to the focus of a lens for microscopy

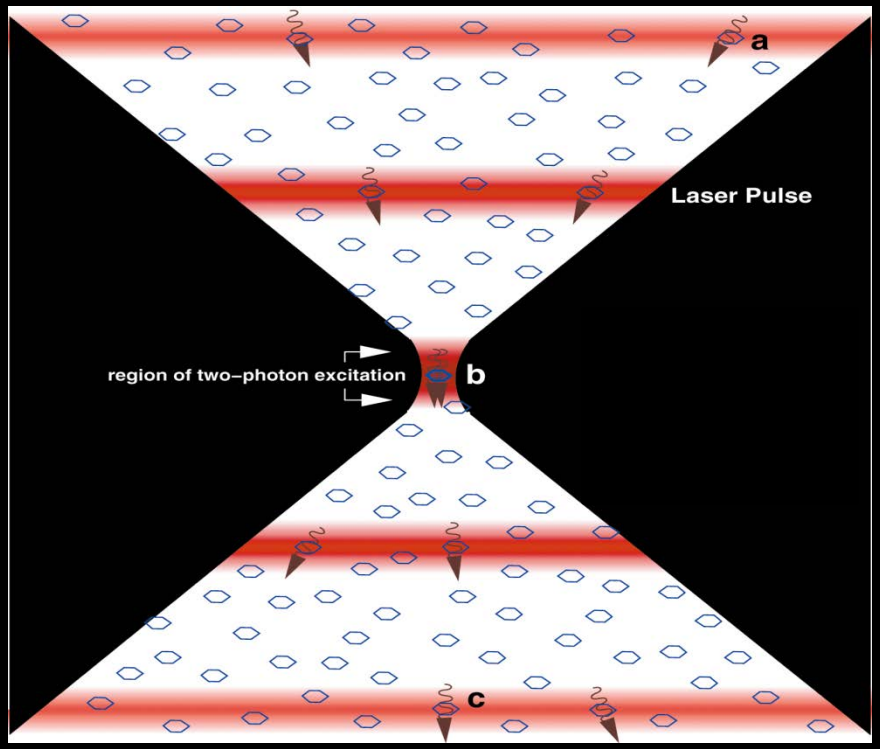




Multiphoton microscopy provides the capability to stimulate fluorescence in a specific sub-femtoliter volume in a sample -

High resolution, 3D microscopy

Multiphoton fluorescence excitation using pulsed (mode-locked) lasers

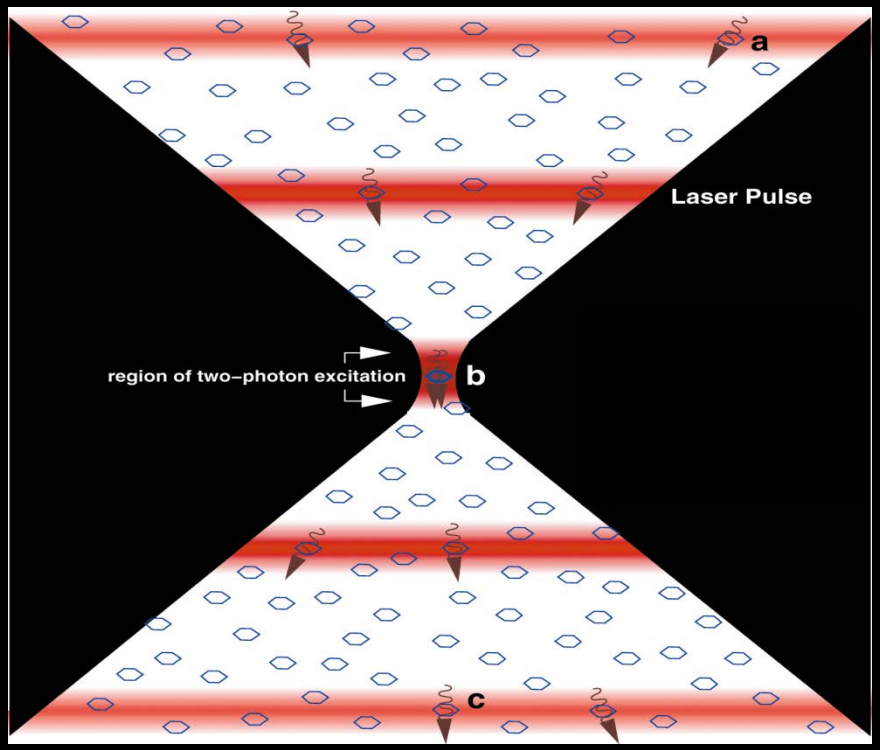


Dave Piston

Pulsing laser illumination into ~100 femtosecond pulses at a rate of 80 MHz provides peak power sufficient for multiphoton excitation, but average power low enough to minimize damage.

(e.g. 28 mW average provides 200 GW/cm^2 peak with 0.8 NA objective)

Misconceptions about multiphoton fluorescence excitation – Pulsing the laser limits the volume of excitation

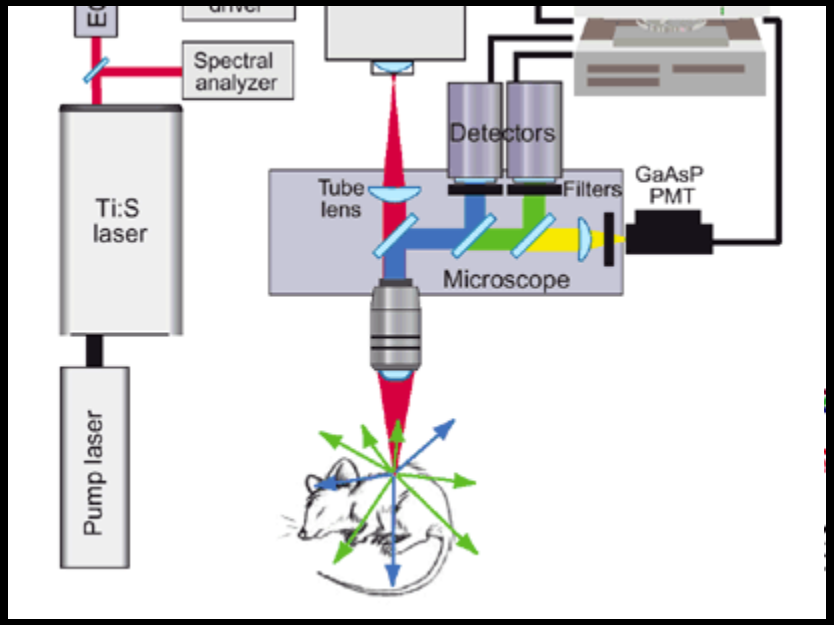


Dave Piston

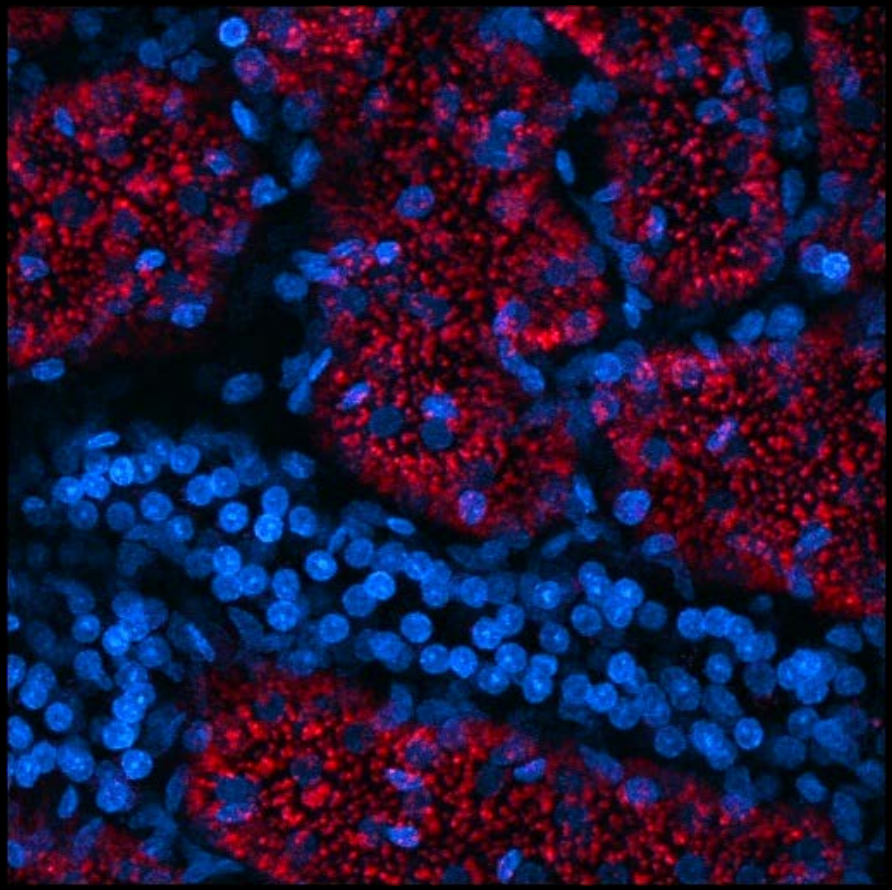
Pulses are actually ~ 45 microns long and ~ 4 meters apart

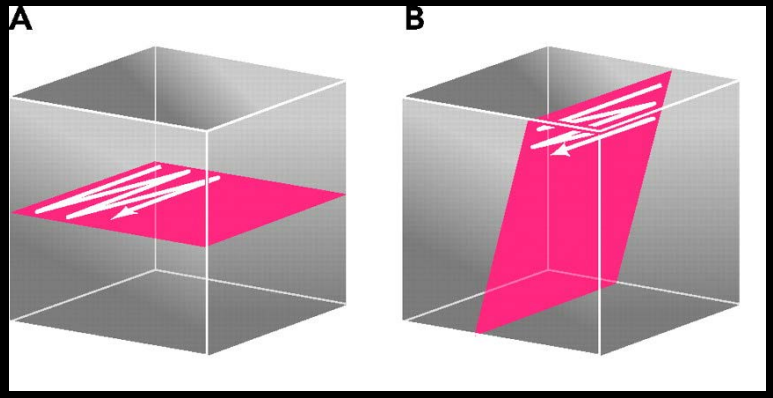
To be accurate, the pulses as displayed here should be ~ 5 miles apart

Making an image using multi-photon microscopy – point scanning



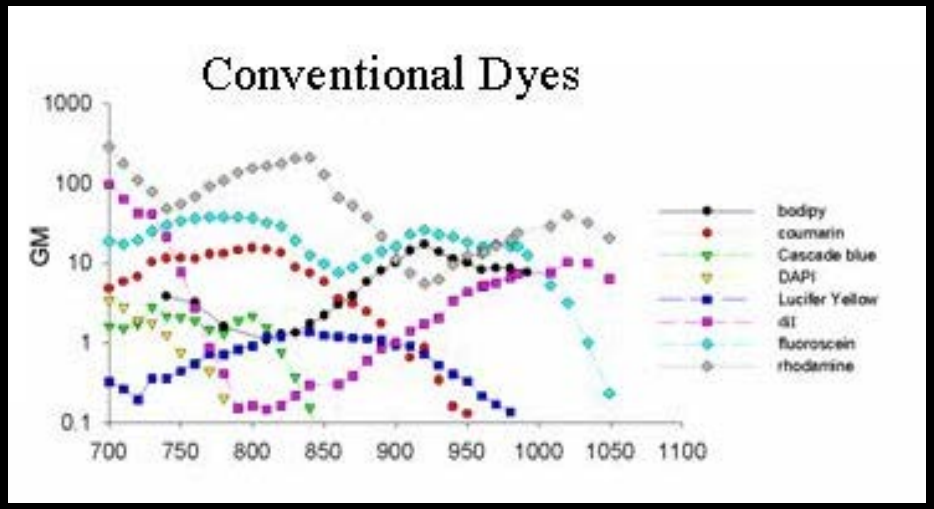
Zipfel et al., 2003

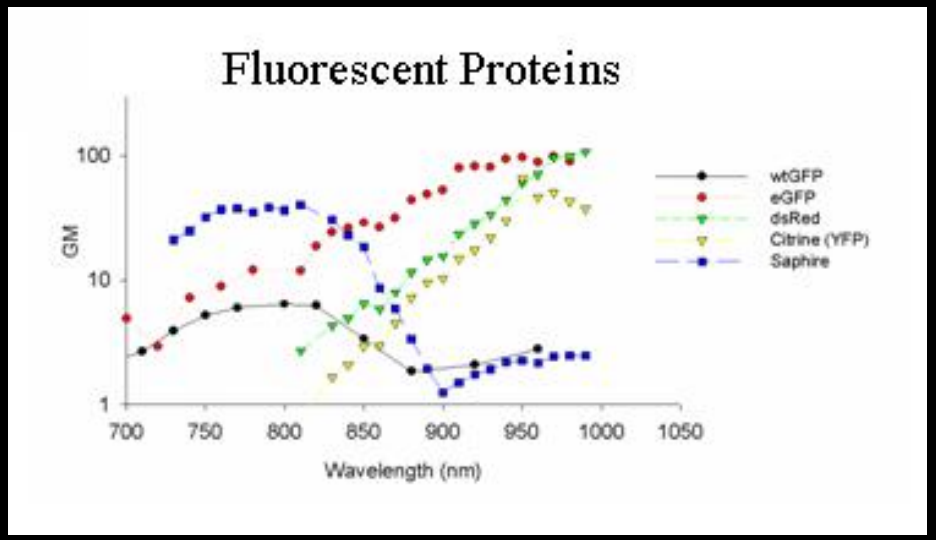




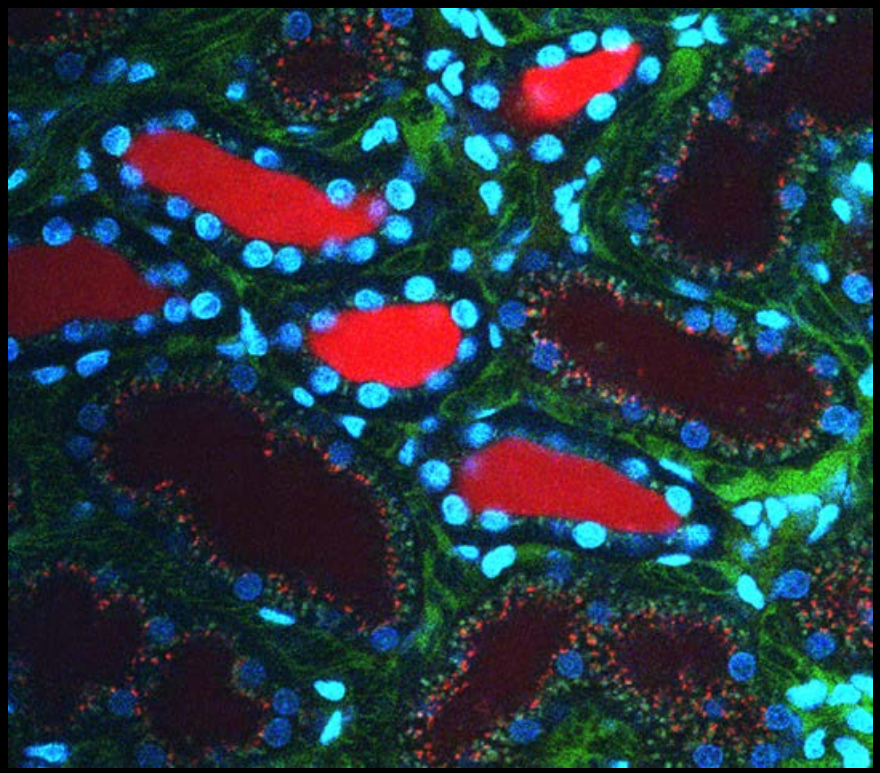
Göbel and Helmchen 2007

Two-photon action cross sections





Developmental Resource for Biophysical Imaging Opto-electronics, Cornell
(Watt Webb and Warren Zipfel)



10 kD Rhodamine Dextran,
500 kD Fluorescein Dextrn
Hoechst 33342

Dunn et al 2002

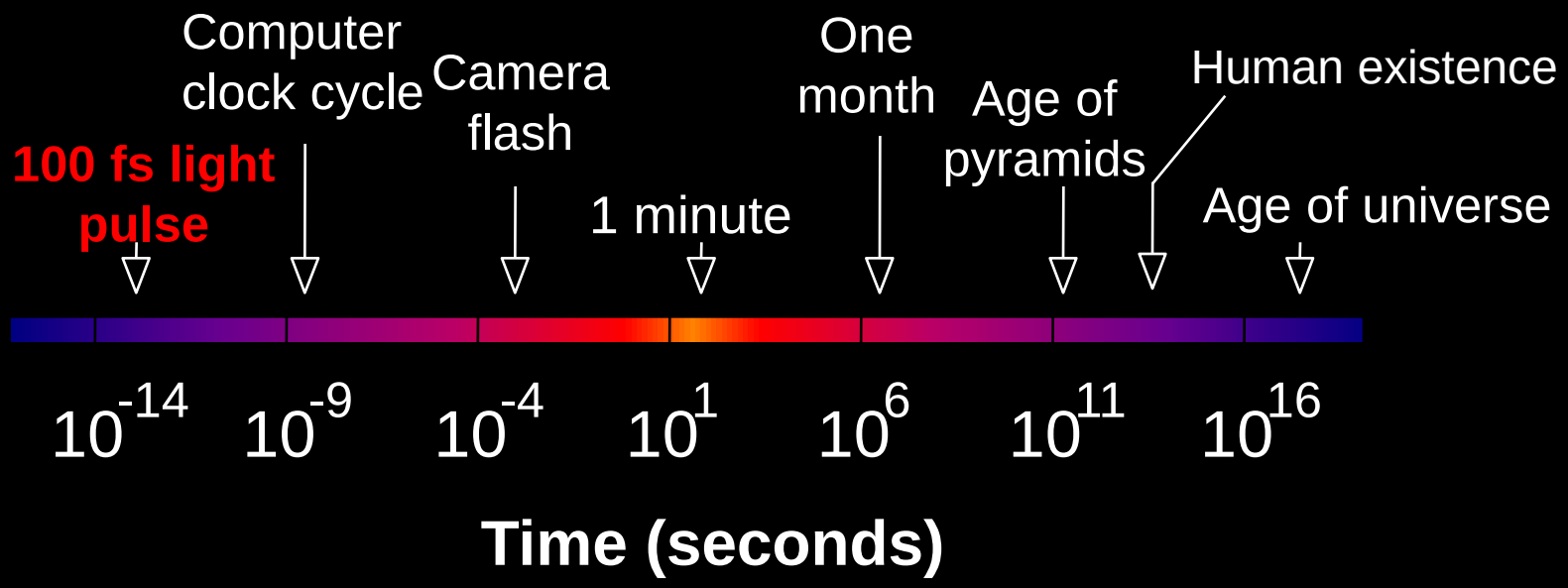
Fun facts about multiphoton microscopy

How frequently does multiphoton fluorescence excitation occur outside of a microscope?

Calculations made by Winfried Denk indicate that a molecule of rhodamine B exposed to direct sunlight will experience:

- A one-photon absorption around once per second.
- A two photon absorption once every 10,000 years.
- A three-photon absorption . . . never in the history of the universe.

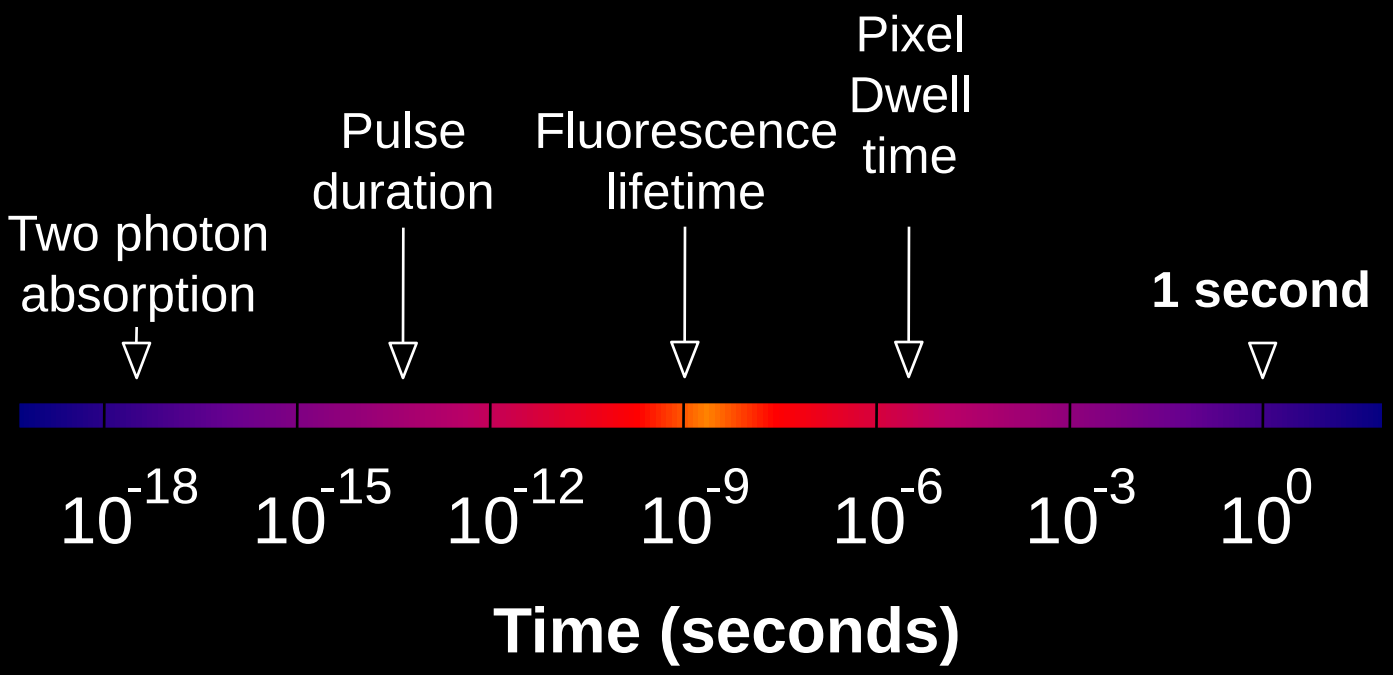
Fun facts about multiphoton microscopy
Relative time scales



100 fs is to 10 minutes as 10 minutes is to the age of the universe.

Rick Trebino, University of Georgia

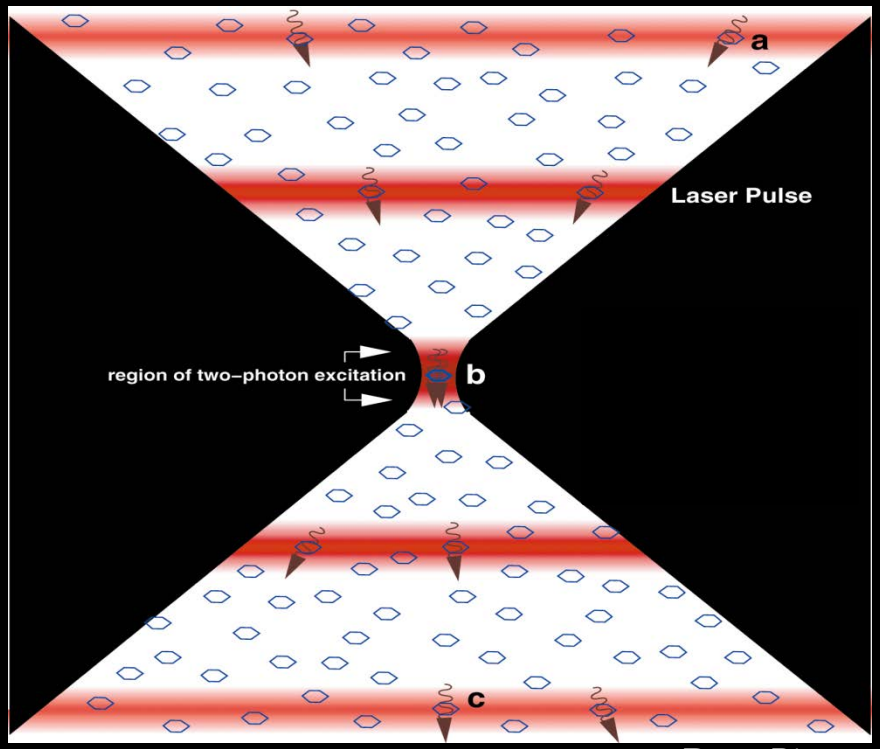
Fun facts about multiphoton microscopy
Relative time scales



- Fluorescence lifetimes are ~ 1 billion times the length of the virtual intermediate state
- At 80 MHz, the laser pulses are spaced ~ 3 fluorescence lifetimes apart (serendipitous)
- A fluorophore will be illuminated ~320 times during a 4 microsecond pixel dwell time

Fun facts about multiphoton microscopy

When the Titanium-sapphire laser is on, it's mostly off.



Dave Piston

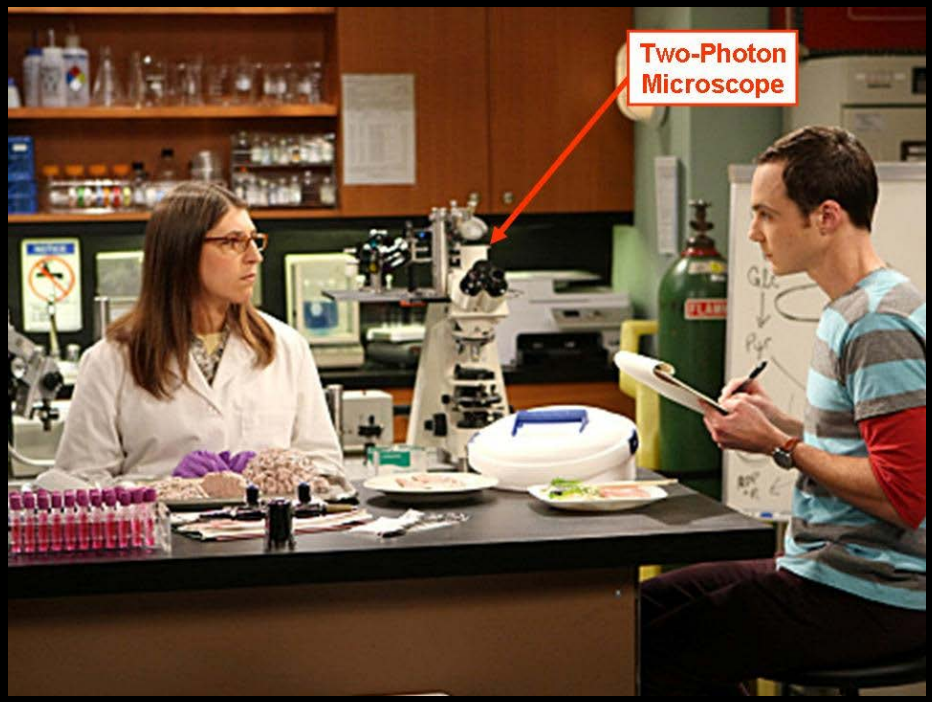
The sample is illuminated with 100 femtosecond pulses at a rate of 80 million times per second.
Only 0.001% of the time

For sense of scale, this duty cycle is equivalent to a one second pulse occurring once a day.

Multiphoton microscope designs

Gregor Lab, Physics Princeton

Nikon A1RMP



Google search result

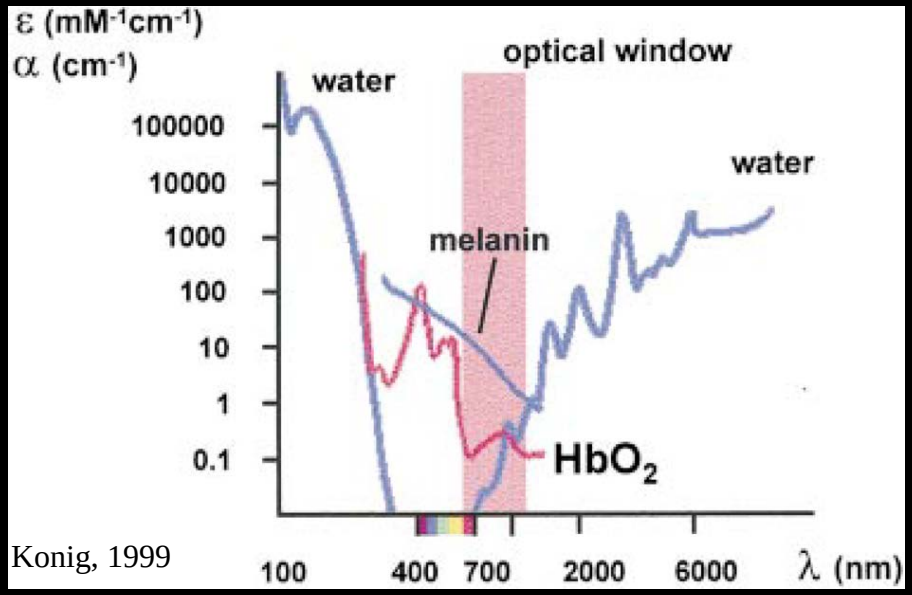
Why multiphoton microscopy?

Complex

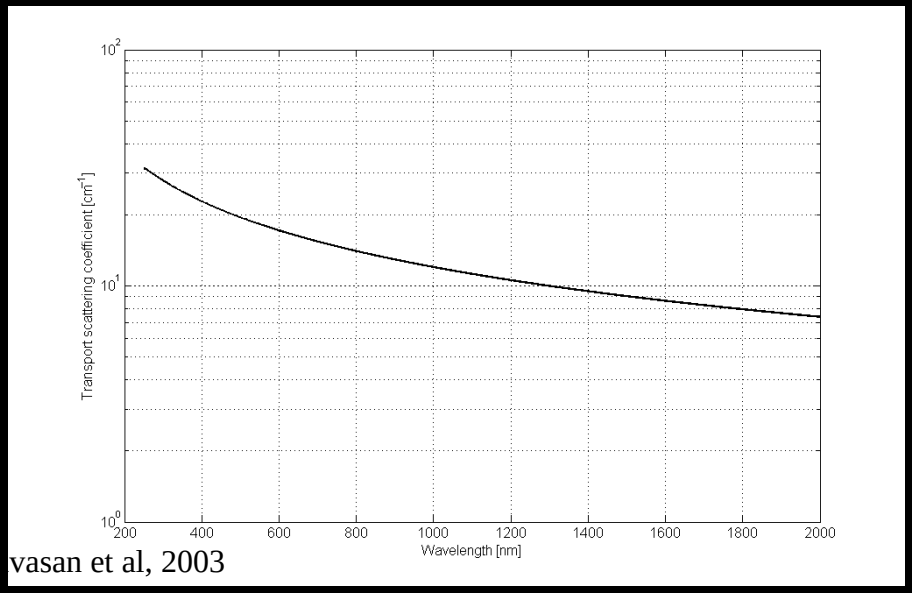
Expensive

Poorer resolution - 0.3 by 0.9 microns versus .21 by .75 microns

Why multiphoton microscopy?



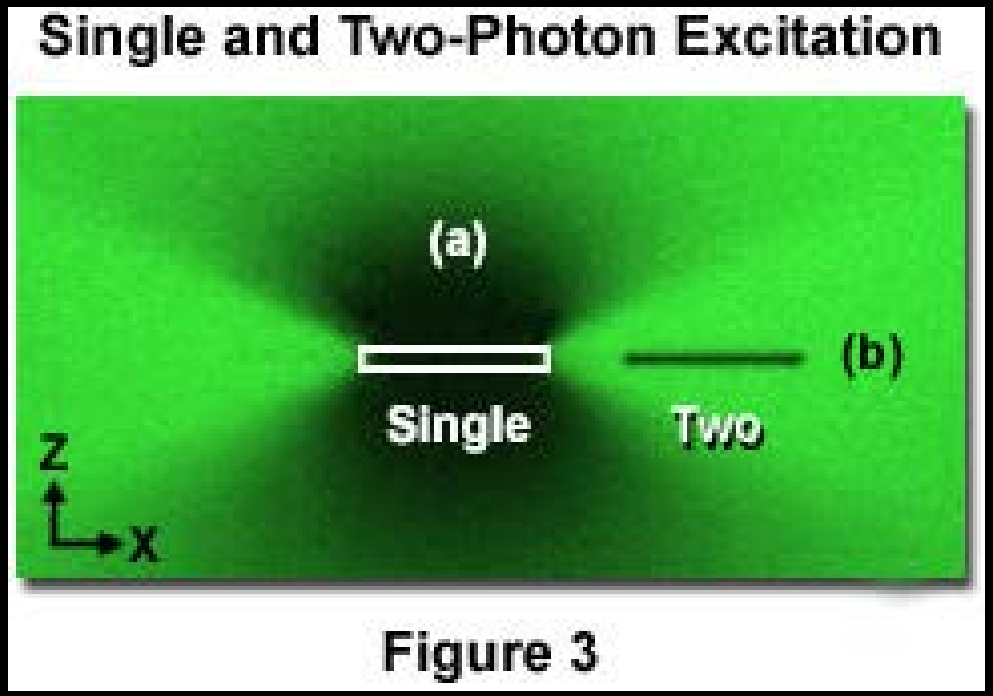
Konig, 1999



vasan et al, 2003

- IR light penetrates deeper, with less damage

Why multiphoton microscopy?



Nikon MicroscopyU

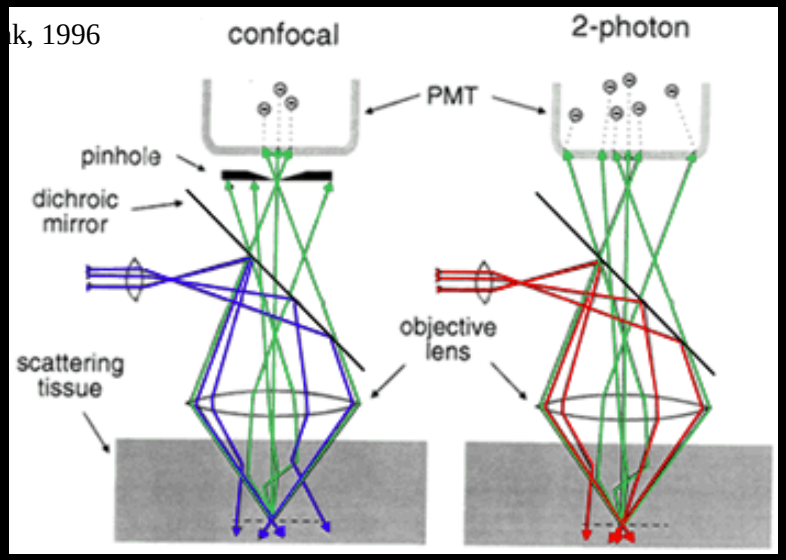
- IR light penetrates deeper, with less damage
- Photobleaching only in the focal plane

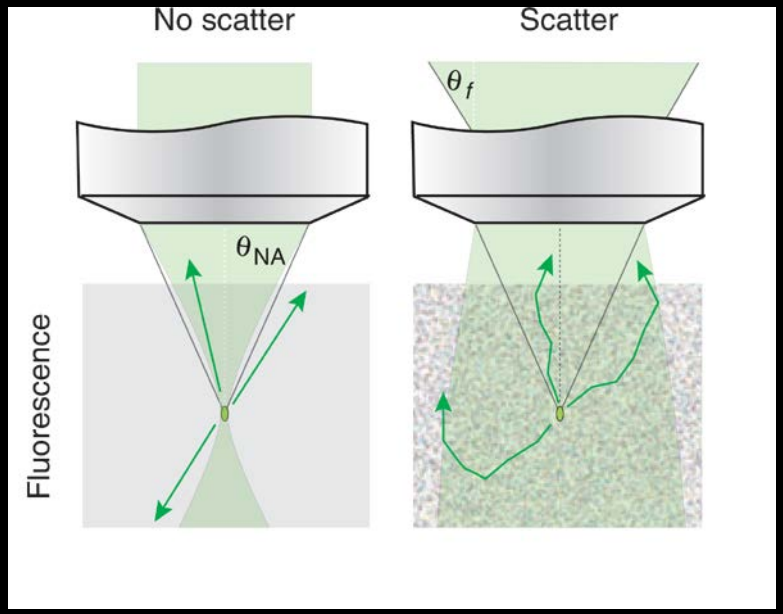
Misconceptions about multiphoton fluorescence excitation - Multiphoton microscopy causes less photobleaching

Since only the focal plane is excited, image volumes can be collected without cumulative photodamage and photobleaching. Reimplanted hamster embryos develop normally after being imaged every 15 minutes for 24 hours.

However, the rate of photobleaching increases disproportionately with illumination, increasing with the third or even fourth power at high levels. Cellular toxicity increases abruptly at power levels above around 10 mW.

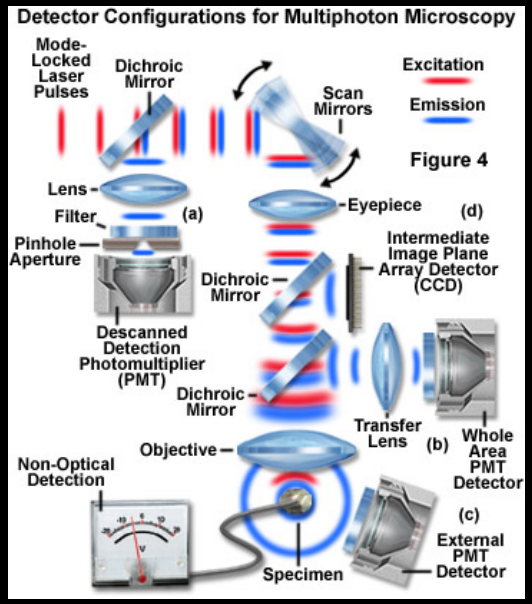
Why multiphoton microscopy?

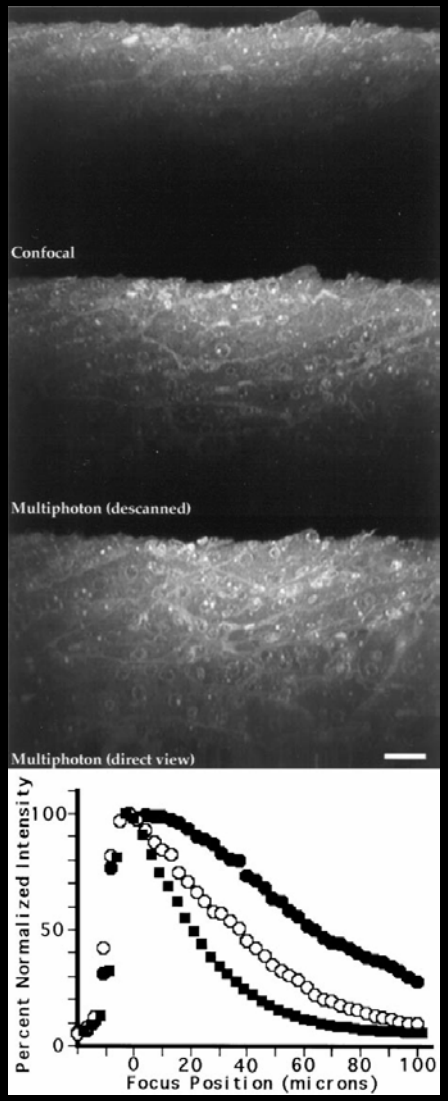




- IR light penetrates deeper, with less damage
- Photobleaching only in the focal plane
- Optical sectioning without an emission aperture - less loss to scattering

Why multiphoton microscopy?

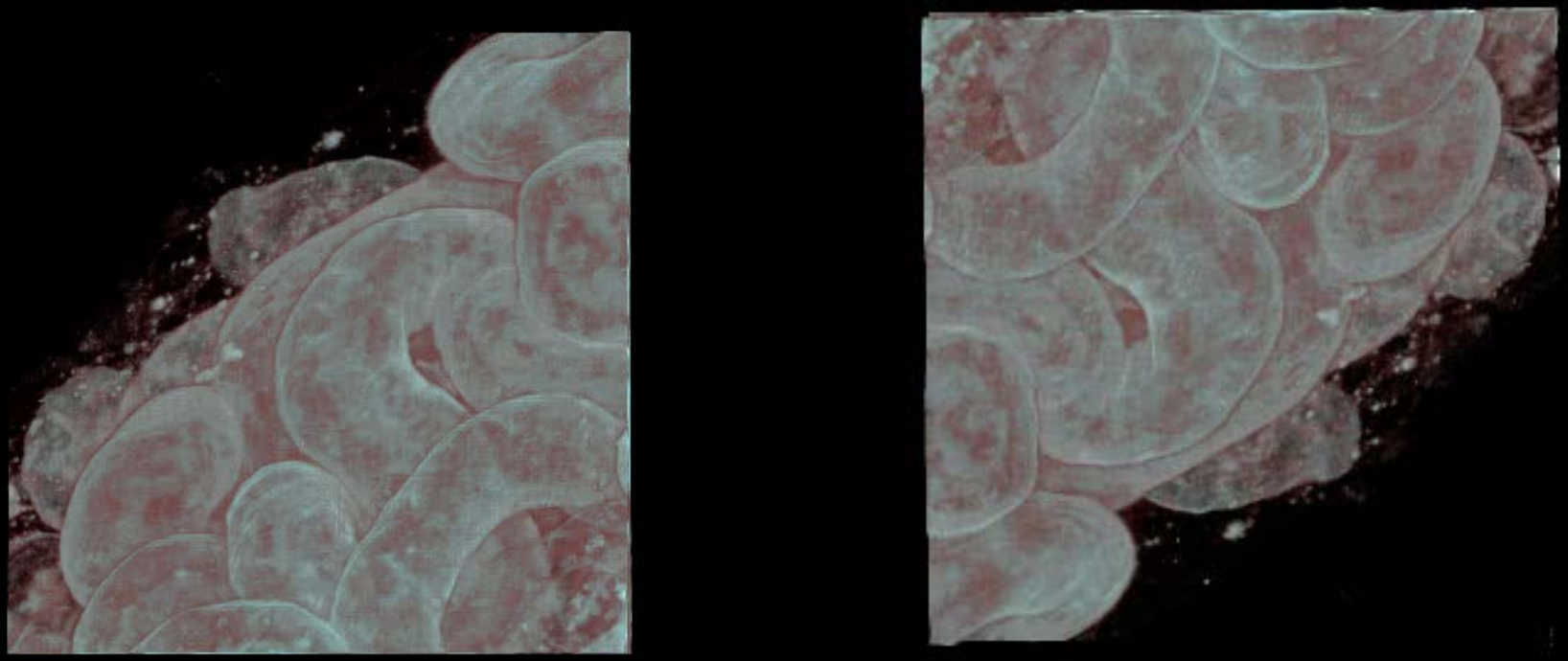




Centonze and White, 1998

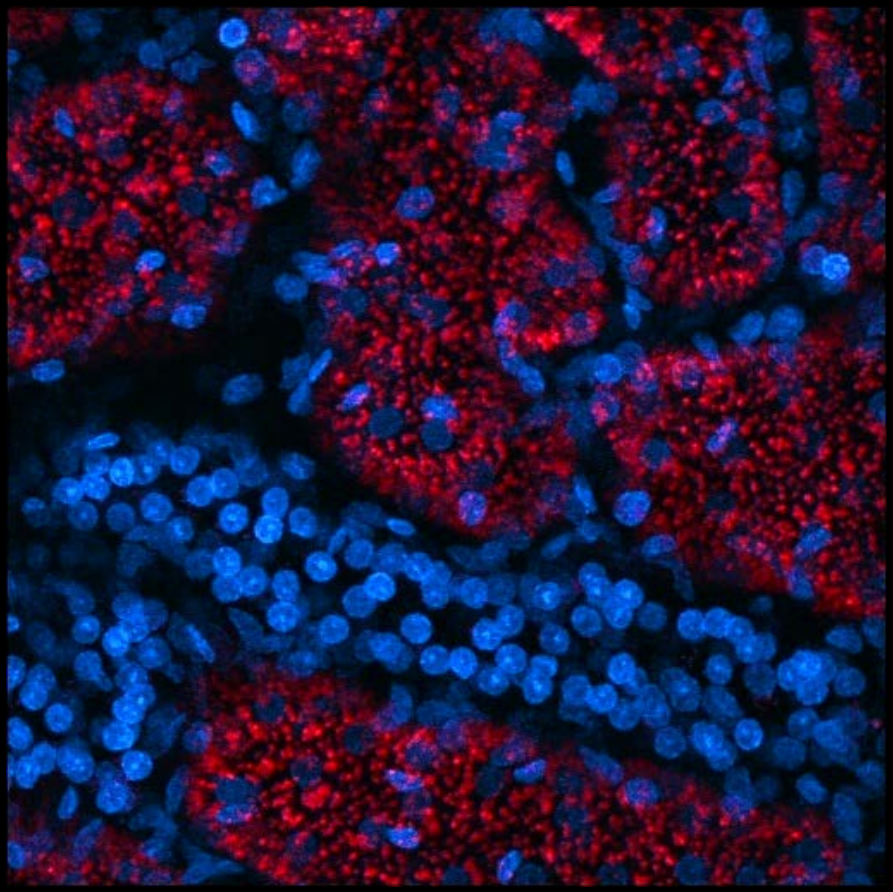
- IR light penetrates deeper, with less damage
- Photobleaching only in the focal plane
- Optical sectioning without an emission aperture - less loss to scattering

Why multi-photon microscopy?
Deep-tissue imaging at high resolution



120 micron thick volume of mouse embryonic kidney -
Carrie Phillips, Nephrology

Why multi-photon microscopy? Intravital microscopy



- Intravital microscopy provides submicron resolution and nanomolar sensitivity in under a second.
- Multiphoton microscopy has extended the reach of intravital microscopy to the scale of tissues and the functional components of organs.

Applications of intravital multiphoton microscopy in biomedical research

Brain function and pathology

- Neural development and activity - Winfried Denk, Karel Svoboda
- Neural activity and blood flow in epilepsy - Gyorgy Buzsaki
- Alzheimer's disease - Brian Bacskai, Brad Hyman
- Astrocyte-neuron signaling - Jan Nedergard
- Vascular function in the brain - David Kleinfeld

Tumor biology

- Tumor cell dynamics and metastasis - John Condelis, John Segal
- Angiogenesis/vascular function, gene expression - Rakesh Jain

Immunology

- T-cell interactions and dynamics- Michael Dustin, Ullrich Von Andrian, Michael Cahalan
- Immune surveillance in the brain - Fritjof Helmchen, W.B. Gan

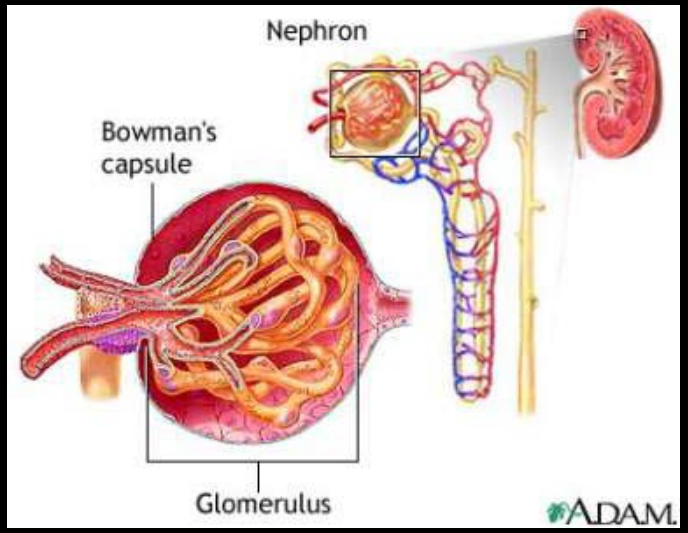
Liver function

- Mitochondria and liver injury - John Lemasters
- In vivo analysis of hepatobiliary transport - H.S Lee

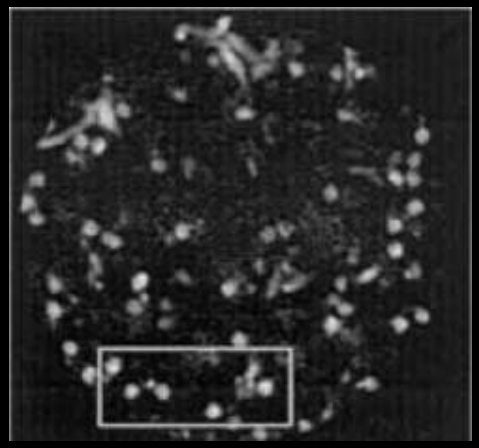
Kidney function

- Pathology and treatment of renal ischemia - Bruce Molitoris
- Pathobiology of diabetes - Katherine Kelly
- Mechanisms of renal septic injury - Pierre Dagher
- Microvascular function in renal injury - Tim Sutton
- Regulation of glomerular function - Janos Peti-Peterdi

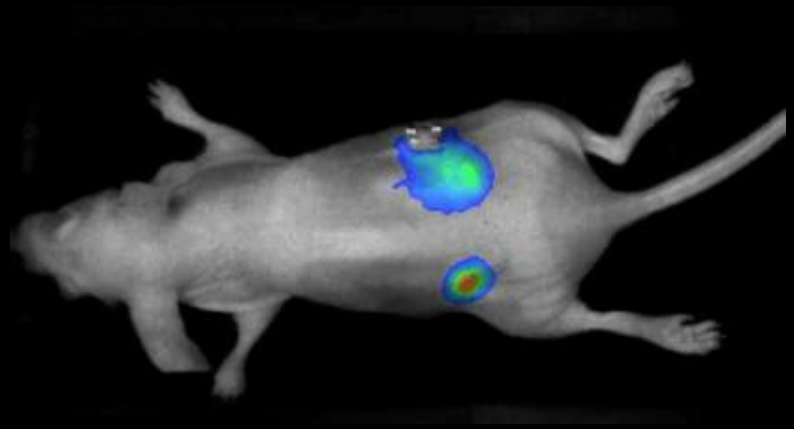
In vivo imaging of the kidney



microPET
Johnstrom et al. 2002. Clin. Sci

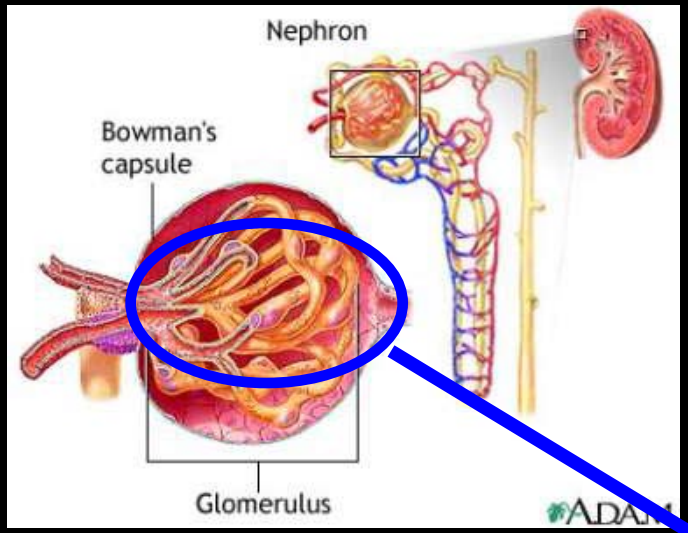


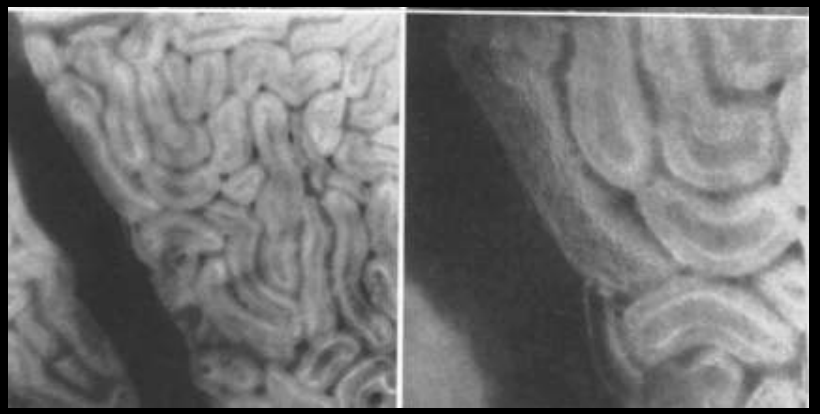
microCT
Toyota et al. 2004. Kid. Int.



Bioluminescence
Que, Kajizel and Löwik, LUMC, Netherlands

Intravital microscopy of the kidney





Wide-field microscopy
Steinhausen et al., 1963.

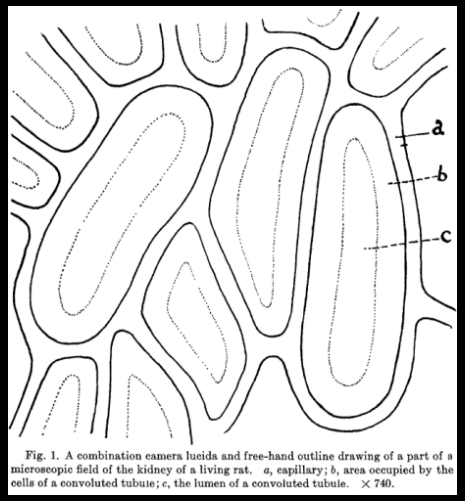
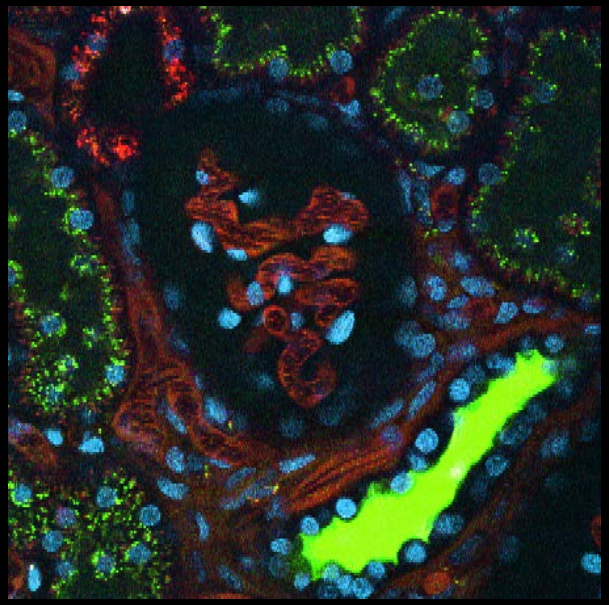
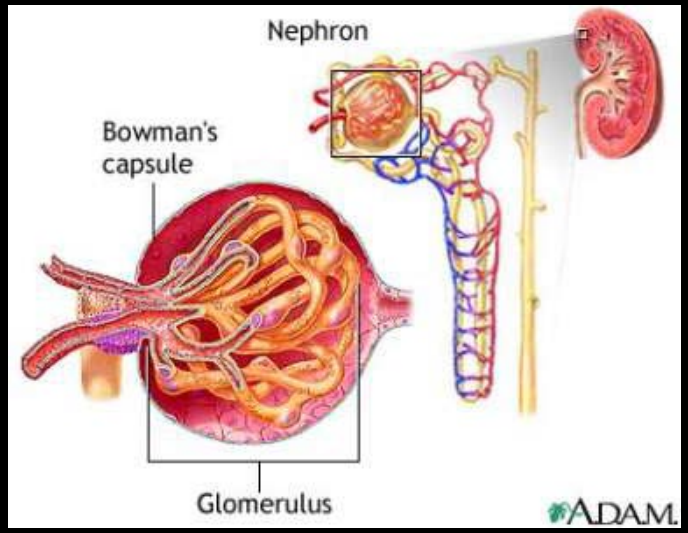


Fig. 1. A combination camera lucida and free-hand outline drawing of a part of a microscopic field of the kidney of a living rat. *a*, capillary; *b*, area occupied by the cells of a convoluted tubule; *c*, the lumen of a convoluted tubule. × 740.

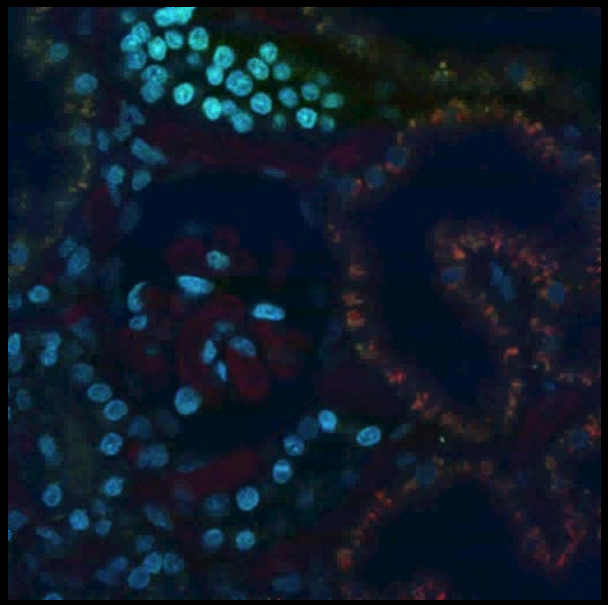
Wide field microscopy
Edwards and Marshall, 1924

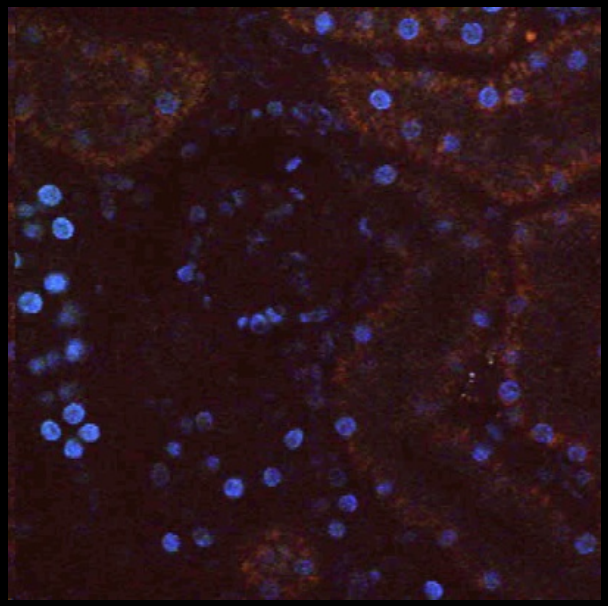


Multiphoton microscopy



Dynamic intravital microscopy of the kidney



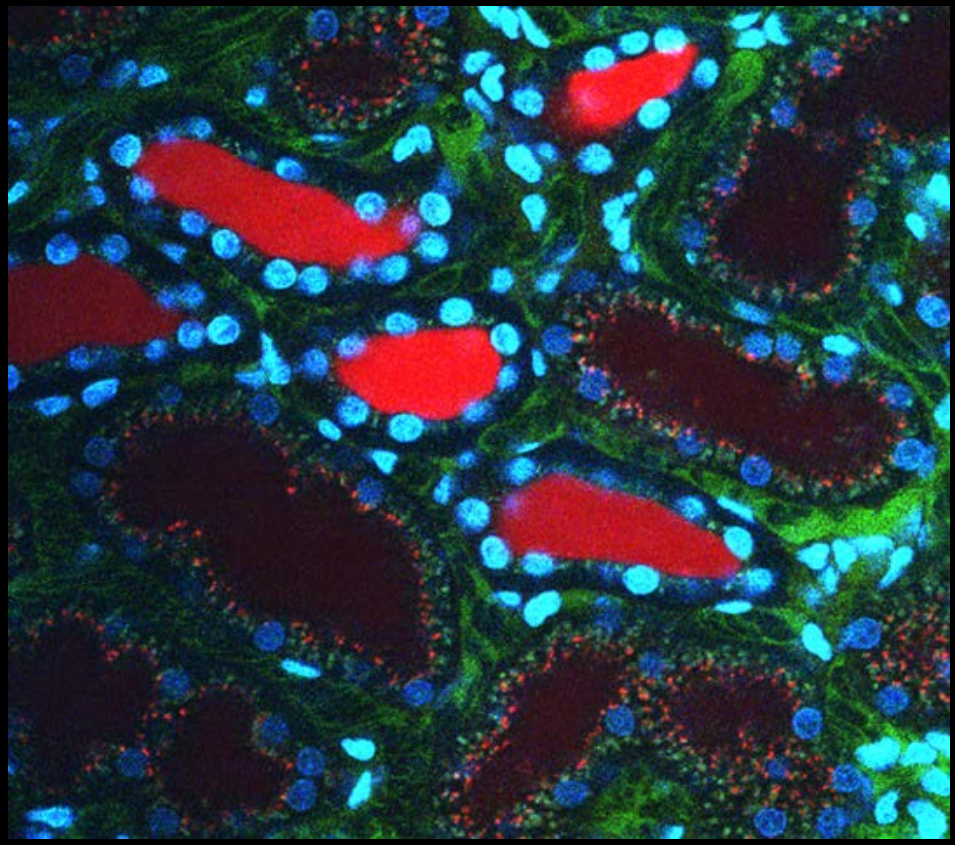


Ruben Sandoval and Bruce Molitoris

Intravital microscopy of multiple kidney functions via intravenous injection of fluorescent probes

Simultaneous imaging of a rat injected with fluorescent dextrans and Hoechst 33342 reveals multiple processes

- glomerular filtration
- proximal tubule endocytosis
- tubular solute concentration
- tubular flow
- capillary blood flow
- vascular permeability
- apoptosis
- tubular sloughing

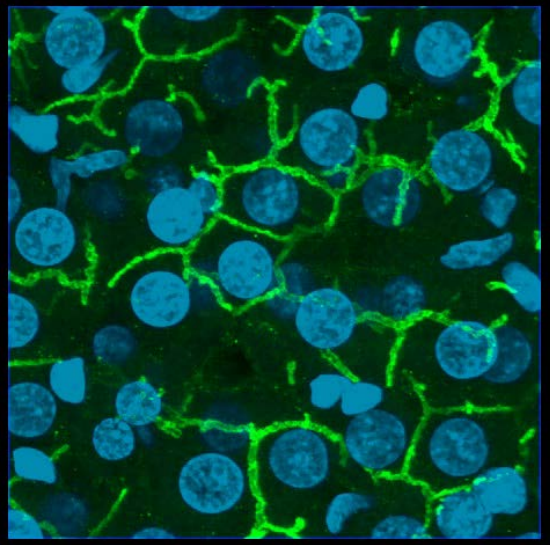


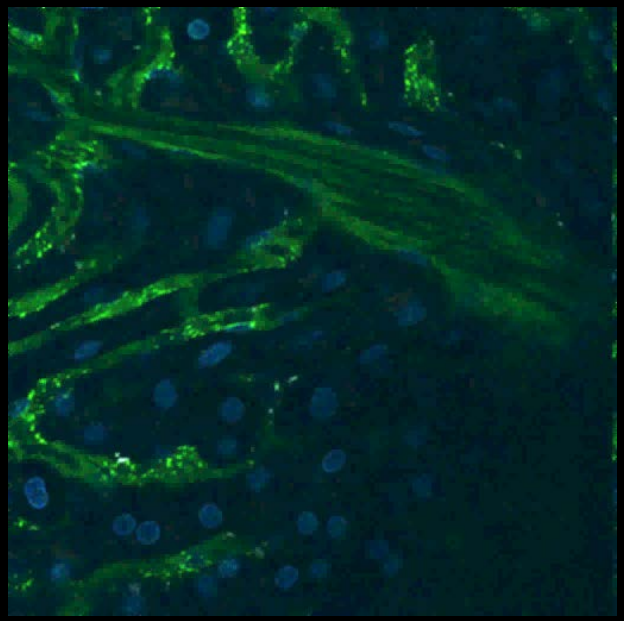
Dunn et al., 2002. AJP Cell. 283:C905-C916

Challenges of intravital microscopy

- Motion artifacts from respiration and heartbeat
- Slow image capture rate
- Limited depth at which useful images can be collected
- Discrimination of different fluors
- Introduction of fluorescent indicators

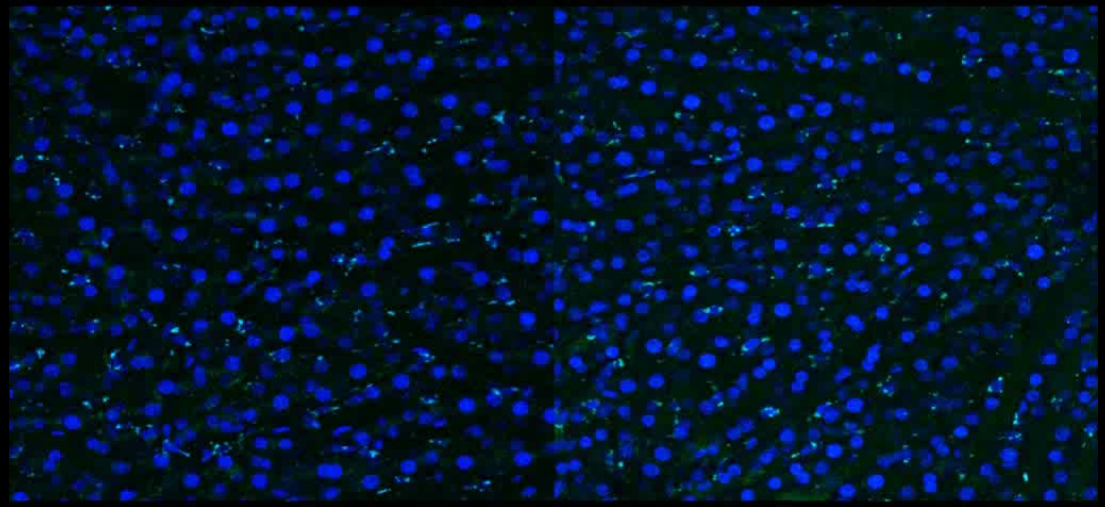
Motion artifacts in intravital microscopy



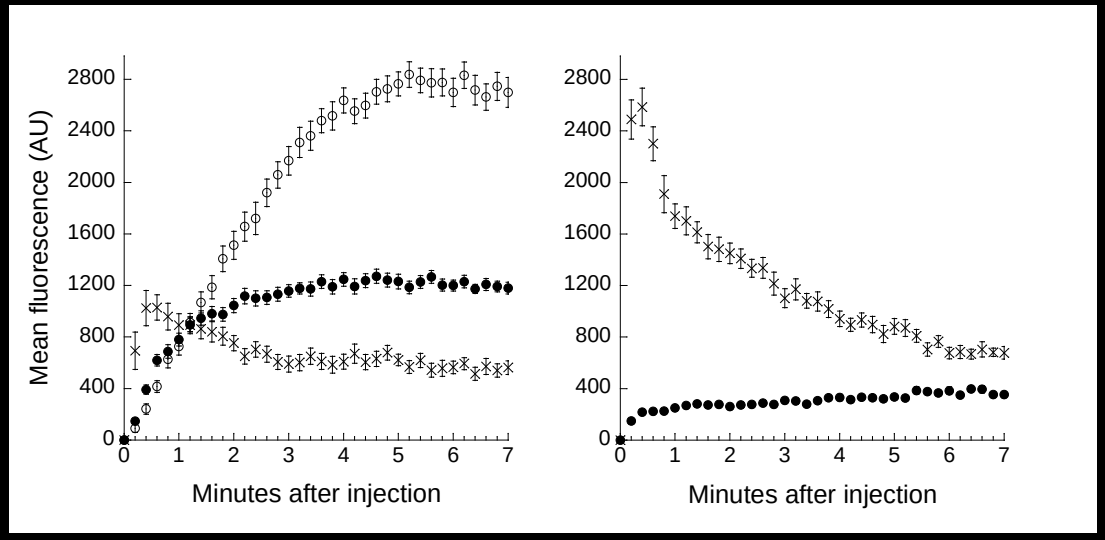


- Motion artifacts can obscure structures

Motion artifacts in intravital microscopy

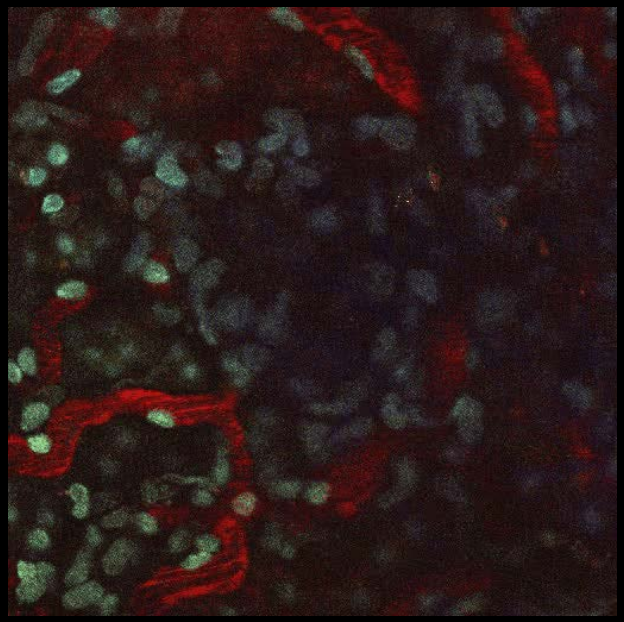


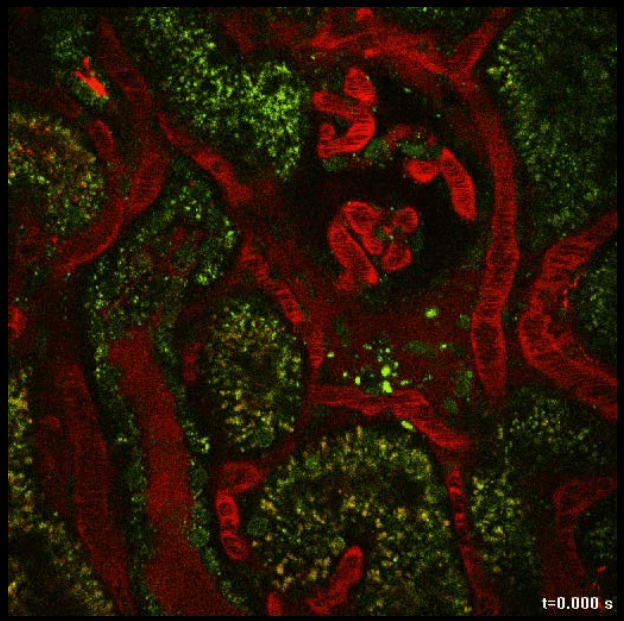
Vehicle Rifampin



- Motion artifacts can obscure structures
- Motion artifacts can confound quantitation

Motion artifacts in intravital microscopy

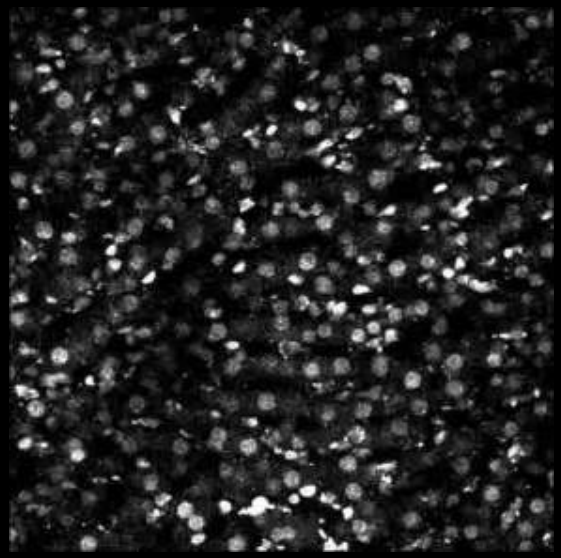




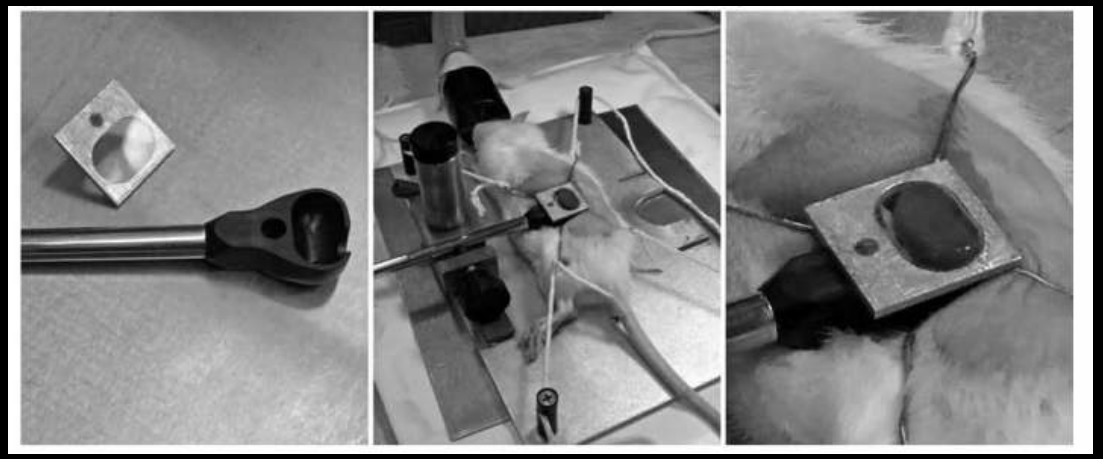
- Motion artifacts can obscure structures
- Motion artifacts can confound quantitation
- Motion artifacts complicate characterizing structural changes

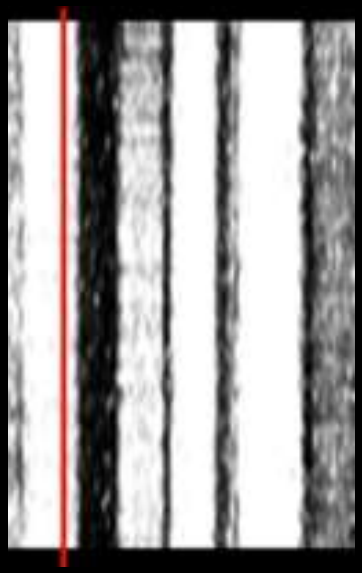
Kang, Toma, Sipos, McCulloch and Peti-Peterdi. 2006. AJP Renal

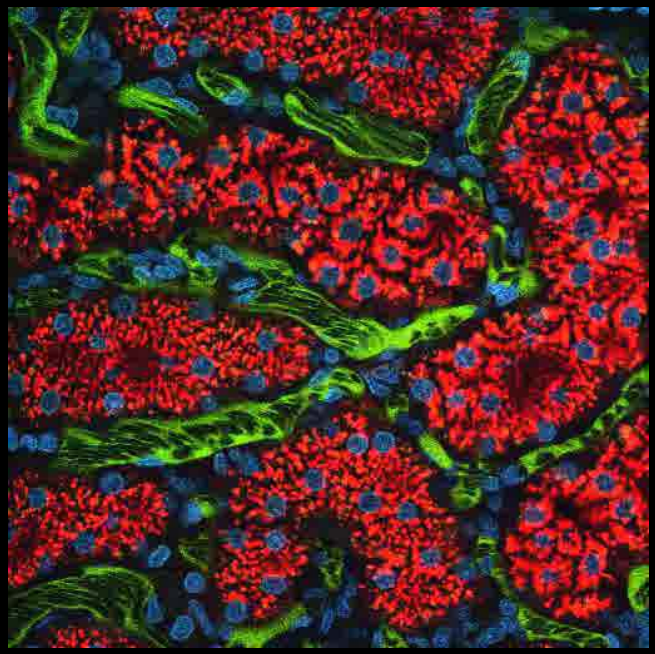
Reducing motion artifacts in intravital microscopy - Immobilize the organ



75 images collected over 30 min

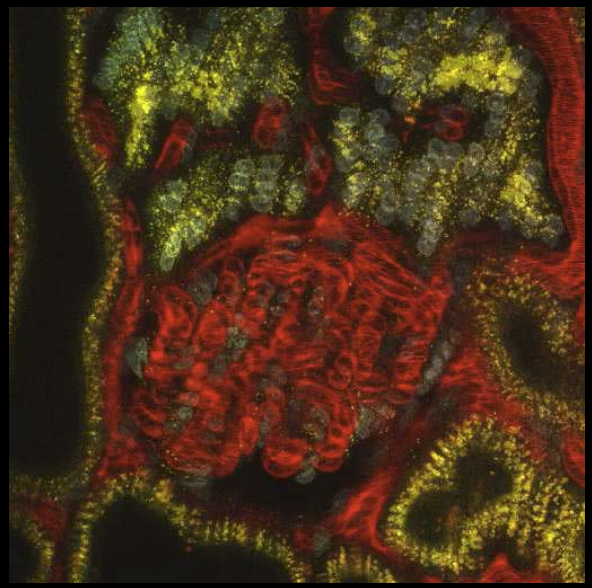




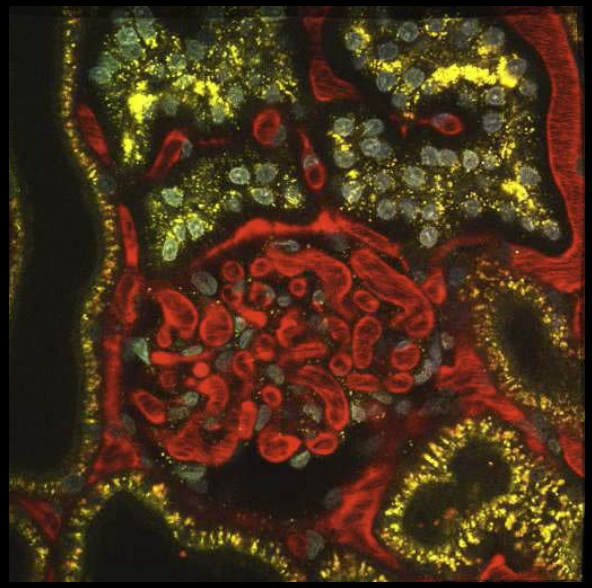


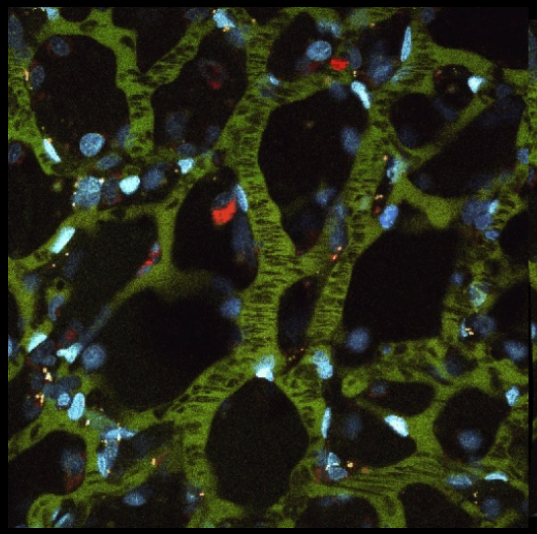
Reducing motion artifacts in intravital microscopy - Non-rigid image registration

Before

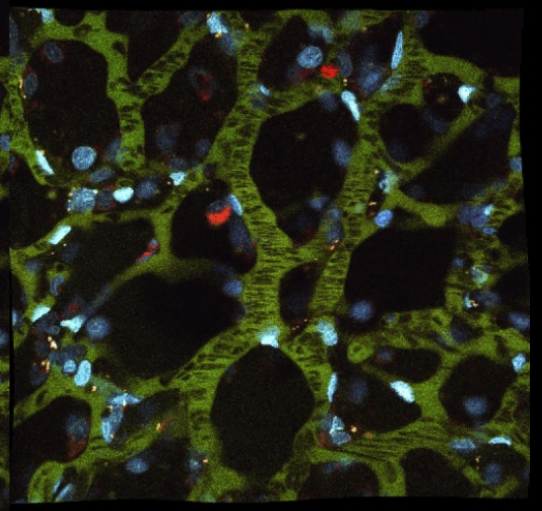


After





Before



After

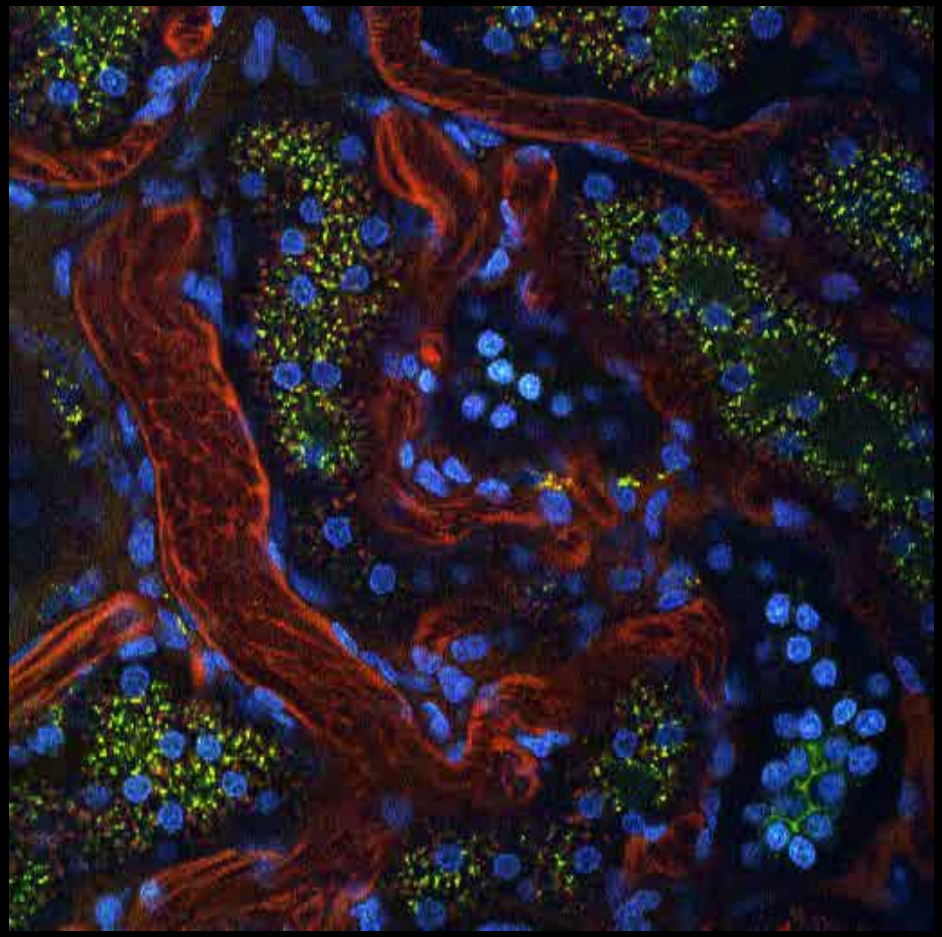
Presson, R., Brown, M., Sandoval, R., Dunn, K., Lorenz, K., Delp, E., Salama, P., Molitoris, B. and Petrache, I. Two-photon imaging within the murine thorax without respiratory and cardiac motion artifact. Am. J. Path. In press

Lorenz, K., Salama, P., Dunn, K. and E. Delp. Digital correction of motion artifacts in microscopy image sequences collected from living animals using rigid and non-rigid registration. Submitted

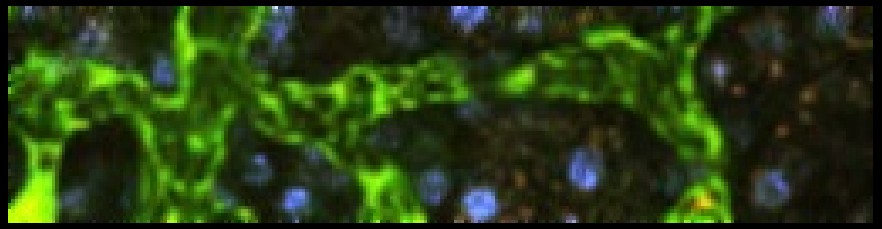
Challenges of intravital microscopy

- Motion artifacts from respiration and heartbeat
- Slow image capture rate

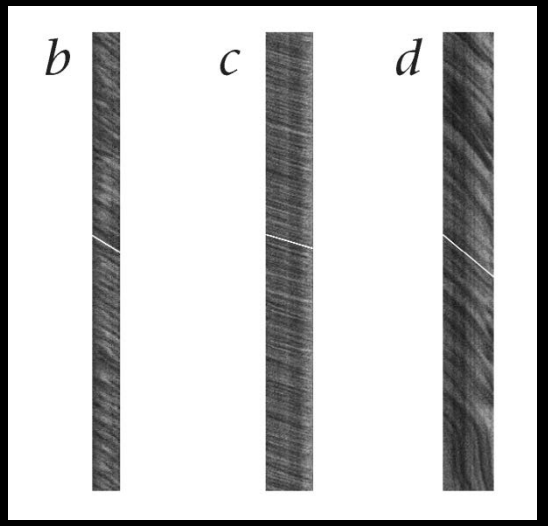
Single point scanning with a galvonometer
One 512 by 512 frame per second



512 lines at 1 frame per second



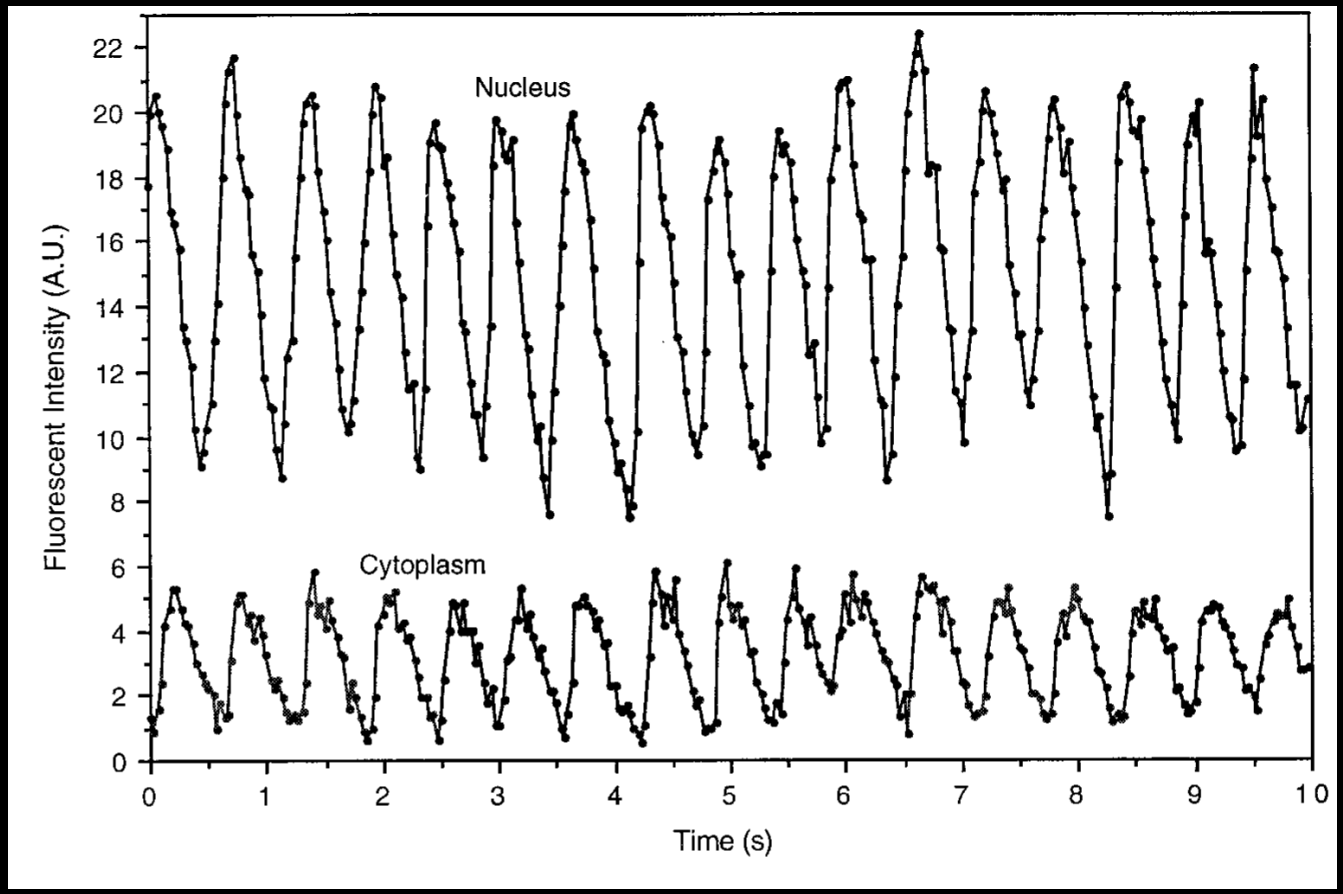
128 lines at 4 frames per second



(Brown et al., 2001)

1 line at 512 lines per second

Single point scanning with a resonant scanner
Calcium transients measured at 30 fps



Fan et al., 1999. Biophys. Journal 76:2412

Speed limits for multi-photon microscopy

30-fold higher frame rates require 30-fold shorter pixel dwell times which then requires 30 fold better signal. For most applications increasing the signal requires increasing the excitation.

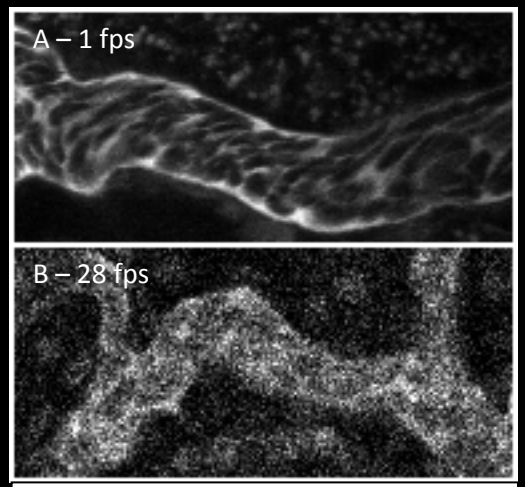


Figure 1 – Image of renal capillary of a living rat collected at 1 frame per second (A) or 28 frames per second (B).

- In many applications of multiphoton microscopy, power is already limiting – you may not be able to extract more signal with more power - 30 fold increase in signal requires 5.5 fold more power

- In many applications, multiphoton imaging is done at power levels very close to fluorophore saturation – you may not get proportional increases in signal

- Photobleaching increases at greater than the cube of power – disproportionately more photobleaching than signal with more power

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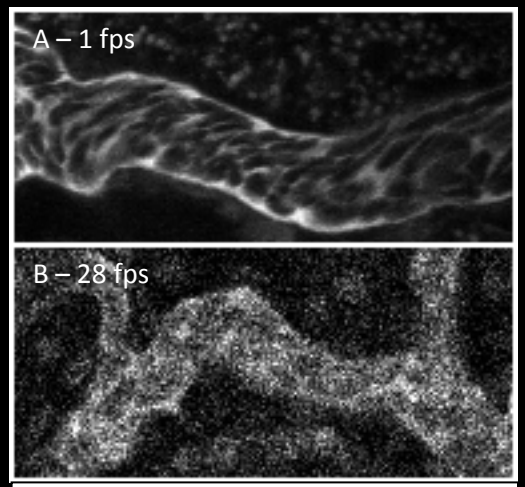

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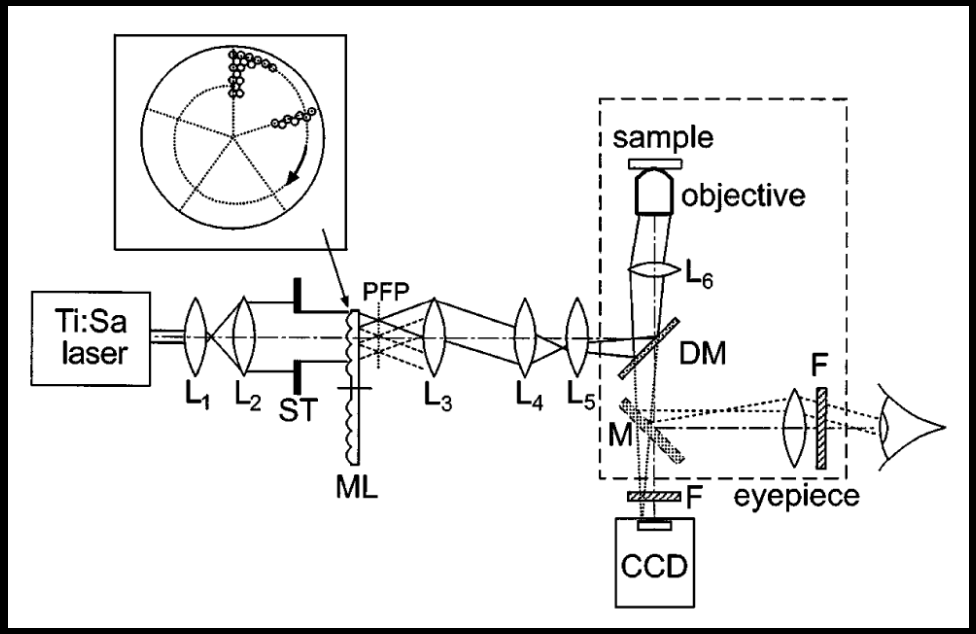
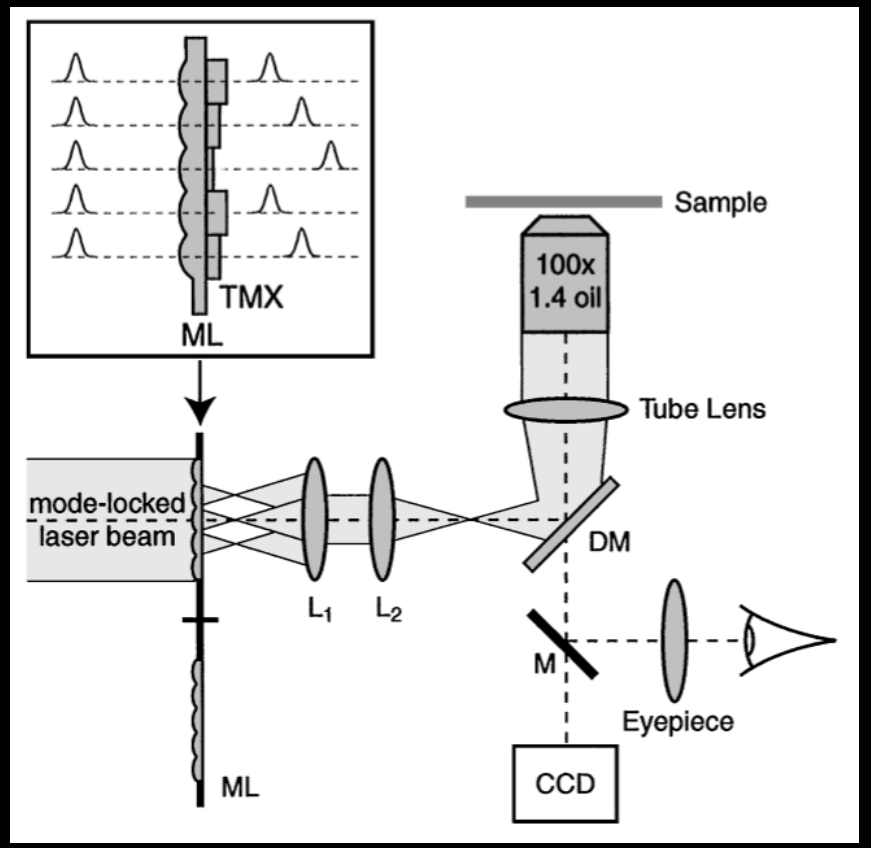
BUT, FOR SYSTEMS WITH SUFFICIENT SIGNAL, RESONANT SCANNERS ARE CAPABLE OF QUANTIFYING DYNAMICS IN VIVO -

TRY THE LEICA MULTIPHOTON SYSTEM

Multi-point scanning multiphoton microscopy



Bewensdorf et al., 1998. Optics Letters 23:655



Andresen et al., 2001. Optics Letters 26:75

Andresen system incorporates temporal delay between adjacent foci,
avoiding overlapping excitation in out-of-focus planes

Speed limits for multi-photon microscopy

- Multi-focal systems require that power be divided among multiple excitation points - again, this presumes sufficient laser power

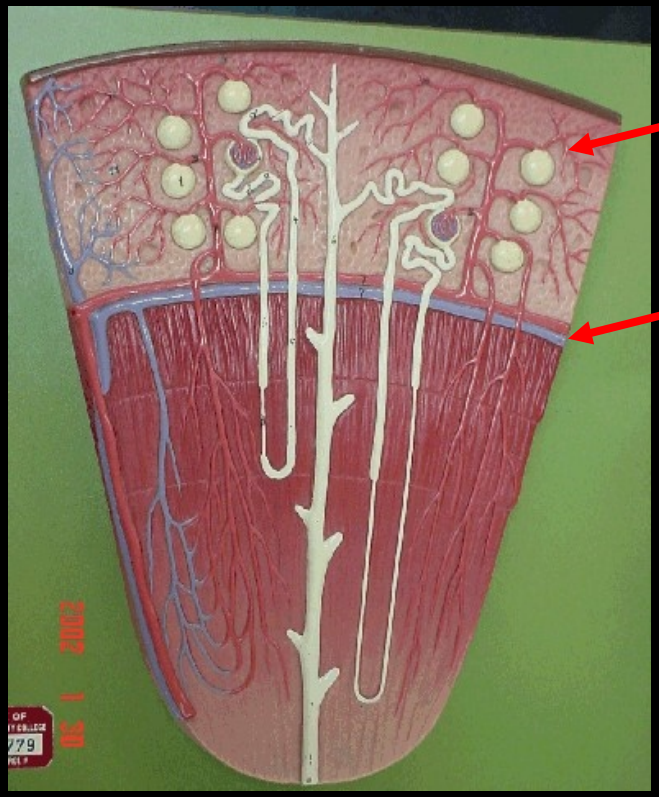
- Imaging onto a CCD results in scattered emissions being attributed to the wrong position in the image, increasing background and reducing resolution

- The system would thus be of limited utility for imaging deep into tissues, the conditions that justify multiphoton microscopy in the first place

Challenges of intravital microscopy

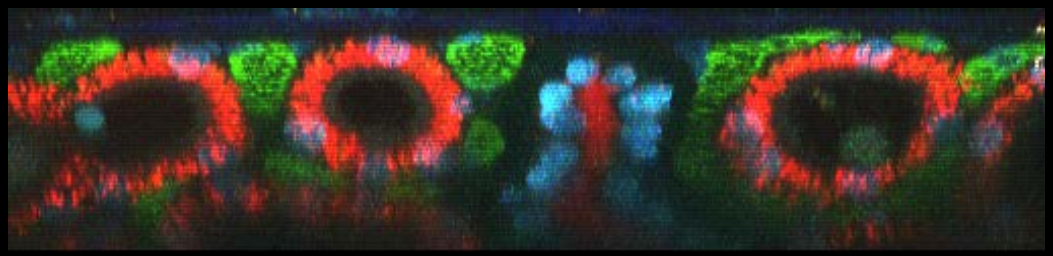
- Motion artifacts from respiration and heartbeat
- Slow image capture rate
- Limited depth at which useful images can be collected

Multiphoton microscopy
Attenuation of signal with depth in the kidney



Glomeruli of most rats, mice are around 400 microns from the surface of the kidney

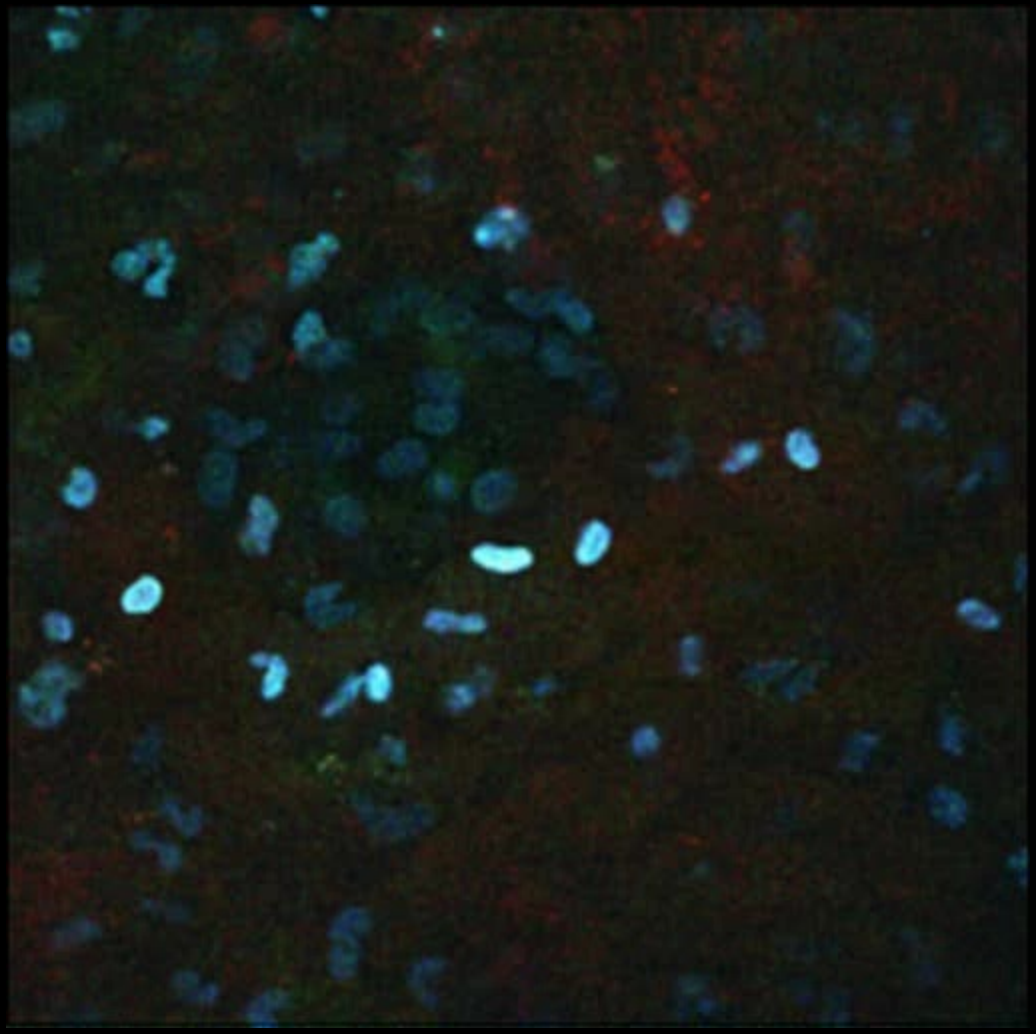
Cortico-medullary boundary is around 2 mm from the surface of the kidney



70 microns

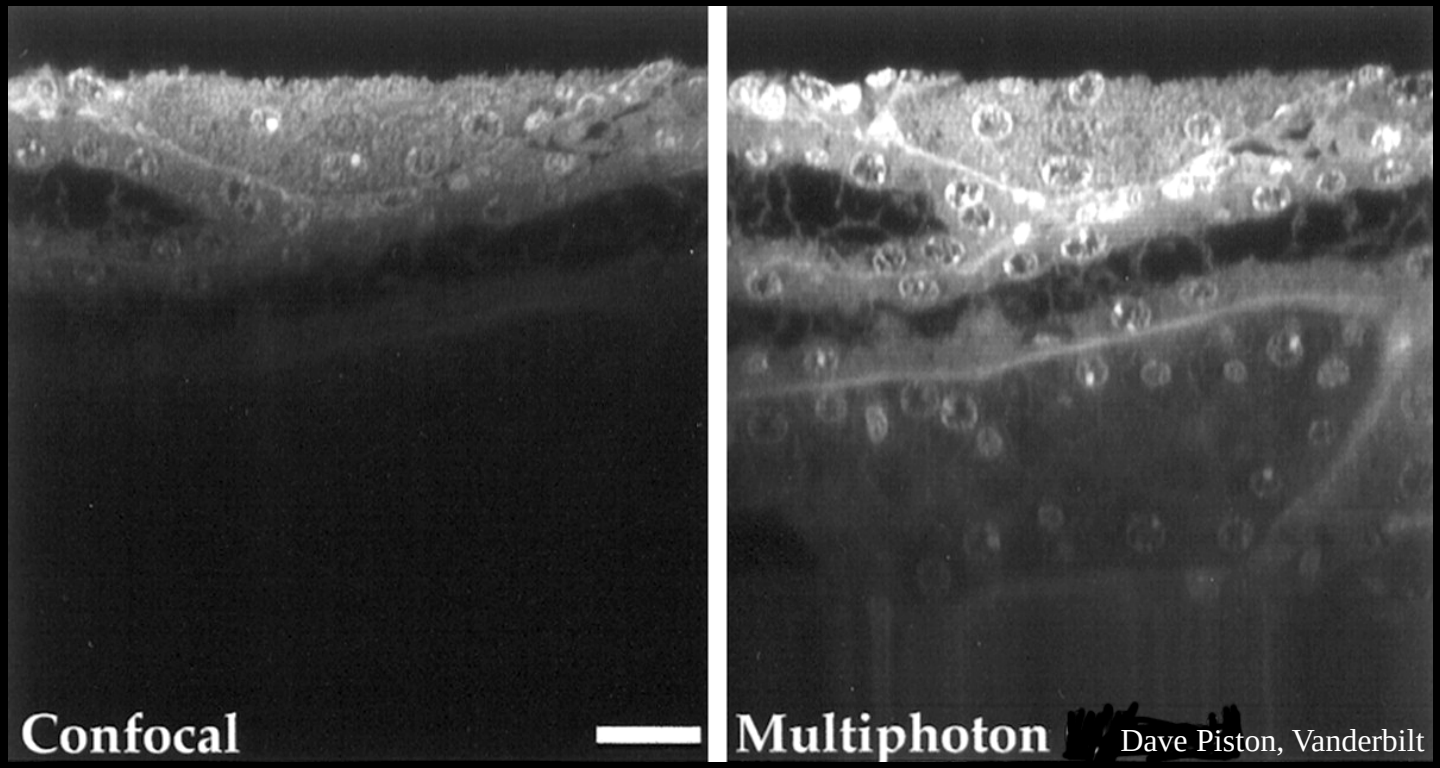
With a 60X NA 1.2 water immersion objective, signal attenuation prevents imaging deeper than around 100 microns in kidney of living rats

Multiphoton microscopy
Attenuation of signal with depth in the kidney



Ruben Sandoval

Multiphoton microscopy
Sources of signal attenuation of signal with depth



- Light absorption
- Light scattering
- Spherical aberration

Multiphoton microscopy
Light absorption as a source of signal attenuation with depth

Model light extinction as - $I_z = I_0 e^{-az}$

Where

I_z = intensity of light at depth z

a = the absorption coefficient

For fluorescence excitation

Near infrared light, a = .05 to 2 cm^{-1} in biological tissues

So transmission attenuated by 50% in 0.35 - 14 cm

But excitation is proportional to I^2,

Excitation is attenuated by 75% in 0.35 - 14 cm

- Excitation reduced by 75% in ~ 3500 to 140,000 microns - not a big deal

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For collection of fluorescence emissions

For 550 nm light, a = 4 to 20 cm^{-1} in biological tissues

so transmission attenuated to 50% in 350-1730 microns

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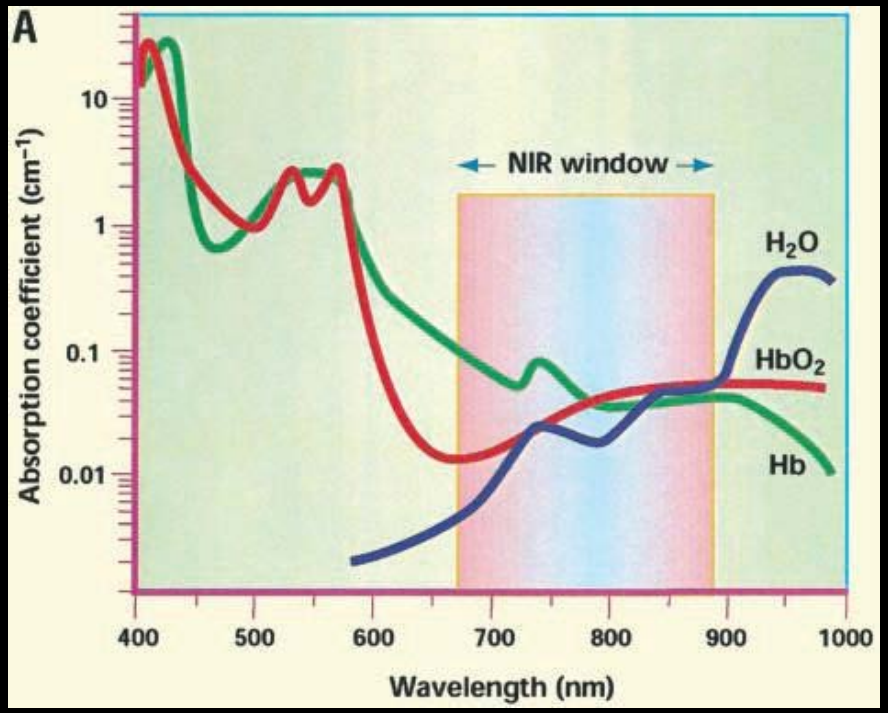
Most investigators agree, absorption is seldom a significant issue in biological tissues, except in skin (melanin) and under blood vessels (hemoglobin)

Multiphoton microscopy
Absorption of light by blood (hemoglobin)



For fluorescence excitation

a = 4 cm^{-1} at 780, - excitation attenuated by 75% at 1730 microns

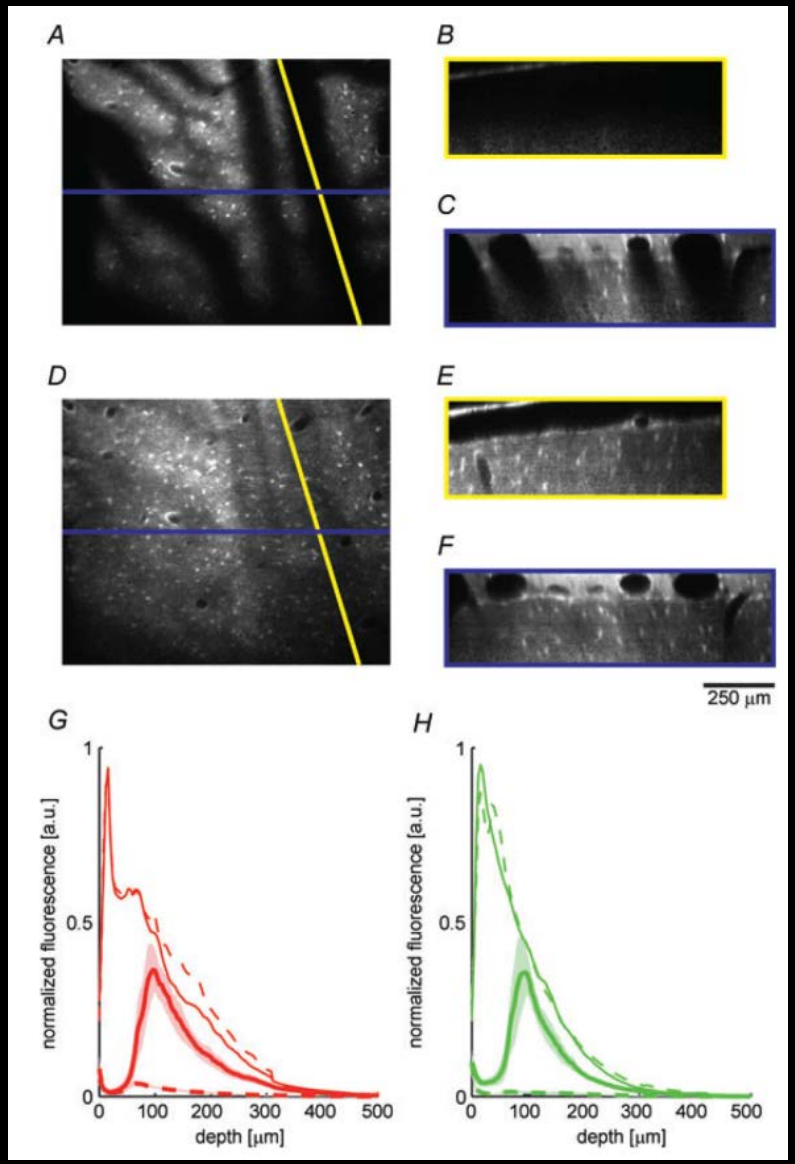
would lose < 1% over 10 micron capillary

For collection of fluorescence emissions

a = 250 cm^{-1} at 550, transmission attenuated to 50% at 28 microns

would lose ~20% under a 10 micron capillary

Multiphoton microscopy
Absorption of light by blood (hemoglobin)



J Physiol 587.13 (2009) pp 3153–3158

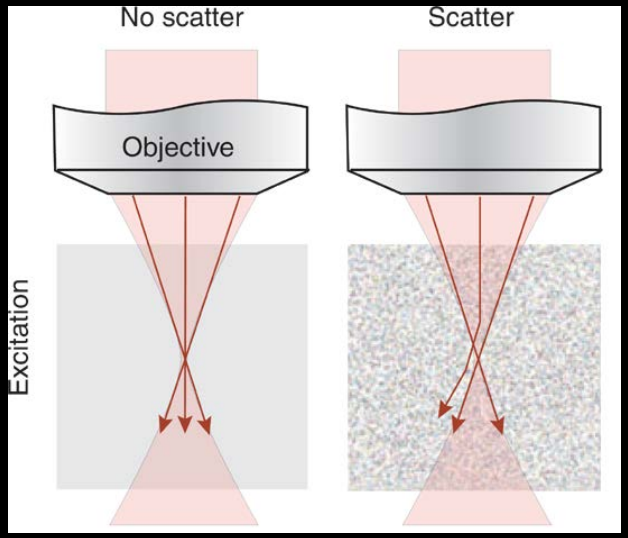
Improved *in vivo* two-photon imaging after blood replacement by perfluorocarbon

F. Haiss[1], R. Jolivet[1], M. T. Wyss[1], J. Reichold[2], N. B. Braham[3], F. Scheffold[3], M. P. Krafft[4] and B. Weber[1]

[1] *Institute of Pharmacology and Toxicology, University of Zurich, 8091 Zurich, Switzerland*
[2] *Institute of Fluid Dynamics, ETH Zurich, 8091 Zurich, Switzerland*
[3] *Department of Physics and Fribourg Center for Nanomaterials, University of Fribourg, 1700 Fribourg, Switzerland*
[4] *Université de Strasbourg, Institut Charles Sadron (CNRS), 23 rue du Loess, 67034 Strasbourg Cedex, France*

Figure 2. Structural imaging of cortical astrocytes *in vivo* before and after PFC transfusion
A, two-photon image of astrocytes (SR101 staining) taken at a depth of 150 μm before PFC transfusion. Shadows resulting from absorption and scattering by red blood cells disappear after PFC transfusion (*D*). *B*, *C*, *E* and *F*, *Z*-sections reconstructed from image stacks were taken at the location marked with the blue and yellow line in *A* and *D*. Astrocytes located below large vessels were not visible before PFC transfusion. Contrast has been optimized in each individual frame in the stack. *G*, depth penetration in the red channel was measured at fixed laser intensity before (dashed line) and after the transfusion (continuous line) below a big surface vessel about 50 μm thick (thick lines) and in an open area without vessel at the surface (thin lines). All traces are averages over 4 stacks collected in 3 animals (± S.E.M. shaded area; average region of interest (ROI) is 800 pixels). All curves were normalized to the peak surface intensity measured in the open area before and after transfusion. The transfusion does not significantly change depth penetration in the open area (compare thin lines) but dramatically increases depth penetration below vessels (thick lines). *H*, same as in *G* for the green channel.

Multiphoton microscopy
Light scattering as a source of signal attenuation with depth



Model light extinction as - $I_z = I_0 e^{-sz}$

Where

I_z = intensity of light at depth z

s = the scattering coefficient

For fluorescence excitation

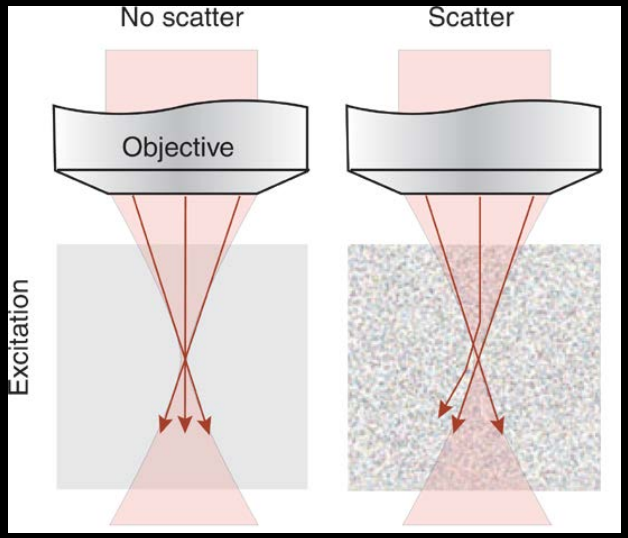
Near infrared light, s = 5 - 100 cm^{-1} in biological tissues

So transmission attenuated by 50% in 70 to 1400 microns

But excitation is proportional to I^2,

Excitation attenuated by 75% in 70 to 1400 microns

Multiphoton microscopy
Light scattering as a source of signal attenuation with depth



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Near infrared light, s = 5 - 100 cm^{-1} in biological tissues

So transmission attenuated by 50% in 70 to 1400 microns

But excitation is proportional to I^2,

Excitation attenuated by 75% in 70 to 1400 microns

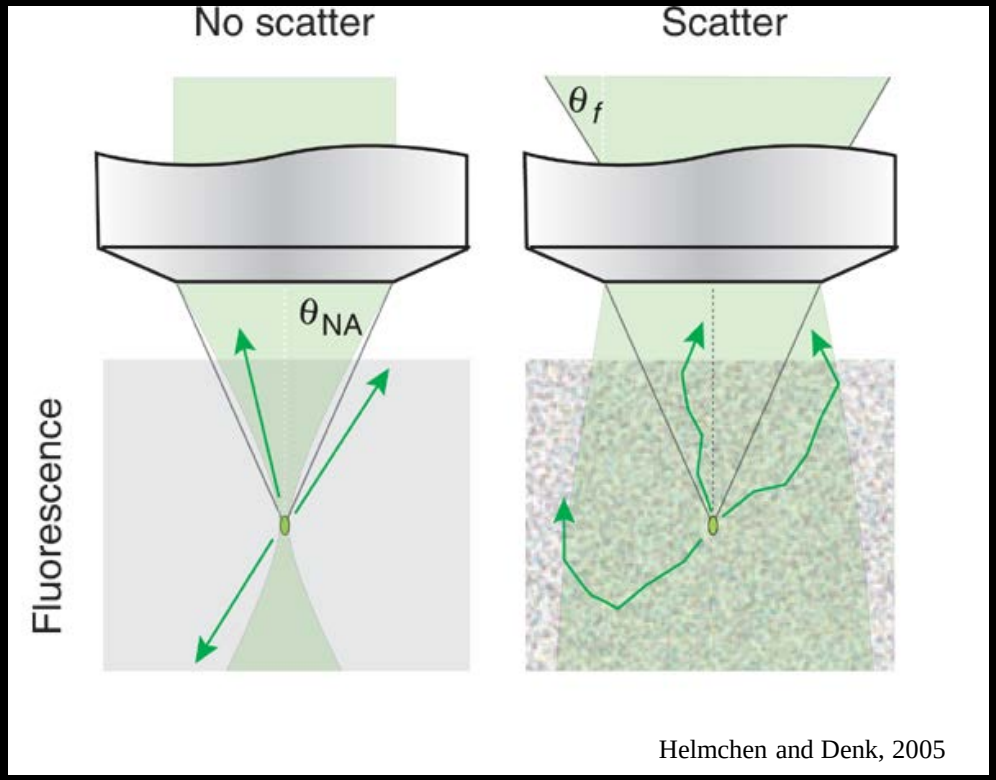
For collection of fluorescence emissions

For 550 nm light, s = 100 - 500 in biological tissues

So transmission attenuated by 50% in 15 - 70 microns

but even scattered emissions collected with non-descanned detectors

Multiphoton microscopy
Light scattering as a source of signal attenuation with depth



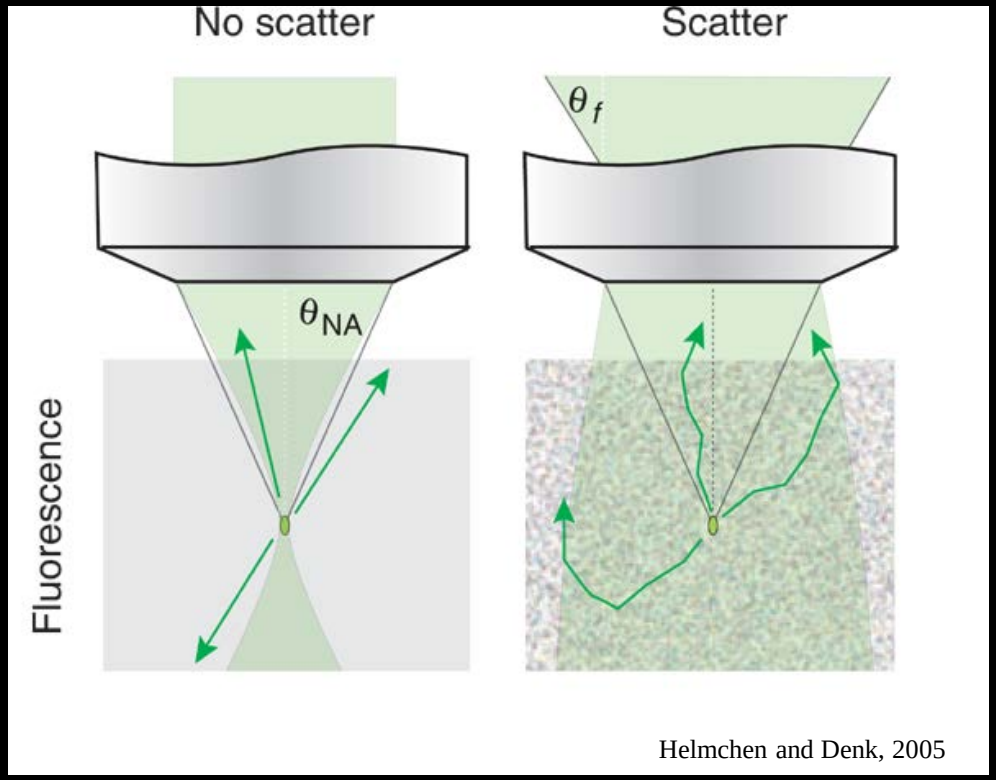
Helmchen and Denk, 2005

In most biological tissues there are effectively no ballistic emissions at 200 microns depth

Consequently, fluorescence arises from a diffuse region whose diameter is 1.5X the focal depth

Under these conditions, the collection angle is larger than the NA

Multiphoton microscopy
Light scattering as a source of signal attenuation with depth



Helmchen and Denk, 2005

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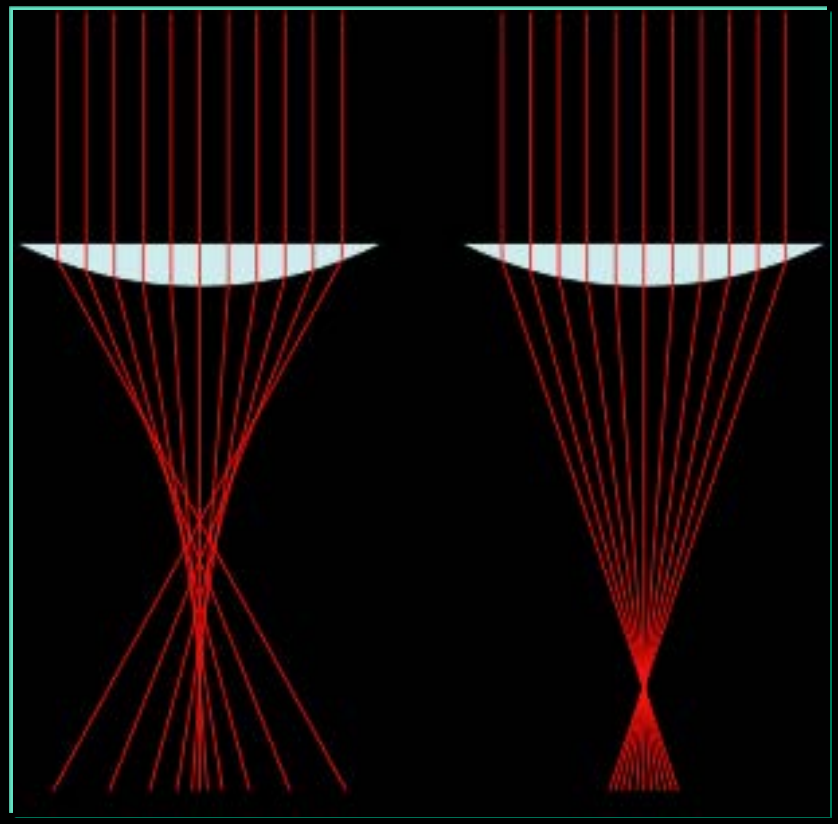
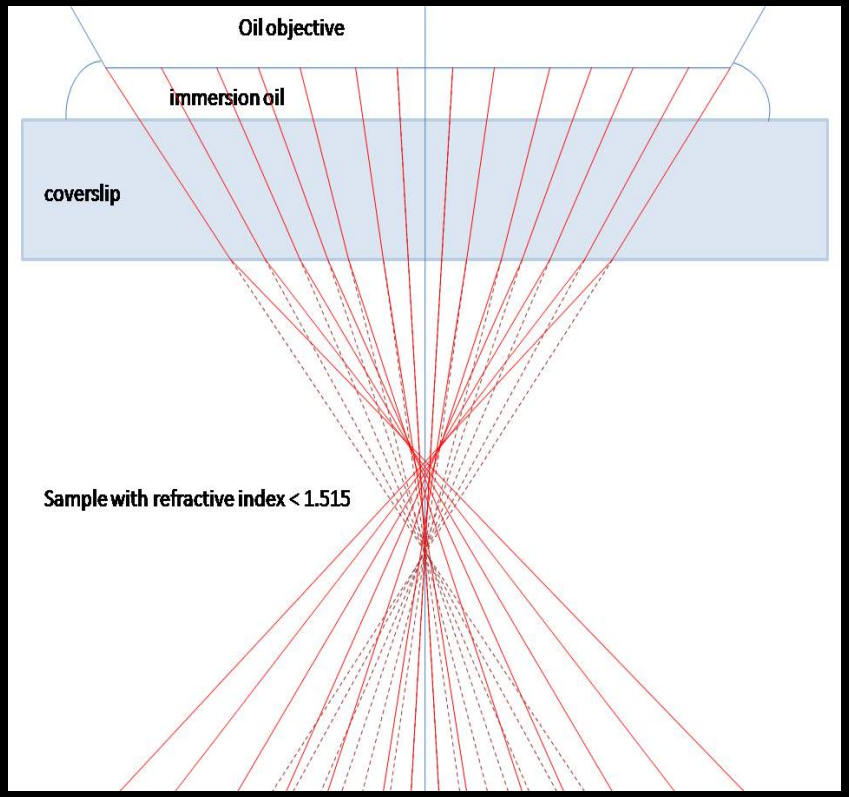
Under these conditions, the collection angle is larger than the NA

Alternatively – IR confocal microscopy –
Both excitation and emissions are in the IR range, minimizing scattering and absorption
Try the Olympus FV1200 IR confocal microscope

Multiphoton microscopy
Refractive index mismatch and spherical aberration



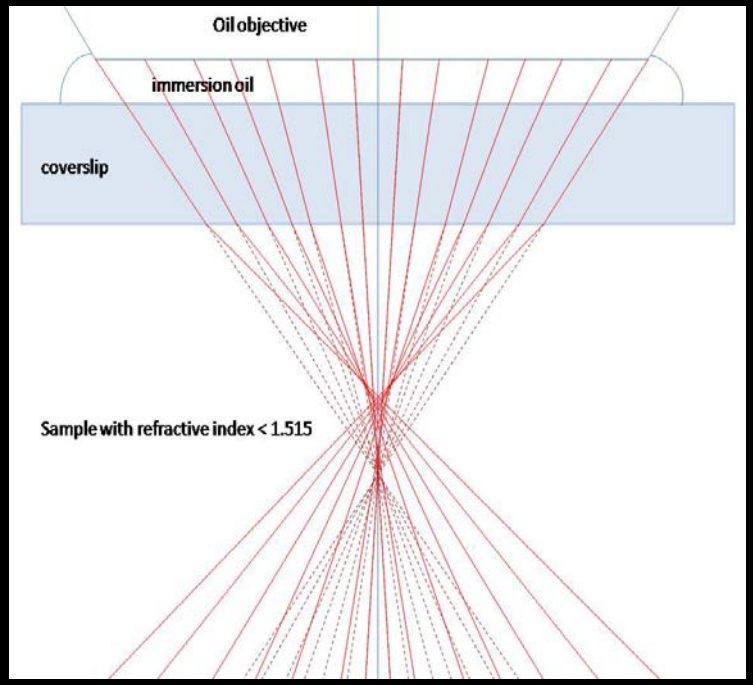


A lens with spherical aberration focuses axial rays and peripheral rays to different points. Spherical aberration commonly results from refractive index mismatch.

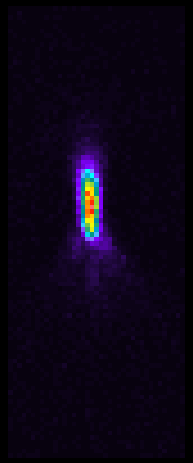
Since the average refractive index of the kidney is ~1.4, we incur spherical aberration with either water immersion or oil immersion objectives

Multiphoton microscopy
Spherical aberration and signal attenuation with depth

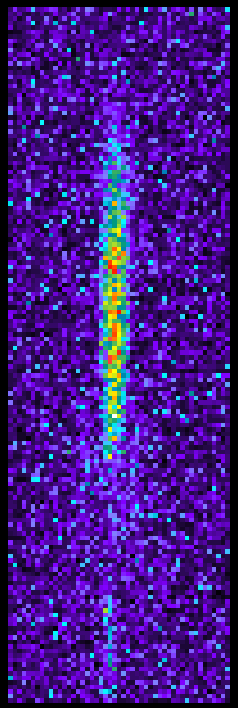
Two photon microscopy of fluorescent beads mounted in different media using an oil-immersion objective



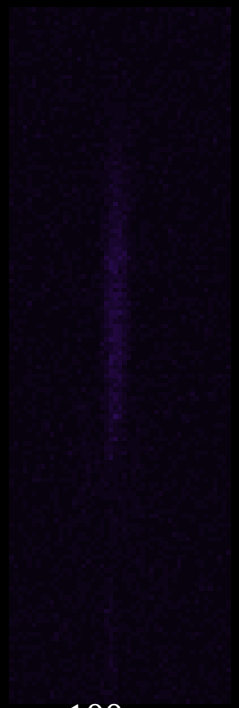
An oil-immersion objective imaging into an aqueous medium



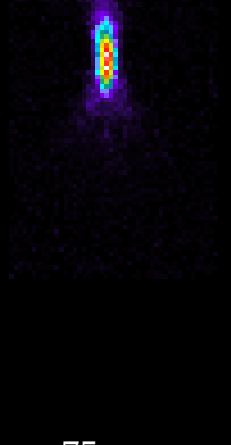
Surface Aqueous



100um Aqueous



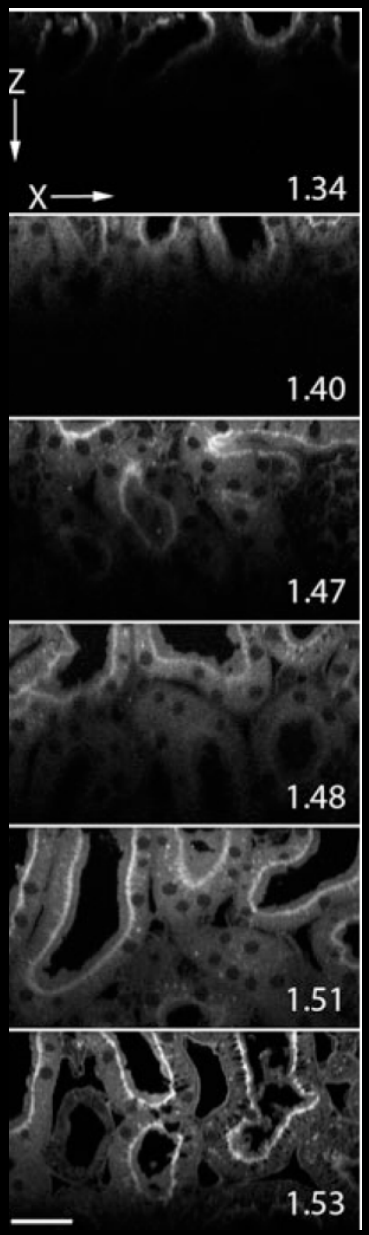
100um Aqueous (w/out stretch)



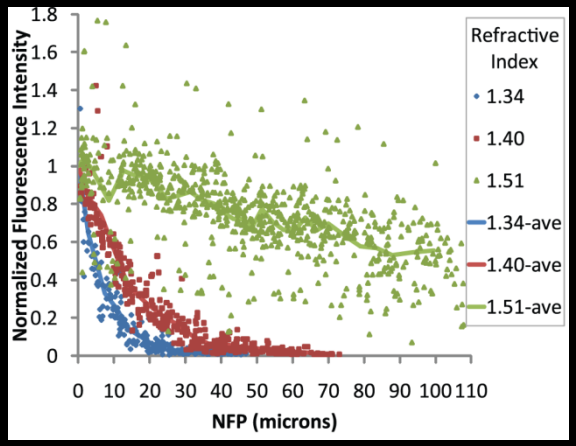
75um 1.5 RI medium

Since fluorescence is quadratically related to illumination, broadening of the focus by spherical aberration seriously decreases multi-photon fluorescence excitation

Predominant source of signal attenuation in kidney tissues?

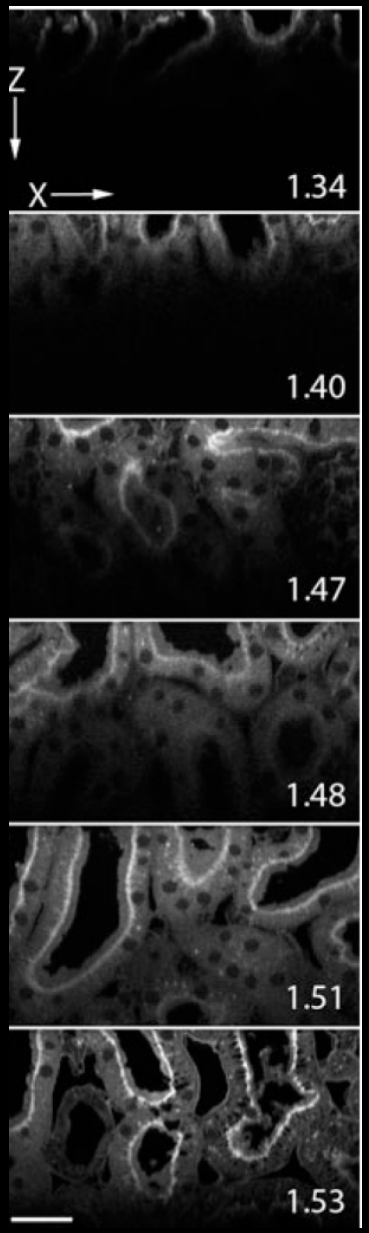


60X – Oil immersion

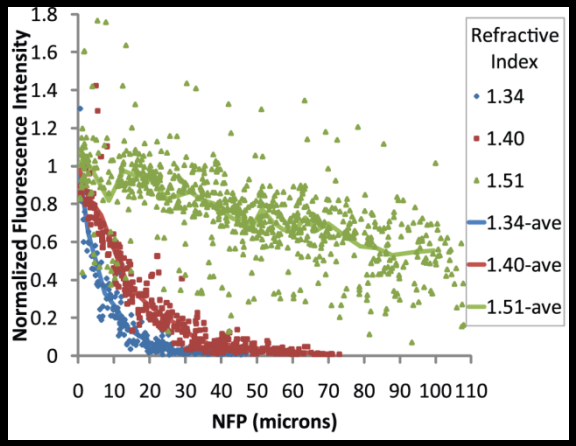


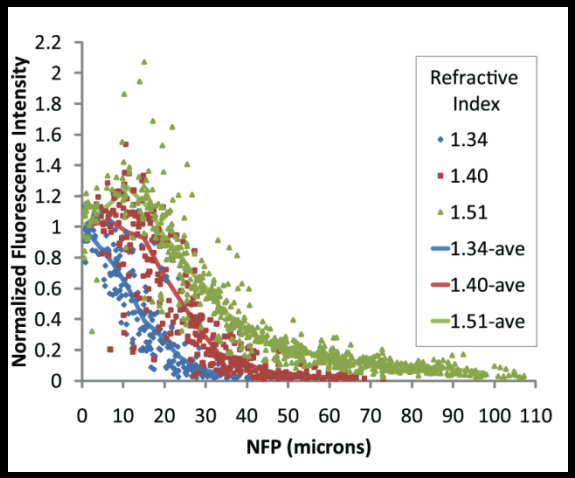
Young, Clendenon, Byars and Dunn. 2011. The effects of refractive index heterogeneity within kidney tissue on multiphoton fluorescence excitation microscopy. J. Microscopy.

Predominant source of signal attenuation in kidney tissues?

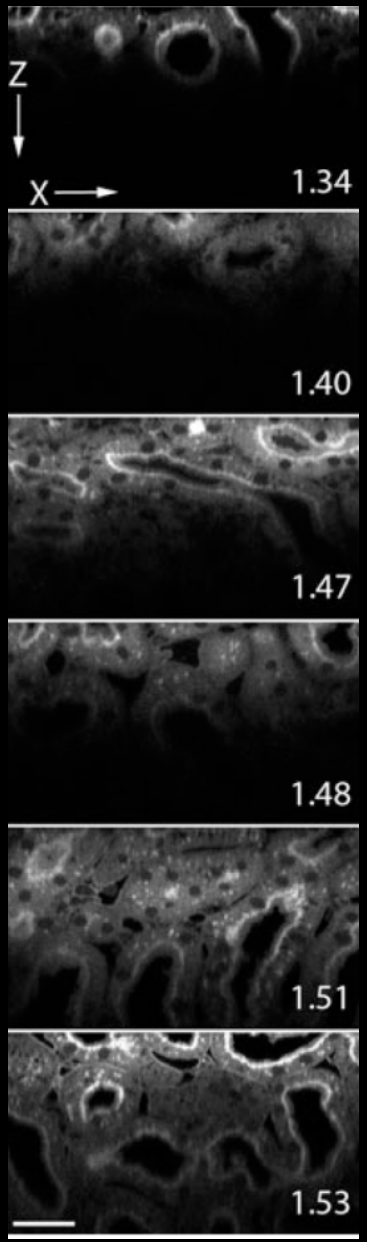


60X – Oil immersion





Young, Clendenon, Byars and Dunn. 2011. The effects of refractive index heterogeneity within kidney tissue on multiphoton fluorescence excitation microscopy. J. Microscopy.

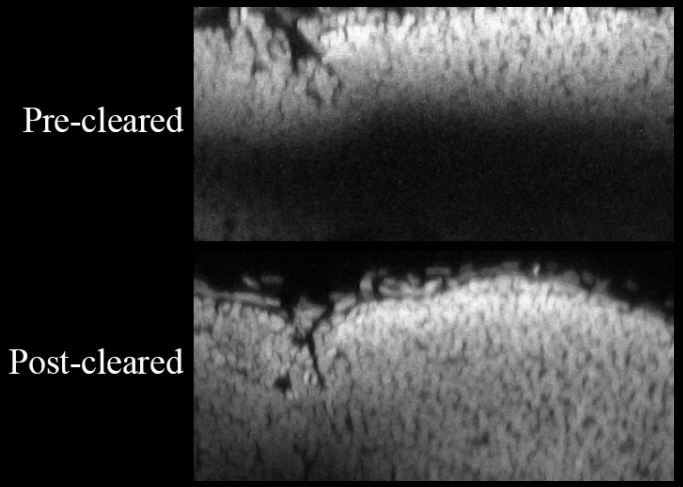


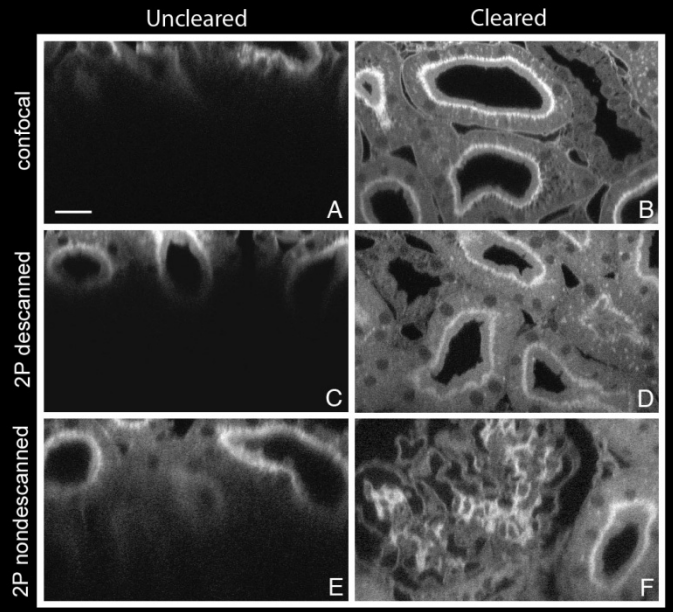
60X – Water immersion

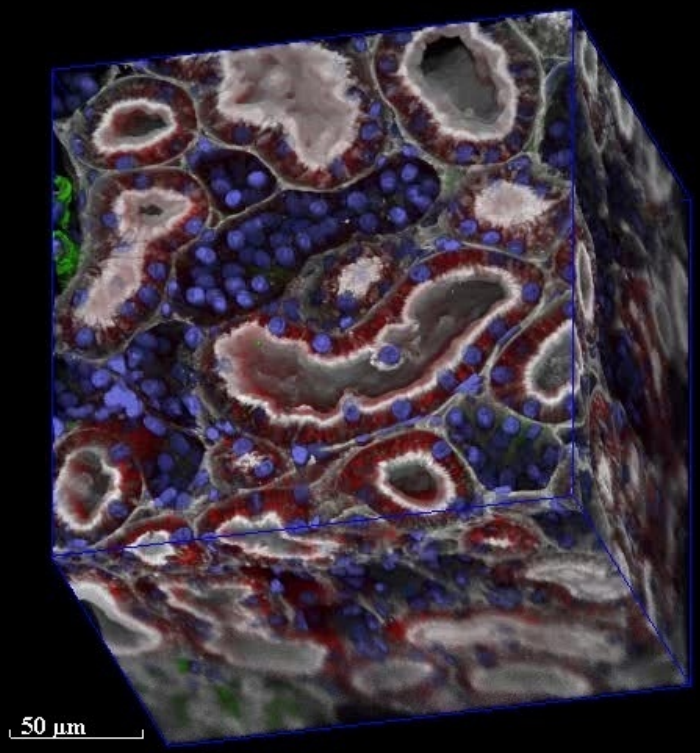
Limited optical reach in the living kidney
Potential solutions

- Reduce scattering – give up and fix the kidney

Tissue clearing - Elimination of refractive index mismatch and scattering

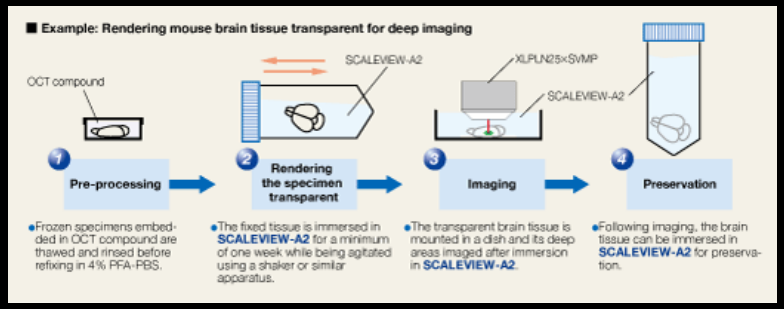


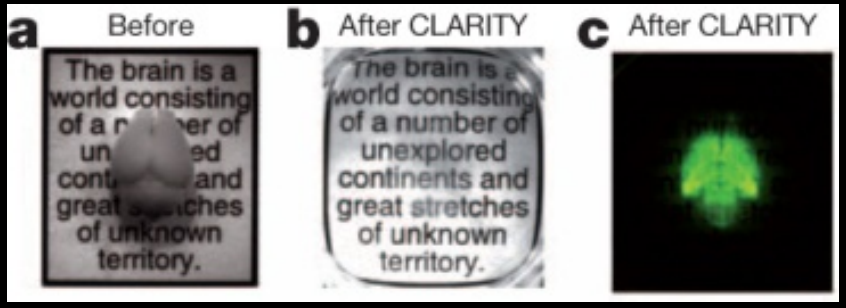




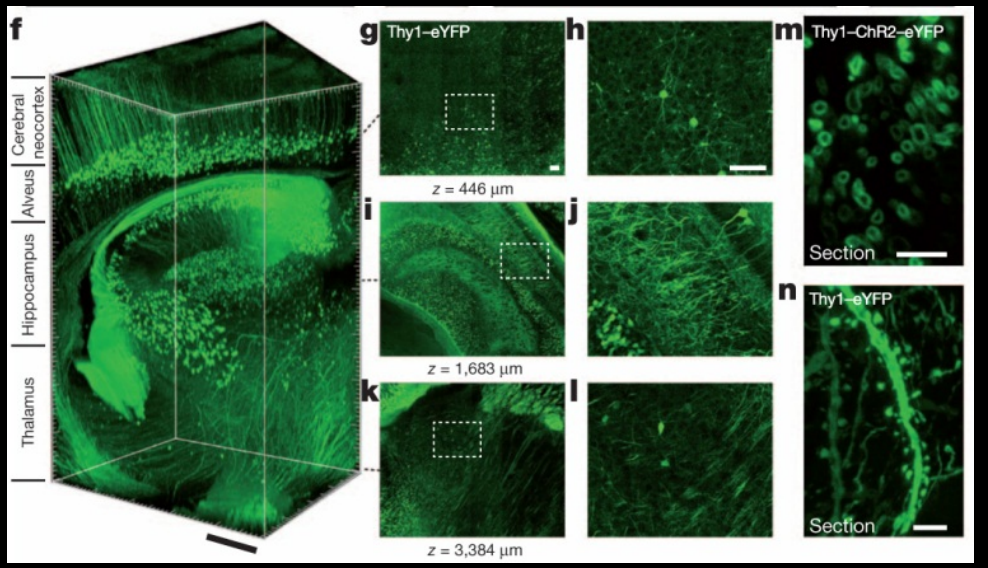
Clendenon, Ferkowicz, Young, and Dunn. 2011. Deep Tissue Fluorescent Imaging in Scattering Specimens Using Confocal Microscopy. Microscopy and Microanalysis.

Tissue clearing and refractive index matching - Olympus Scaleview, Clarity





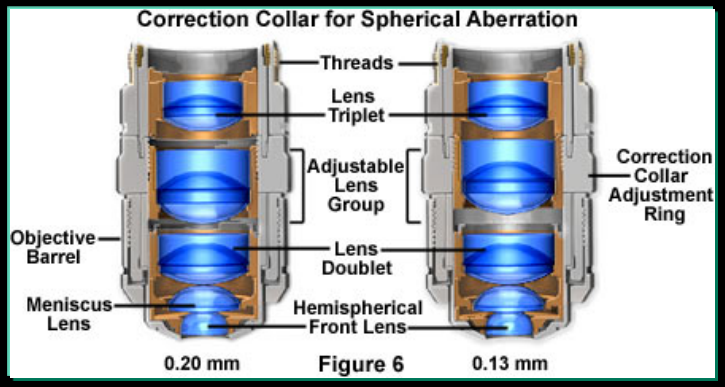
512 by 512 by 512 microns
(water immersion objective)



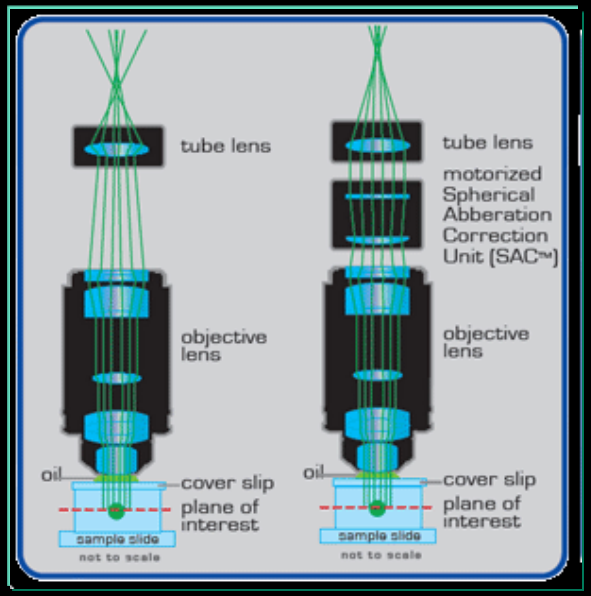
Clarity – Chung et al., 2013. Structural and molecular interrogation of intact biological systems, Nature

Limited optical reach in the living kidney
Potential solutions

- Reduce scattering (change research)
- Reduce optical aberrations

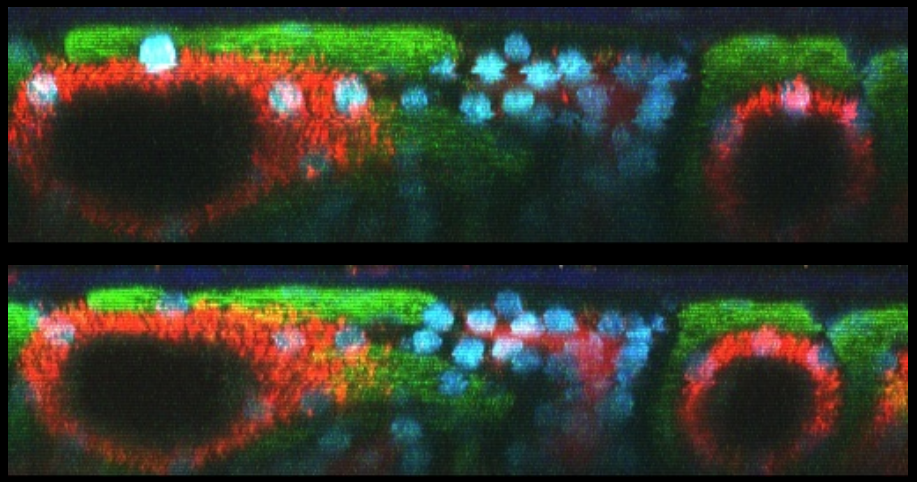


Objective lens correction collars



Spherical aberration correction lenses

Minimizing spherical aberration in living kidney – oil or water immersion objectives?

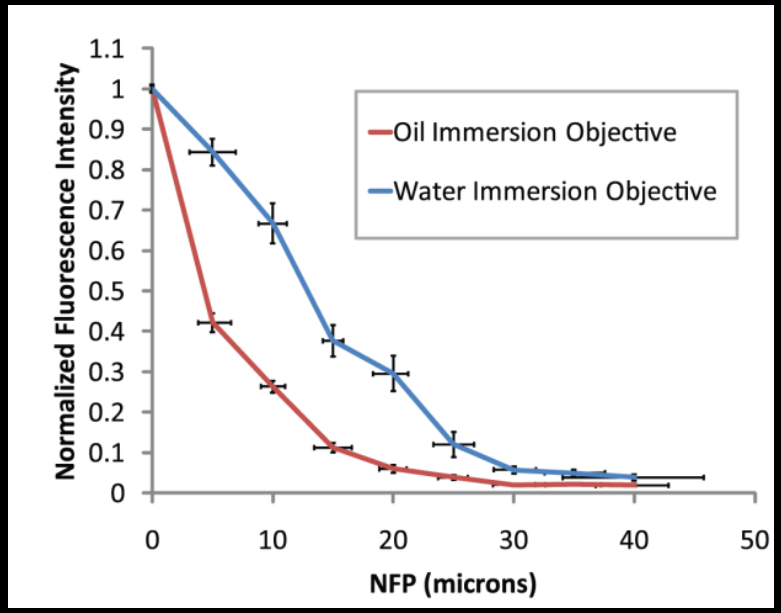


60X – Oil immersion

60X – Water immersion

Ruben Sandoval

Fluorescent beads in fixed kidney mounted in PBS

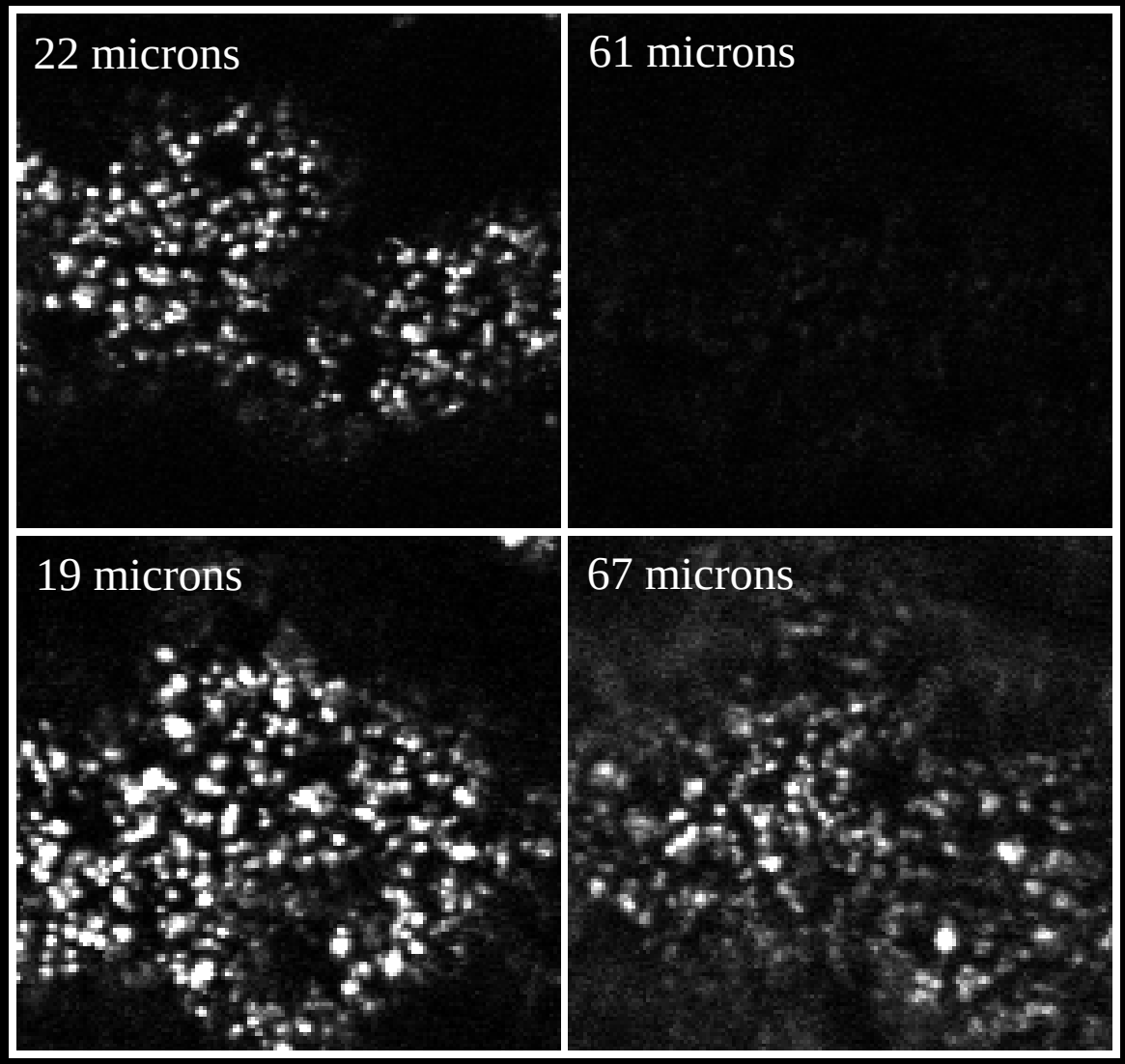


Young, Clendenon, Byars, Decca and Dunn. 2011. J. Microscopy

Minimizing spherical aberration in living kidney – Olympus silicone oil immersion objective

Olympus 25X,
NA 1.05
Water immersion
objective
RI 1.3

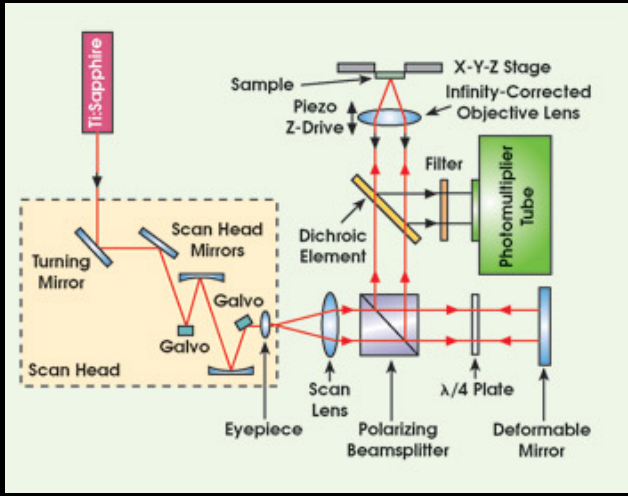
Olympus 30X,
NA 1.05
Silicone oil
immersion
objective
RI 1.4

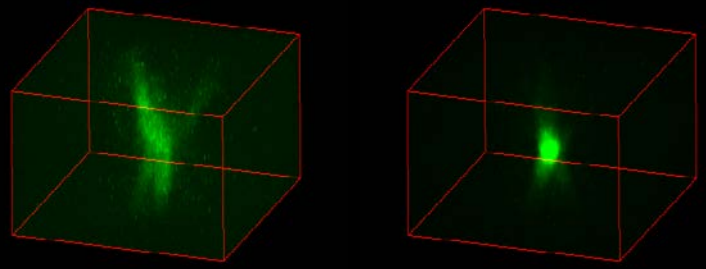


Limited optical reach in the living kidney
Potential solutions

- Reduce scattering (change research)
- Reduce optical aberrations

Adaptive optics



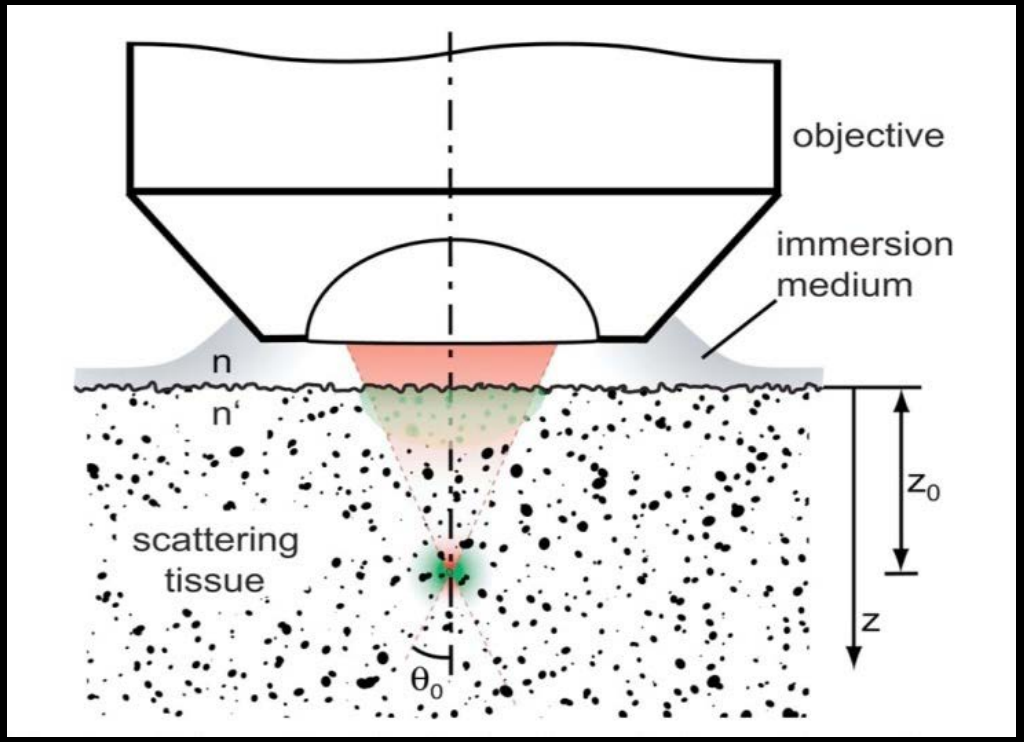


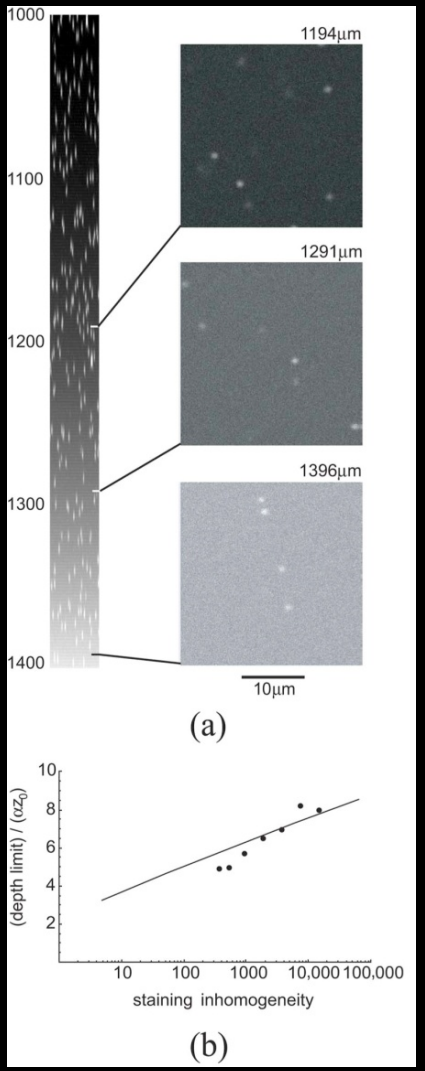
Confocal microscope images of 1 micron fluorescent spheres located below 100 microns of kidney tissue before (A) and after (B) adaptive optics correction – Joel Kubby, UCSC

Limited optical reach in the living kidney
Potential solutions

- Reduce scattering (change research)
- Reduce optical aberrations
- Jack up the laser? Up to a point . . .

Fundamental imaging depth limit in two photon microscopy

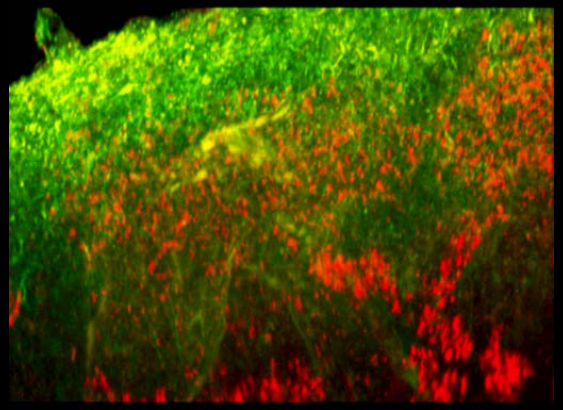


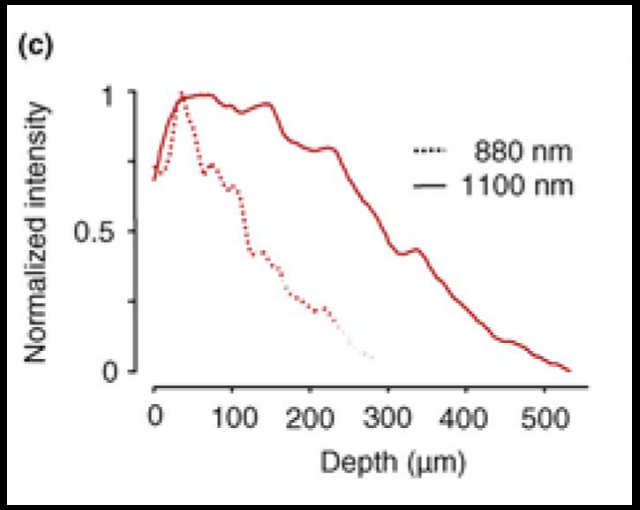


Imaging depth increases with labeling inhomogeneity, with decreasing pulse-width and with decreasing scattering anisotropy, but is ultimately limited by near-surface fluorescence - Theer and Denk, 2006

Limited optical reach in the living kidney
Potential solutions

- Reduce scattering (change research)
- Reduce optical aberrations
- Jack up the laser? Up to a point . . .
- Longer wavelengths of light

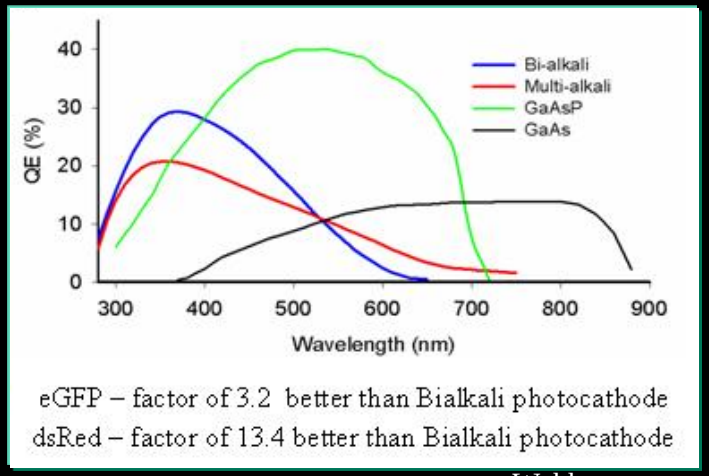




Andresen et al., 2009. Infrared multiphoton microscopy: subcellular-resolved deep tissue imaging. Curr. Opin. Biotech.

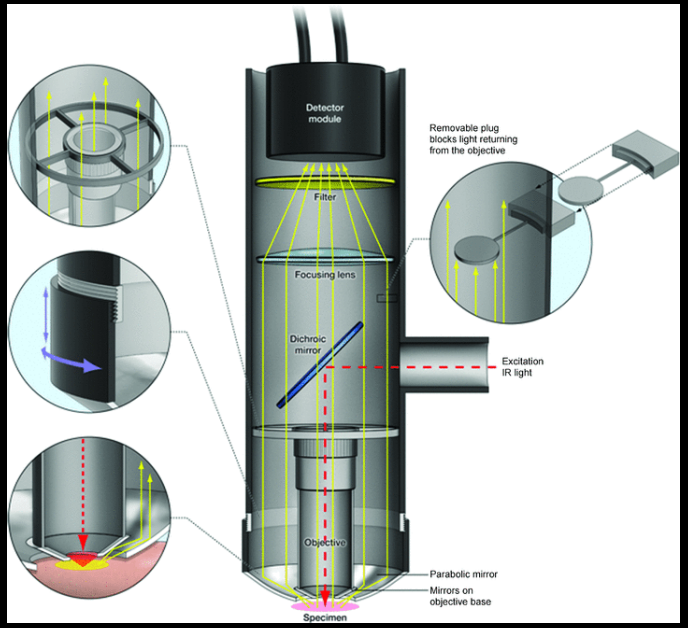
Limited optical reach in the living kidney
Potential solutions

- Reduce scattering (change research)
- Reduce optical aberrations
- Jack up the laser? Up to a point . . .
- Longer wavelengths of light
- Improve fluorescence collection



eGFP – factor of 3.2 better than Bialkali photocathode
dsRed – factor of 13.4 better than Bialkali photocathode

Webb group



COMBS, SMIRNOV, CHESS, MCGAVERN, SCHROEDER, RILEY, KANG, LUGAR-HAMMER, GANDJBAKHCHE, KNUTSON, and BALABAN (2011), Journal of Microscopy

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Potential solutions

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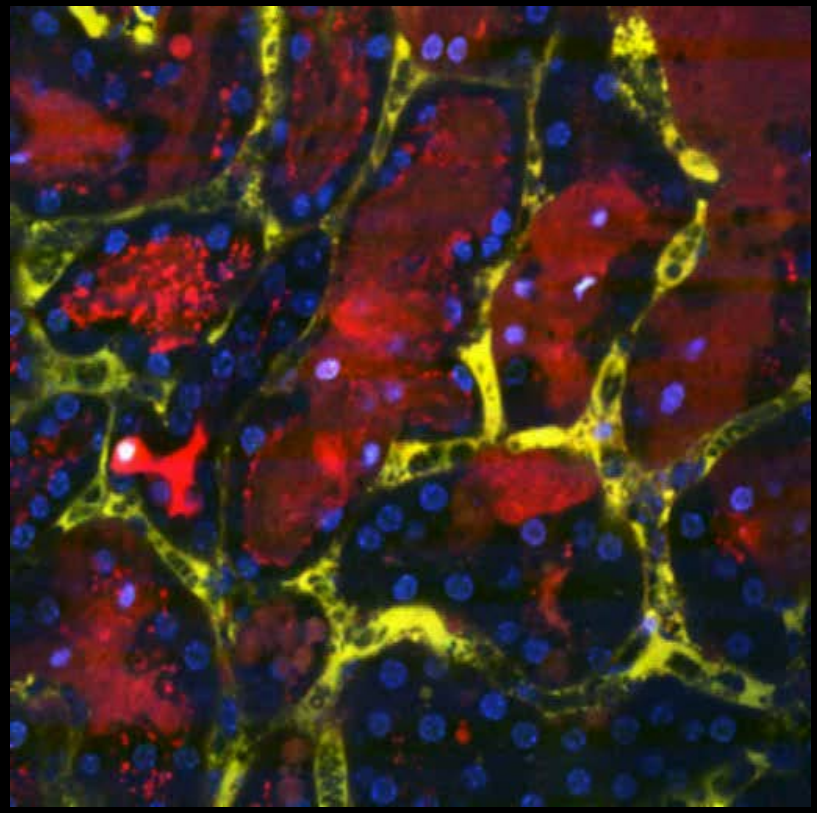
- Jack up the laser? Up to a point . . .
- Longer wavelengths of light
- Improve fluorescence collection
- Invade the kidney - "cut down", GRIN lens, micro-prisms



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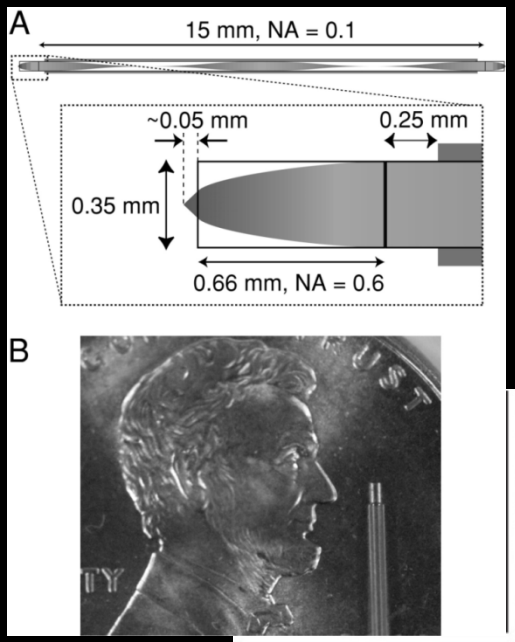

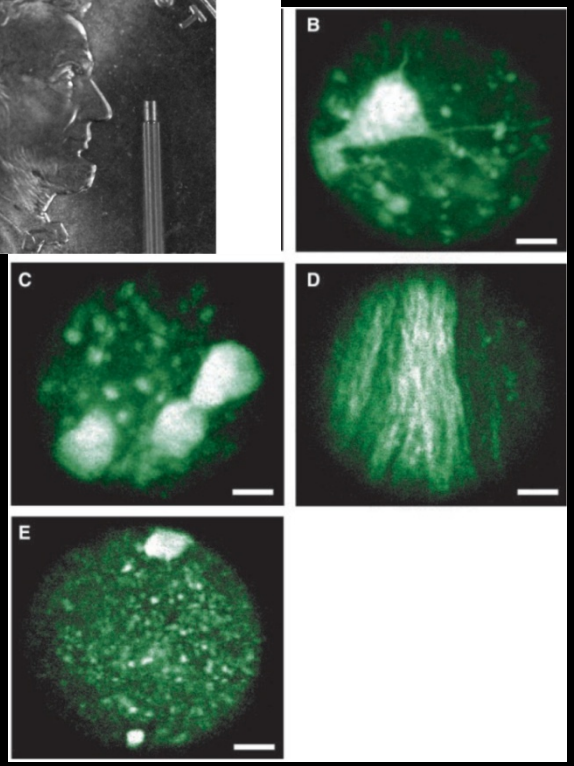

Levene et al., 2004, J. Neurophysiol

Limited optical reach in the living kidney
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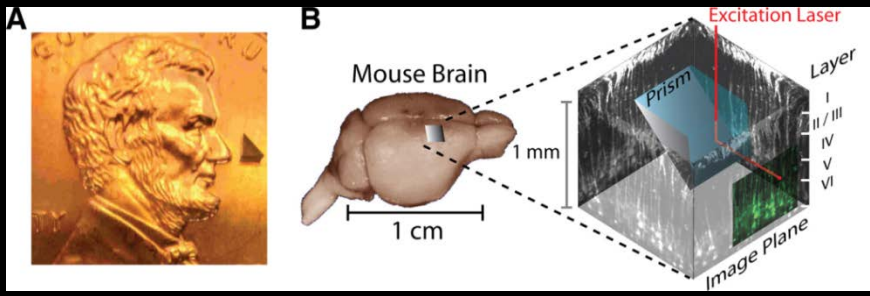

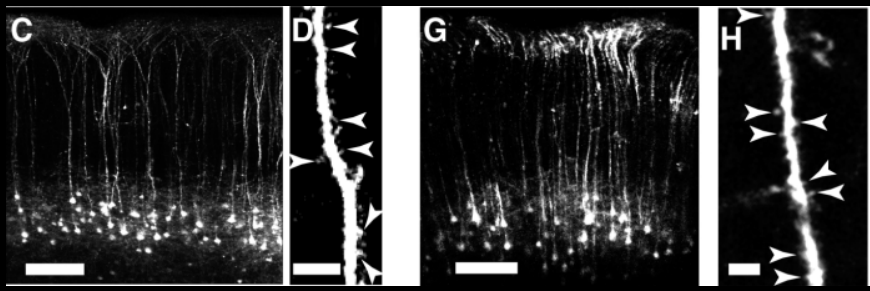

Conventional XY image in brain slice

XZ image in vivo with prism

Chia and Levene. 2009. Microprisms for in vivo multilayer cortical imaging. J. Neurophysio.

Limited optical reach in the living kidney
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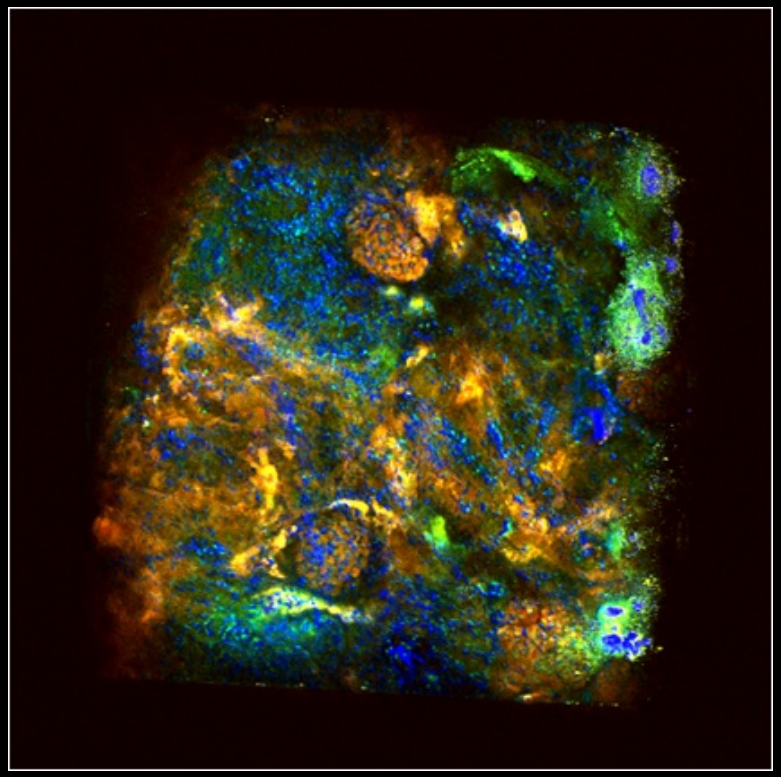
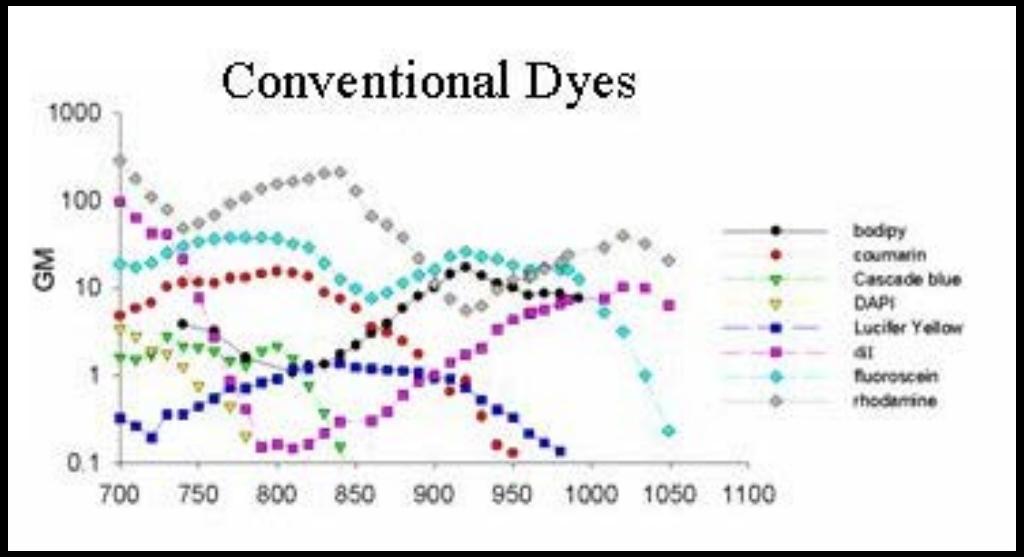


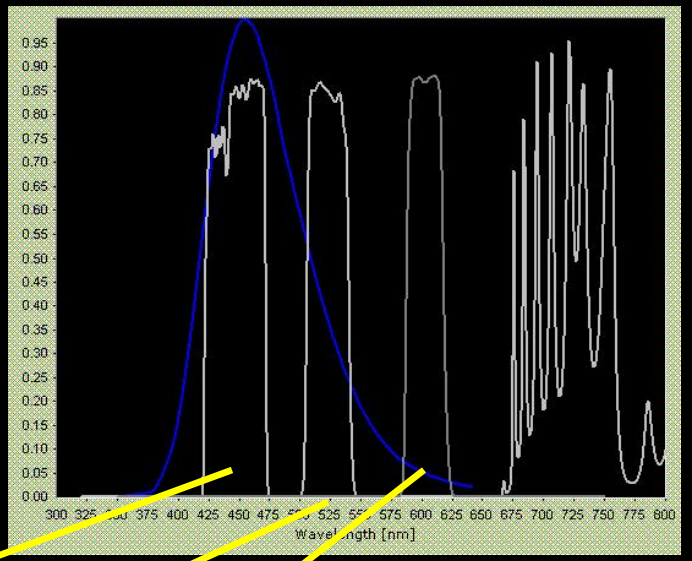
Gosia Kamocka and George Rhodes

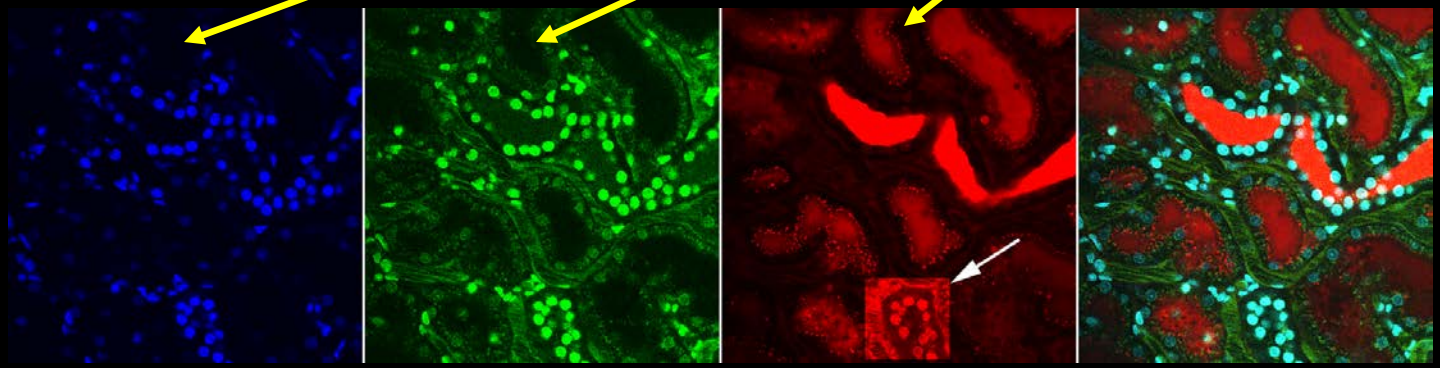
Challenges of intravital microscopy

- Motion artifacts from respiration and heartbeat
- Slow image capture rate
- Limited depth at which useful images can be collected
- Discrimination of different fluors

Exciting (and distinguishing) multiple fluors in multi-photon microscopy



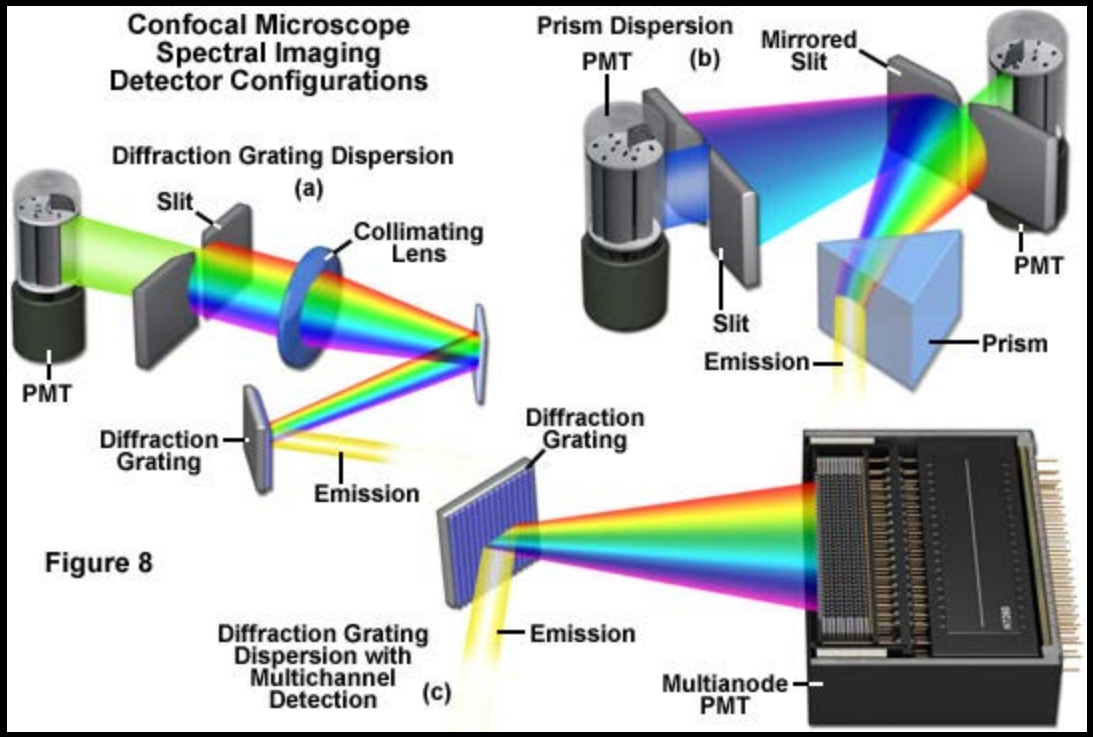




Fluorescence crosstalk correction

- Linear unmixing via spectral detectors

  Effect of scattered light?



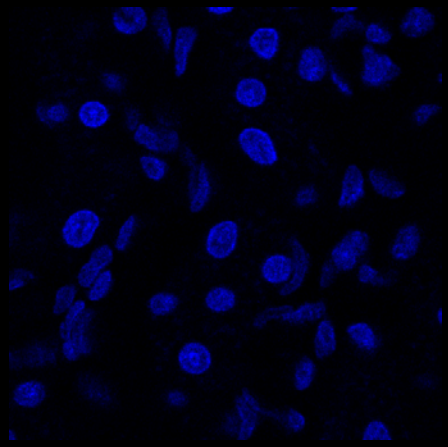
Fluorescence crosstalk correction

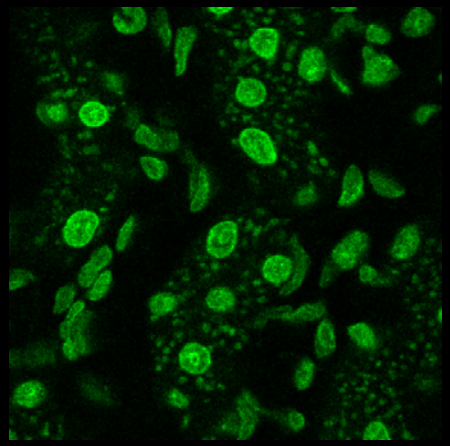
- Linear unmixing via spectral detectors

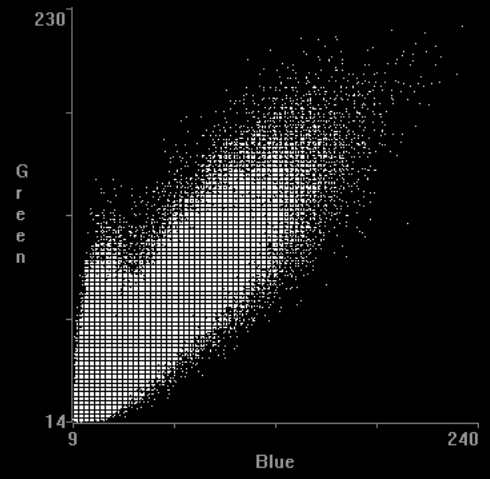
  Effect of scattered light?

- Subtracting a fixed percentage of one channel from another

  Quantitative?



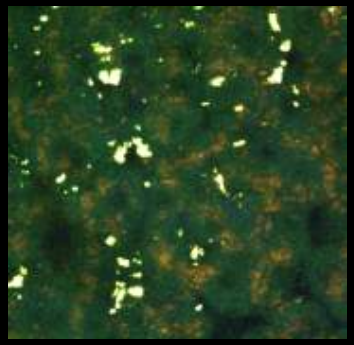


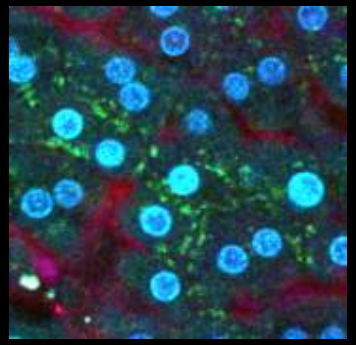


Challenges of intravital microscopy

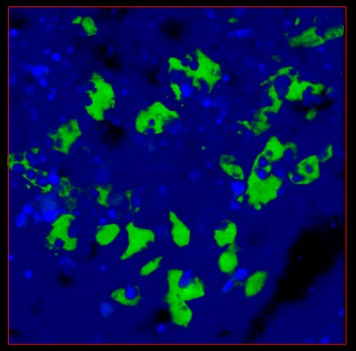
- Motion artifacts from respiration and heartbeat
- Slow image capture rate
- Limited depth at which useful images can be collected
- Discrimination of different fluors
- Introduction of fluorescent probes

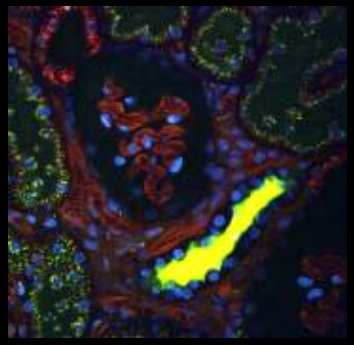
Sources of fluorescence contrast in intravital multiphoton microscopy

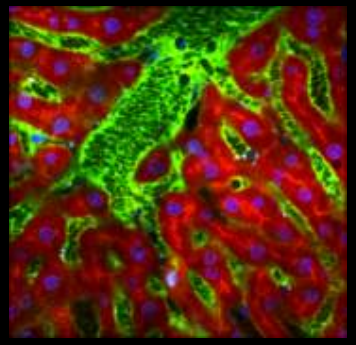




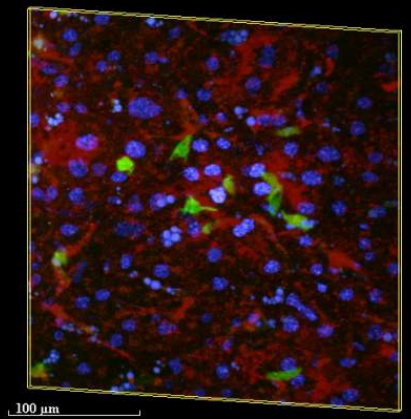
Autofluorescence

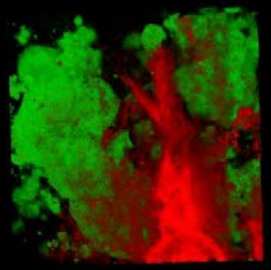


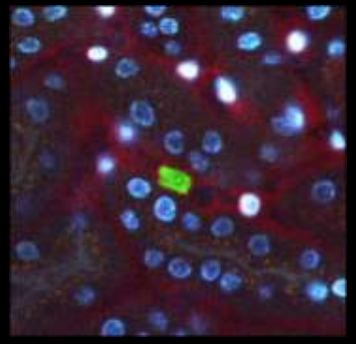




Intravenously introduced probes







Adoptive transfer of fluorescent cells

Transgenic mice

Further information

Websites

http://www.drbio.cornell.edu/ - Watt Webb's laboratory

http://www.microscopyu.com/ - a great general microscopy education website

http://www.aecom.yu.edu/aif/intravital_imaging/introduction.htm - John Condeelis's site

http://www.loci.wisc.edu - Laboratory for Optical and Computational Instrumentation

Reviews

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