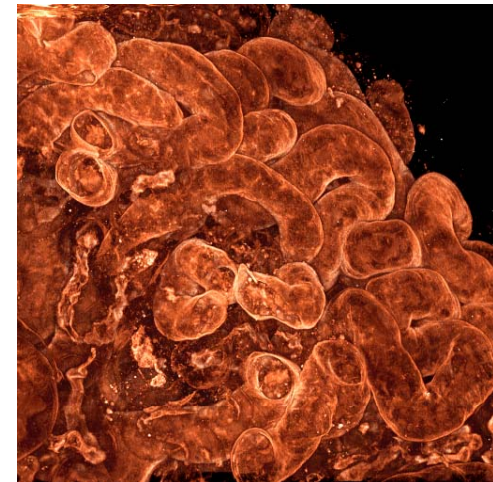
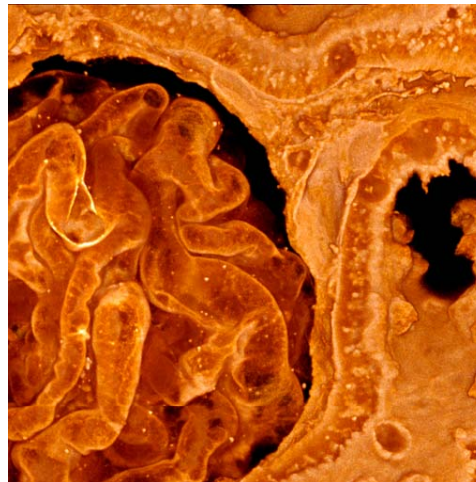
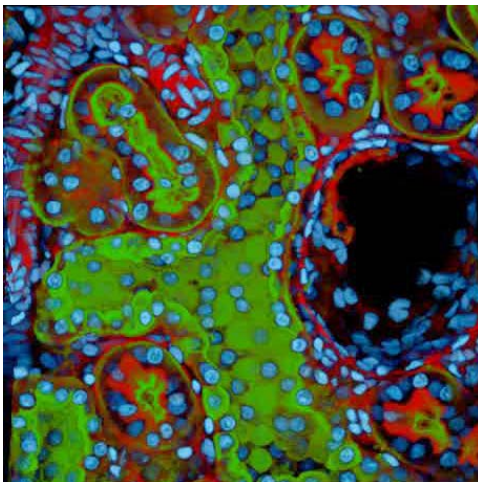
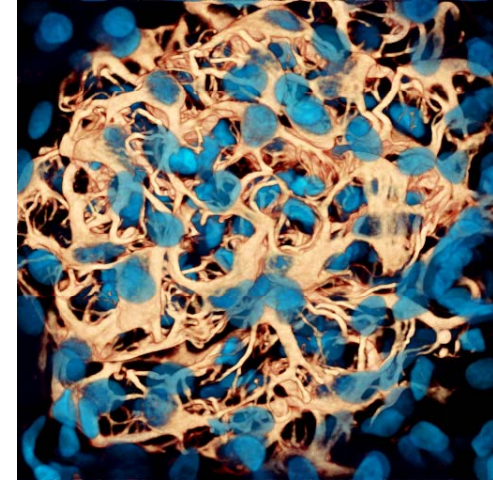
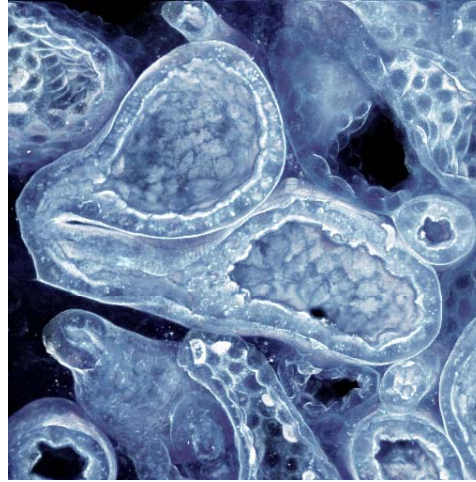
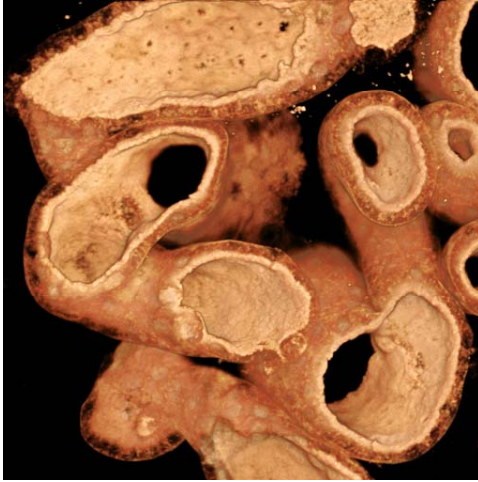


# 3D Visualization and Analysis

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Indiana Center for Biological Microscopy



[www.nephrology.iupui.edu/imaging](http://www.nephrology.iupui.edu/imaging)

# The Problem

**Image-based research is time consuming and labor intensive**

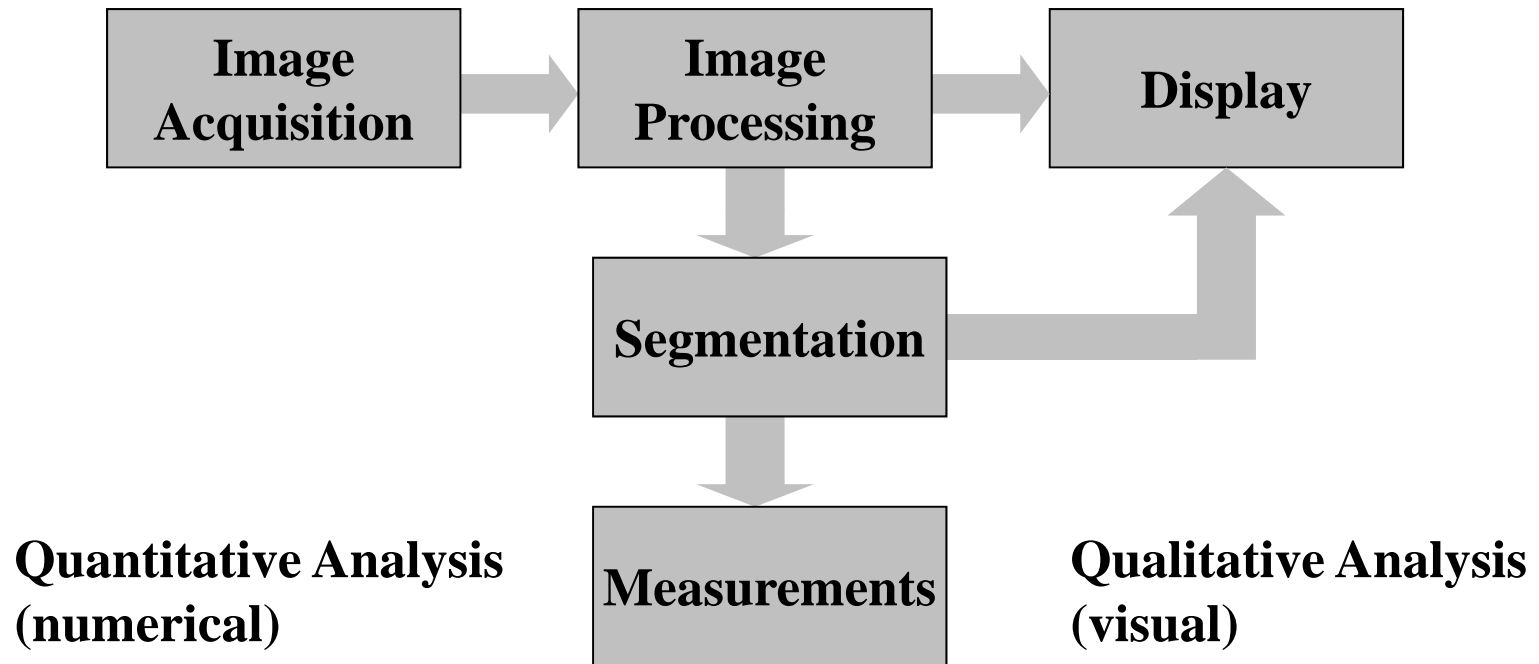
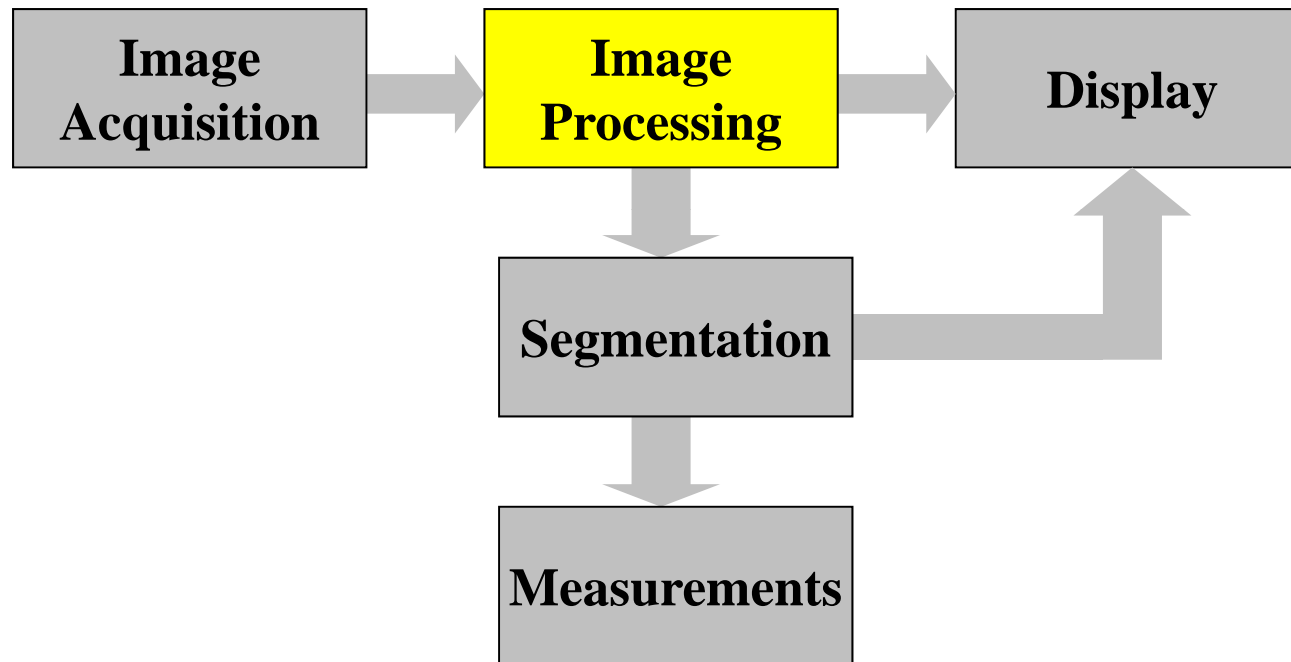


Image processing, segmentation, and rendering are computation-intensive tasks. Some image analysis tasks can be fully automated.

For all of the other tasks, image analysts need (near) real-time visual feedback to perform those tasks...

# Digital Image Processing

The application of any mathematical operation that modifies the original values of pixels in a digitized image



Once you've acquired some images, you often need to process them before they are displayed, or before they can be segmented and measured

# Why do Image Processing? To correct for distortion

Image processing software can sometimes be used to compensate for distortion introduced by instruments and/or tissue during image acquisition



- Limited spatial resolution
- Nonuniform illumination
- Geometric distortion
- Poor signal-to-noise ratio
- Crosstalk
- Chromatic aberration
- etc.

Image acquisition hardware has limitations

# Why do Image Processing? For “image enhancement”

Image processing software can sometimes be used to improve the visibility of biological structures of interest

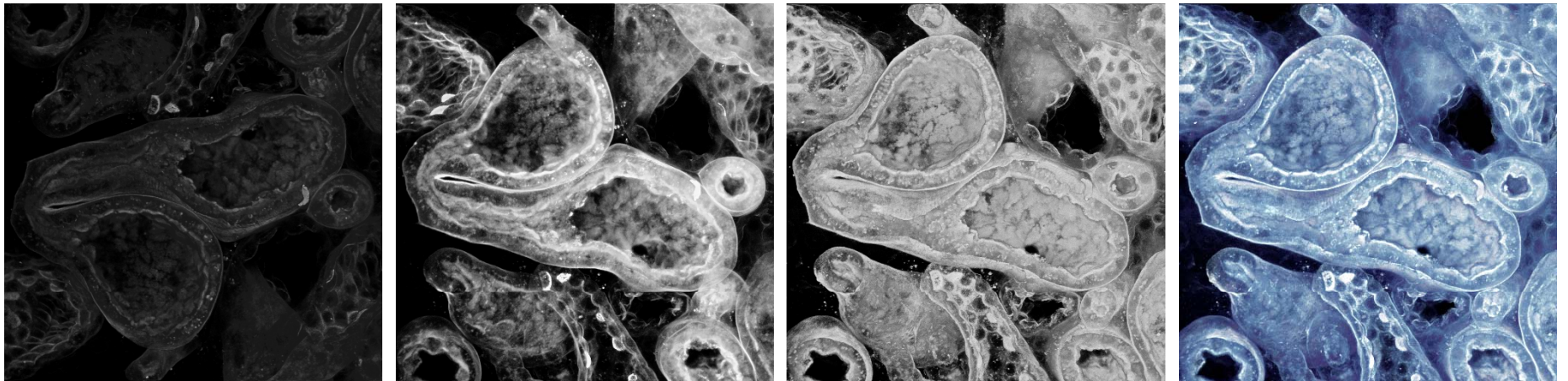


Image display hardware and the human visual system have limitations

# Why do Image Processing? For Segmentation

Before imaging software can be used to measure things, you first have to isolate the structures that you want to measure. That procedure is called segmentation

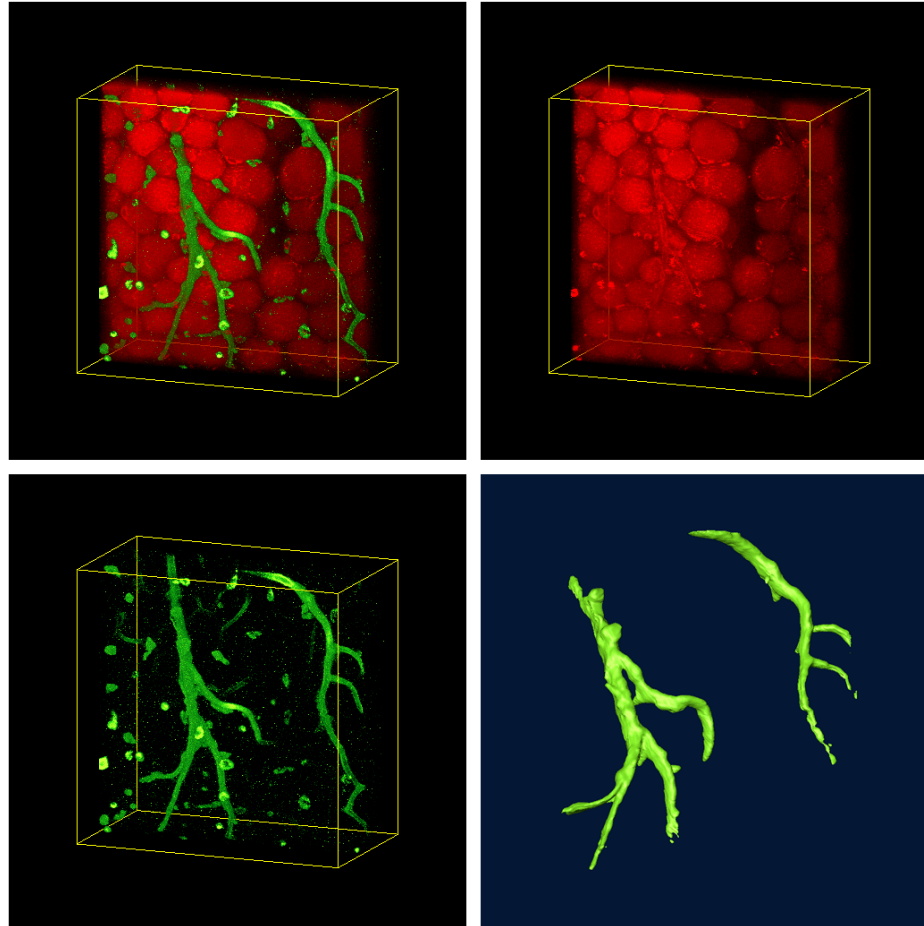
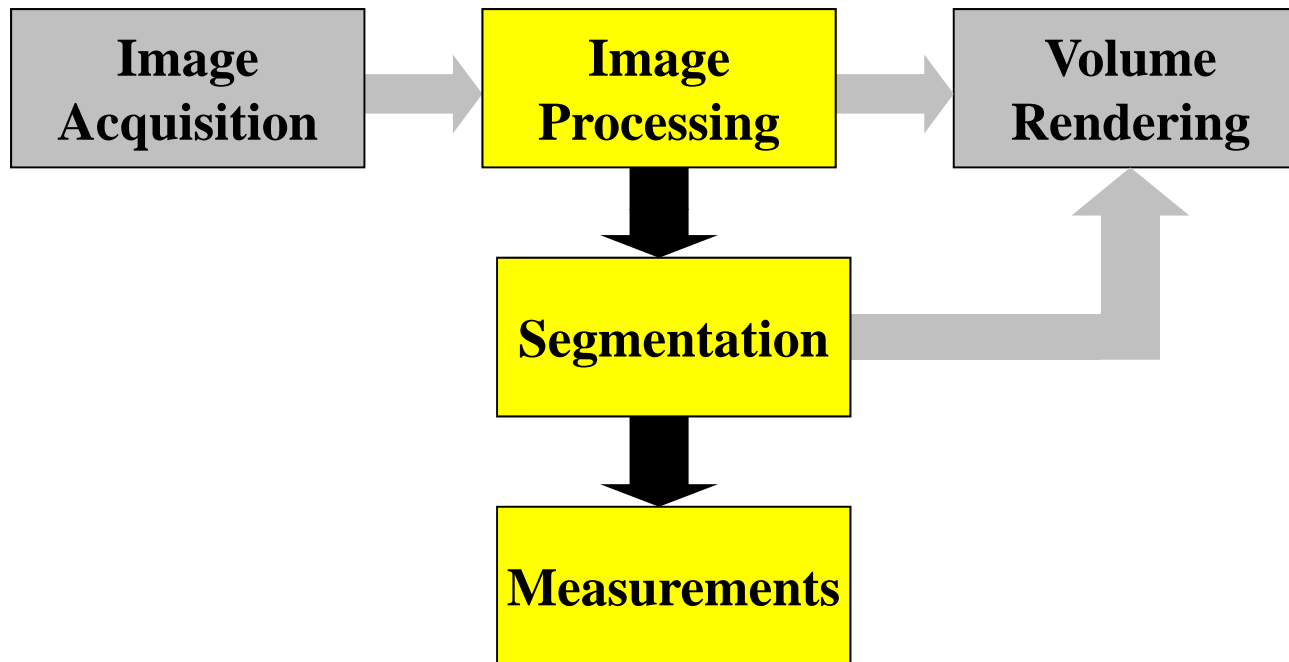


Image processing is required before/during segmentation, to assist in the separation of biological structures from their surroundings

# Automatic Image Segmentation

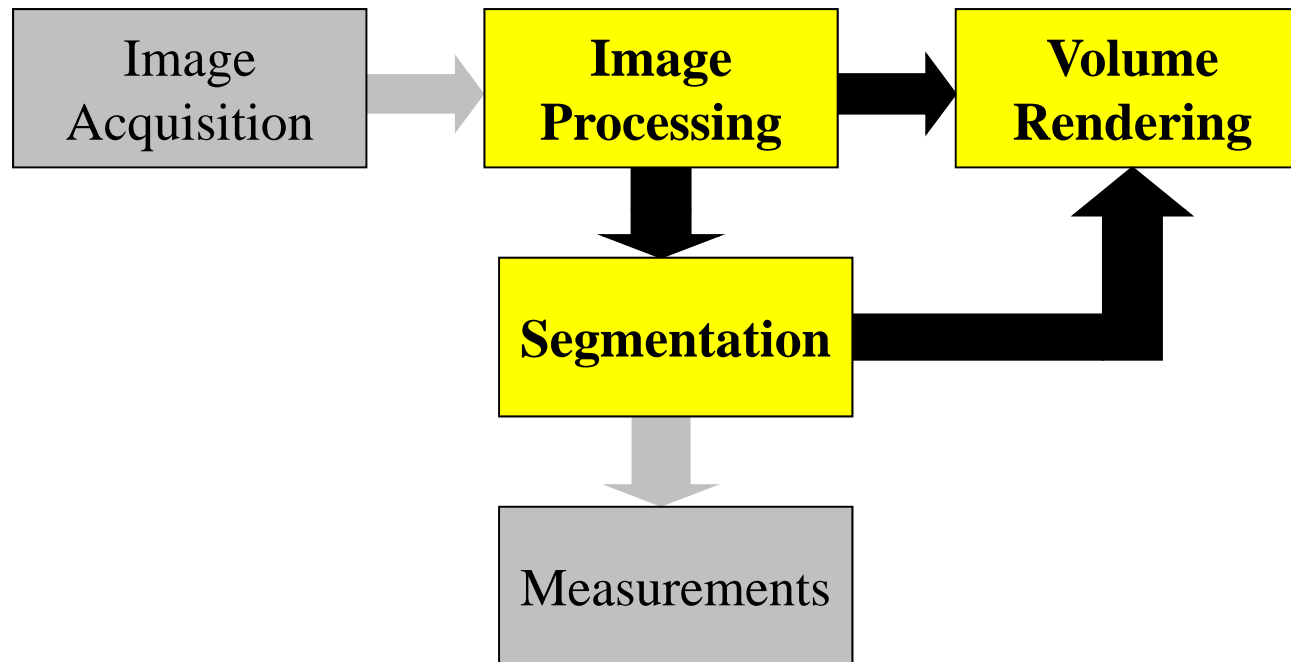
The holy grail for image analysts – reliable, fully automatic segmentation



Quantitative image analysis requires that biological structures in 3D images be recognized and labeled, before measurements can be made

# Interactive Image Segmentation

Interactive segmentation – when fully automatic segmentation isn't feasible

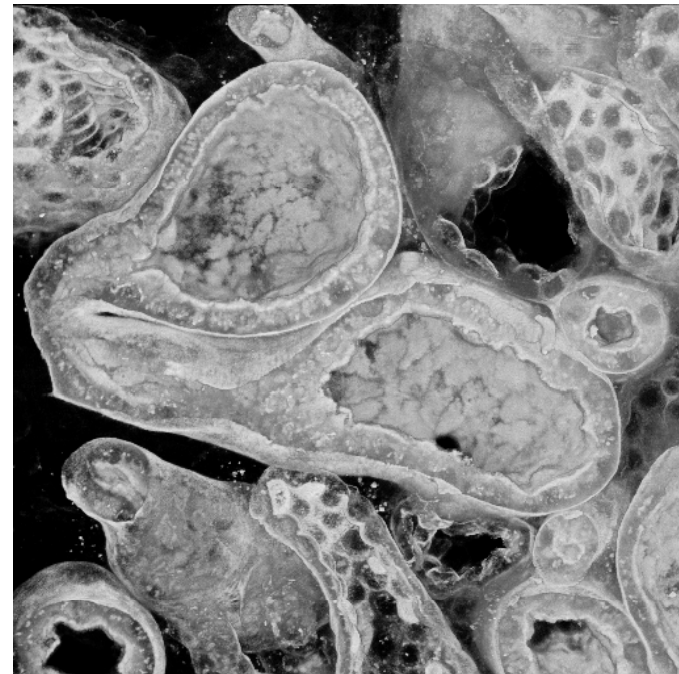
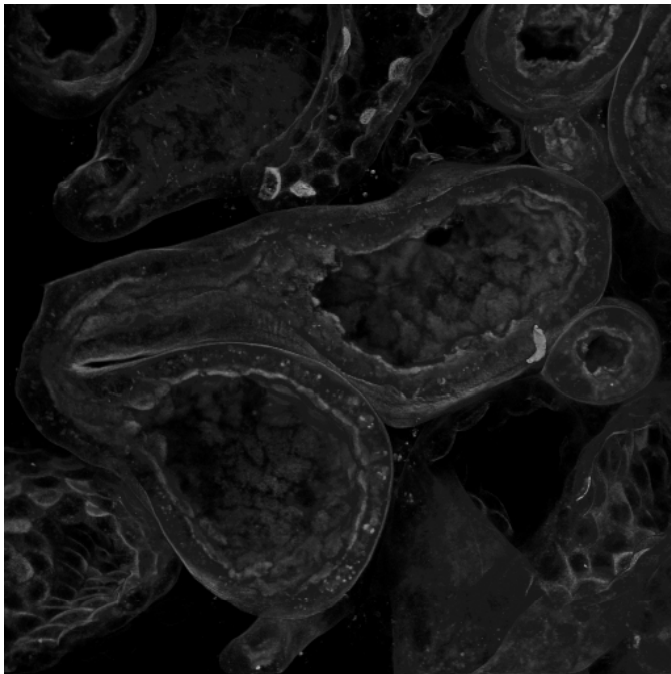


More advanced volume rendering involves both image preprocessing and voxel classification/segmentation. But voxel classification is often still done manually



# Image Processing using Intensity Transforms

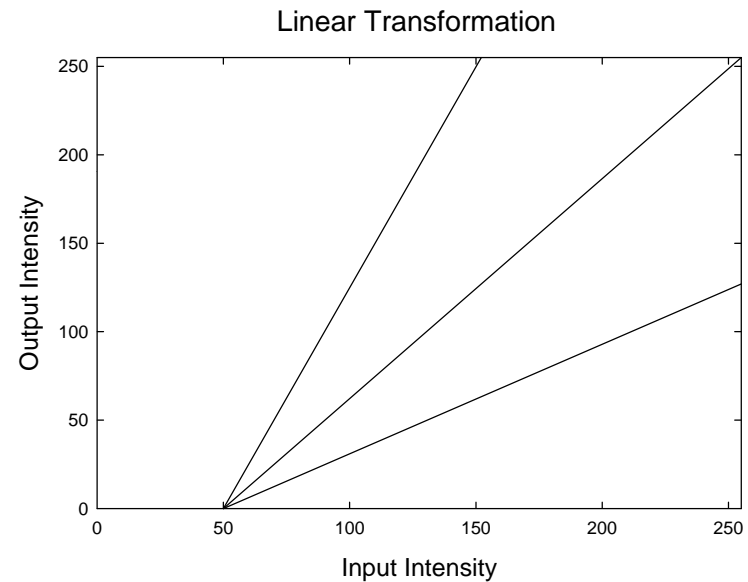
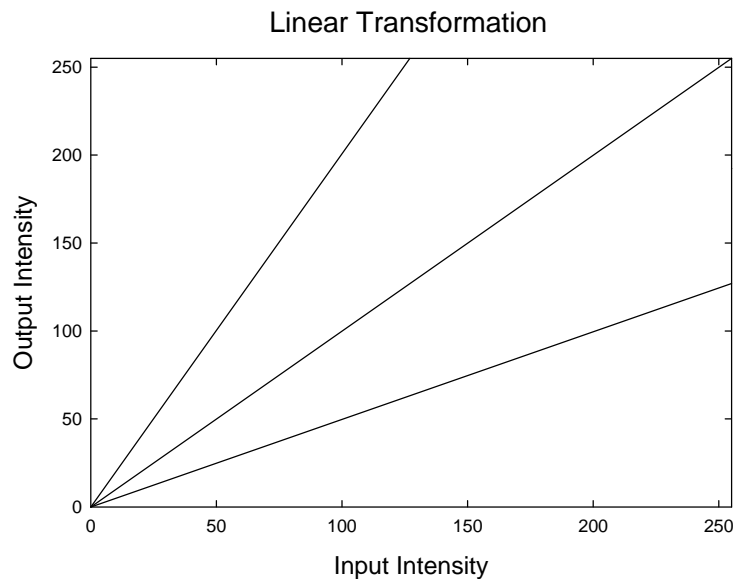
The human eye cannot detect small differences in intensity between regions within an image, even though the pixels have measurably different values



Brightness and contrast stretching operations can make those small differences in intensity visible...

# Intensity Transforms

The most common problem encountered when rendering confocal and two-photon image stacks is that they are dim and low-contrast. Intensity transformations can often be used to improve the visibility of structures.

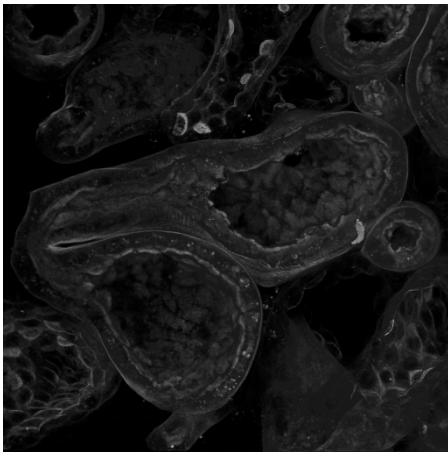


Linear transfer function  $y = m x$ . Setting slope ( $m$ )  $> 1$  will increase the contrast of all pixel values.

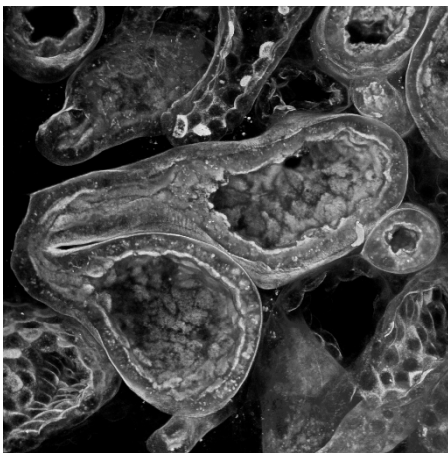
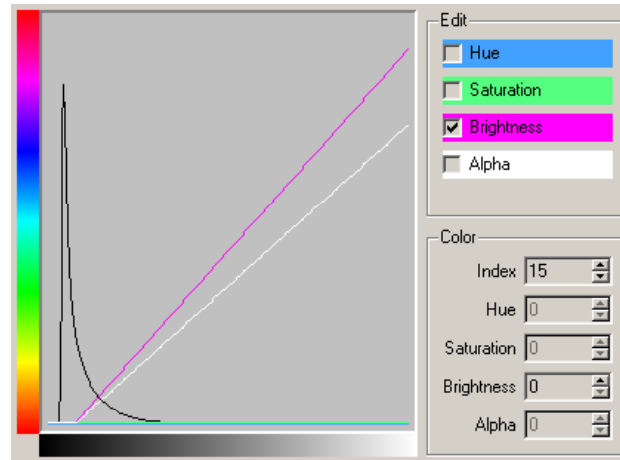
Because of the nonzero background intensity values, you may need to shift the x-axis intercept slightly to the right of the background value(s).

# Intensity Transforms

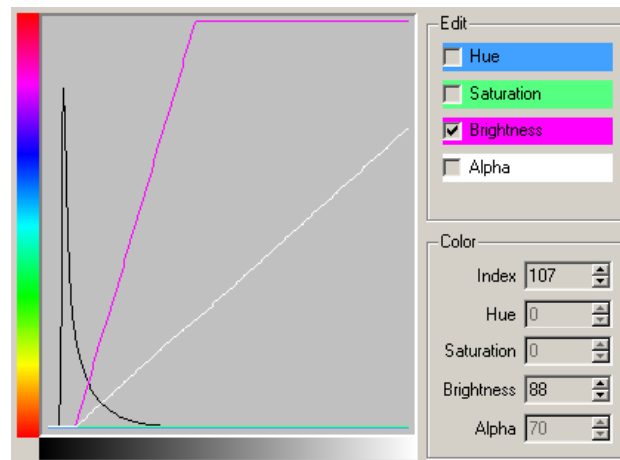
Example of linear transfer functions. Increasing the slope of brightness curve from 1 to 3 increases the brightness and contrast of all pixel values. Also note that x-axis intercept was shifted slightly to the right of the background peak



**Slope=1**

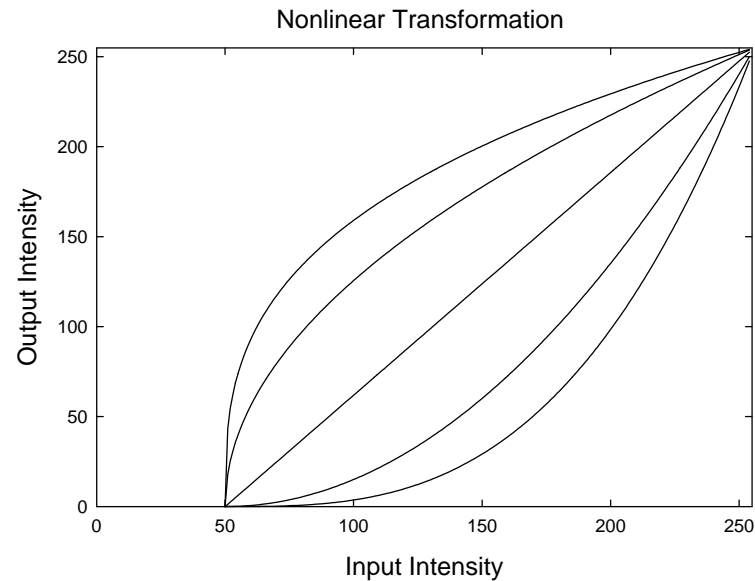
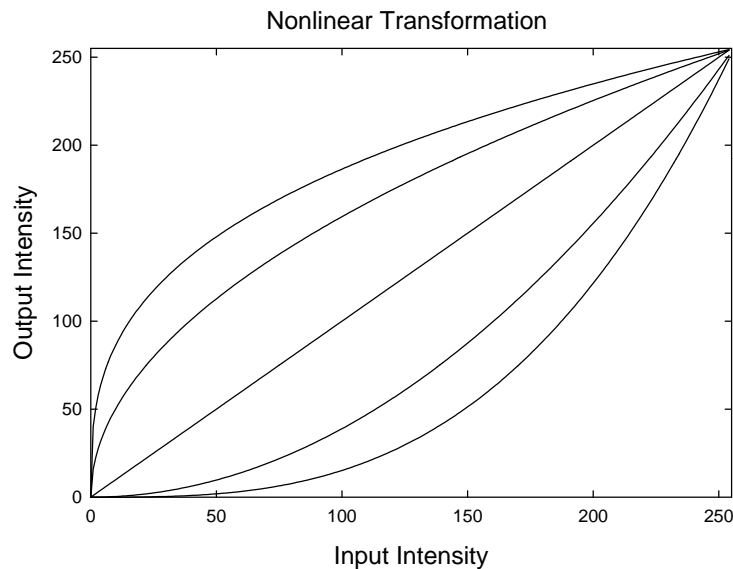


**Slope=3**



# Intensity Transforms

The most common problem encountered when rendering confocal and two-photon image stacks is that they are dim and low-contrast. Intensity transformations can often be used to improve the visibility of structures.

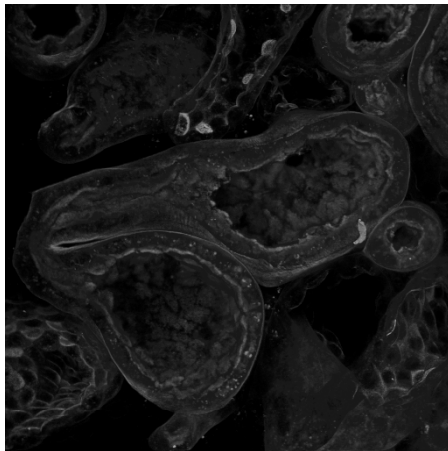


Power-law transfer function  $y = x^a$ . Setting power ( $a$ )  $< 1$  will increase the contrast of dim pixels and decrease the contrast among bright pixels.

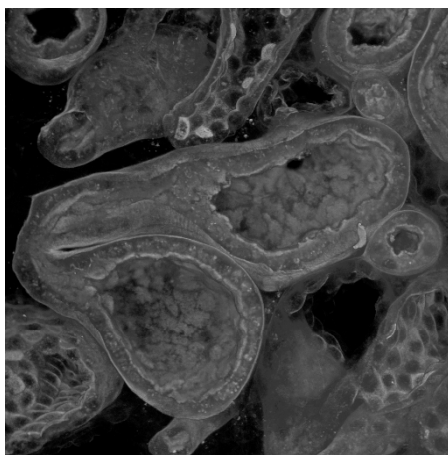
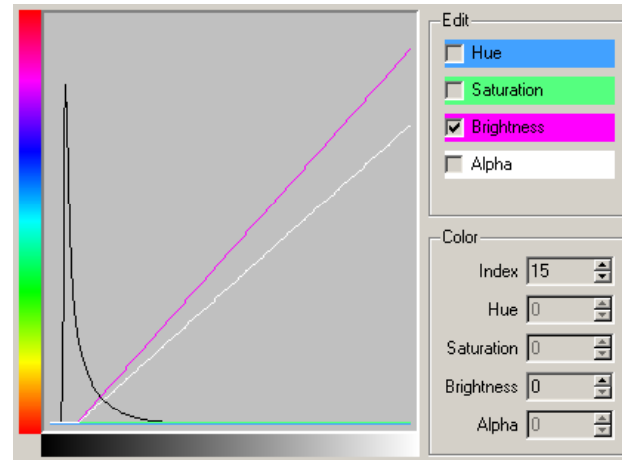
Because of the nonzero background intensity values, you may need to shift the x-axis intercept slightly to the right of the background value(s).

# Intensity Transforms

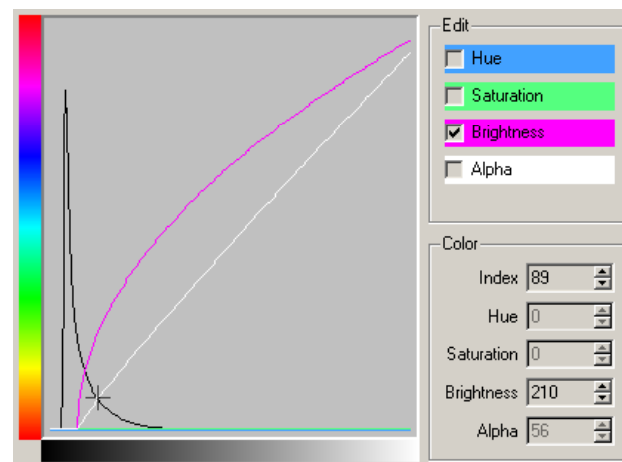
Example of power-law transfer functions. Decreasing the power will increase the brightness and contrast of dim pixels and decrease the contrast among bright pixels.



**Power=1**

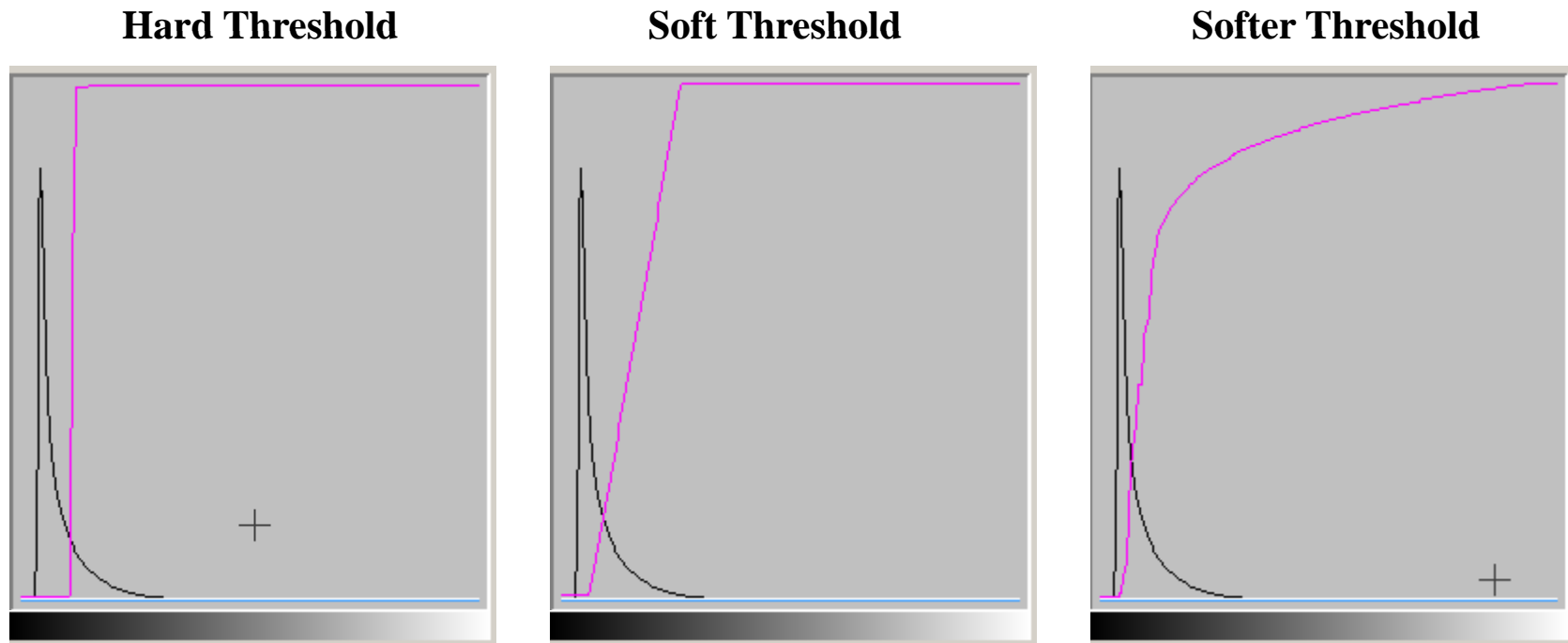


**Power=0.5**



# Intensity Transforms: Hard vs Soft Thresholding

Intensity transformations used to modify brightness and contrast can also be used to implement thresholding, by replacing low-intensity pixel values with zero.

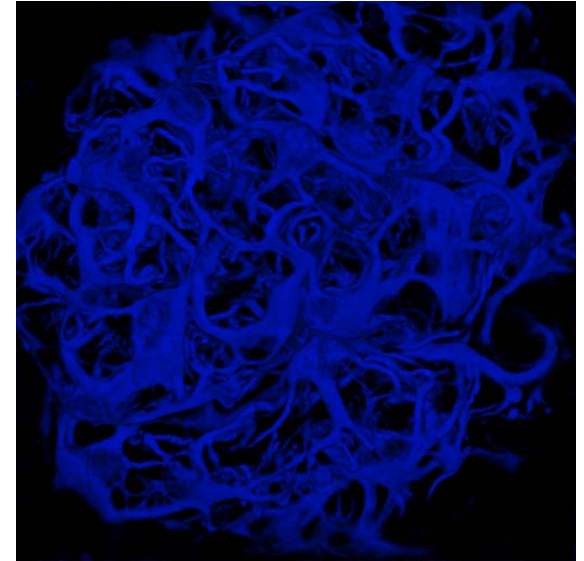
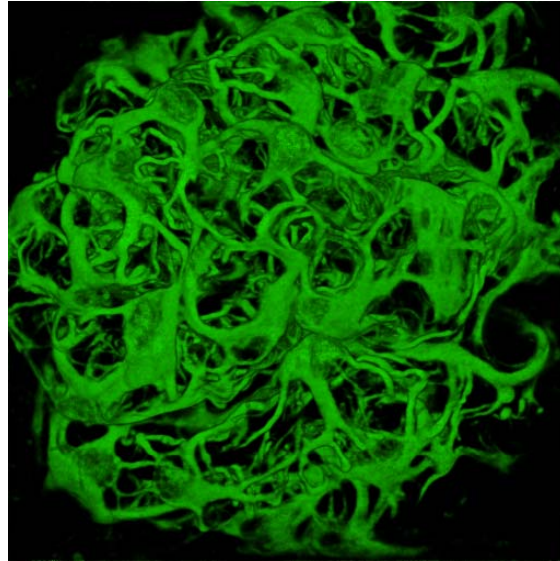
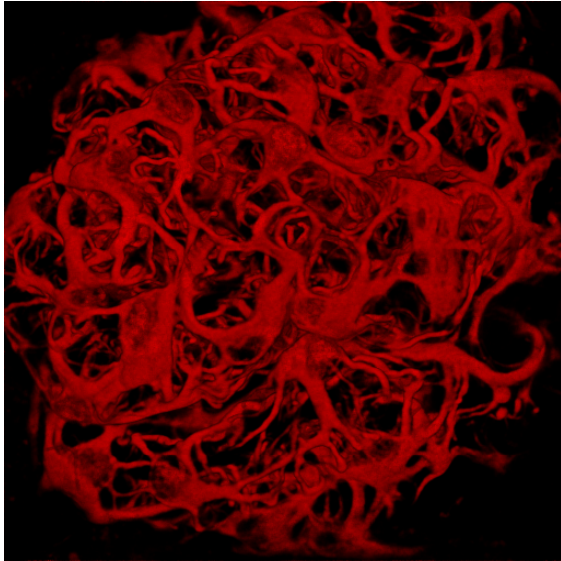


“Hard thresholding” typically replace all pixel values below a given value with zero, and replaces all pixel values greater than or equal to that value with a nonzero value.

“Soft thresholding” has a transition region, instead of a single threshold value, in which pixel intensities are made to rapidly increase from zero to some nonzero value

# Color Transforms

Your choice of colors affects the visibility of structures, and whether the printed image will look more or less like the on-screen image

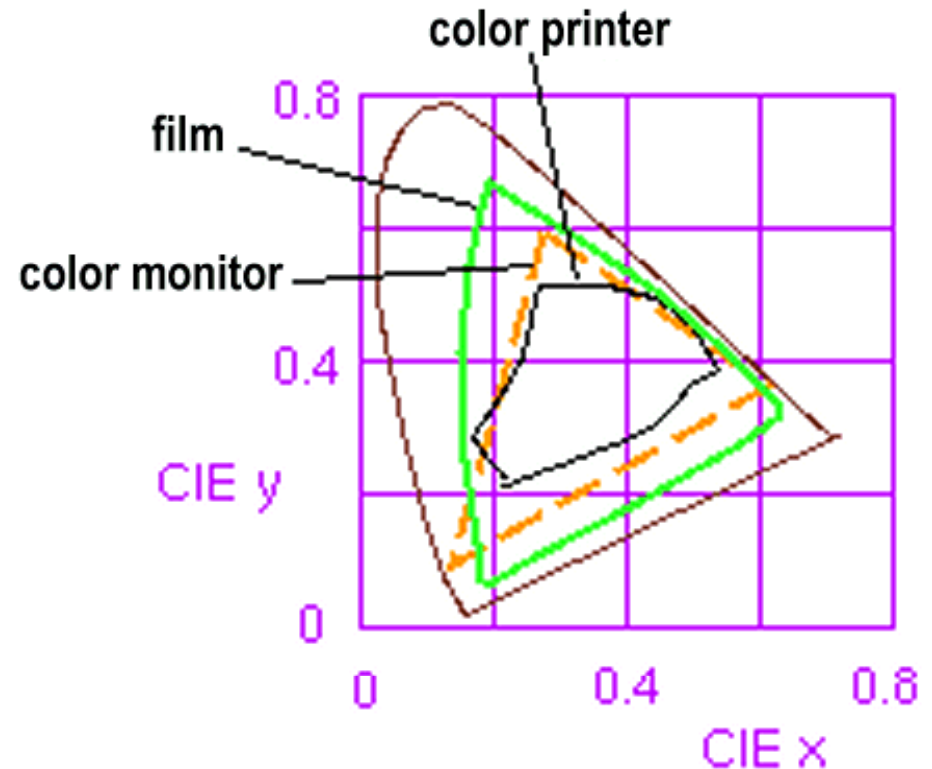
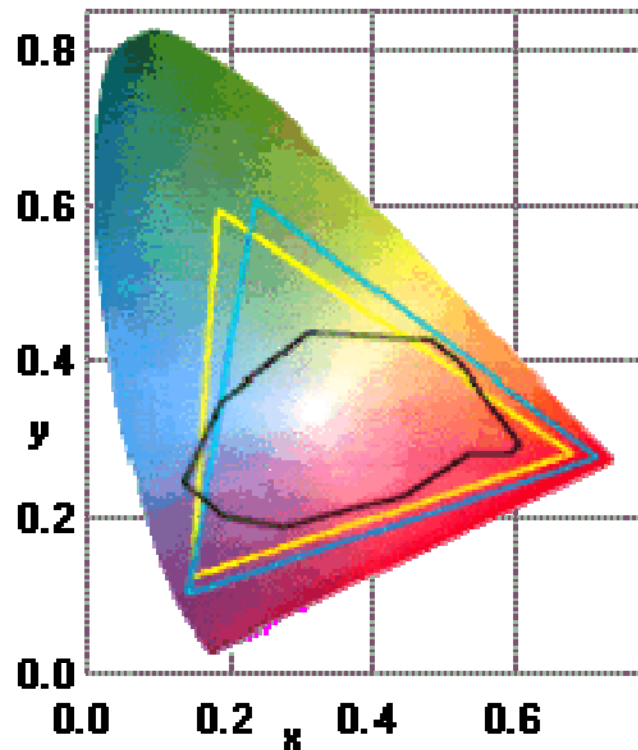


Red and blue objects can be difficult to see in on-screen images, because the human eye is less sensitive to red and blue, and more sensitive to green. Try using a mixture of the primary RGB colors (e.g. reddish-orange instead of red for Rhodamine, cyan instead of blue for DAPI).

And it can be difficult to get printed images to look like the on-screen image when using highly saturated red, green, or blue. Monitors can display colors that are not reproducible on printers. Try slightly reducing color saturation...

# Colorization

The choice of colors affects the visibility of structures, and whether the printed image will look more or less like the on-screen image



It can be difficult to get printed images to look like the on-screen image when using highly saturated red, green, or blue. Monitors can display colors that are not reproducible on printers. Try slightly reducing saturation in HSV, to move a color inside the printer's color gamut...



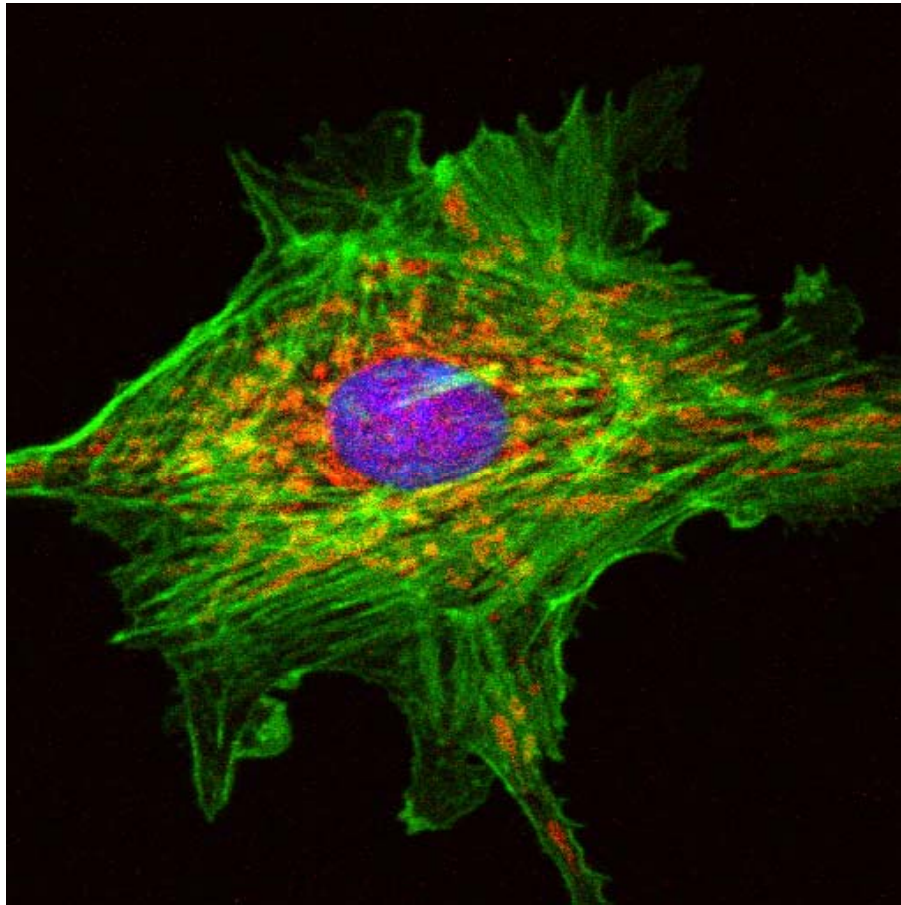
# Image Processing using Filters

Filters use the values of neighboring pixels to compute a new/filtered value for each pixel in an image

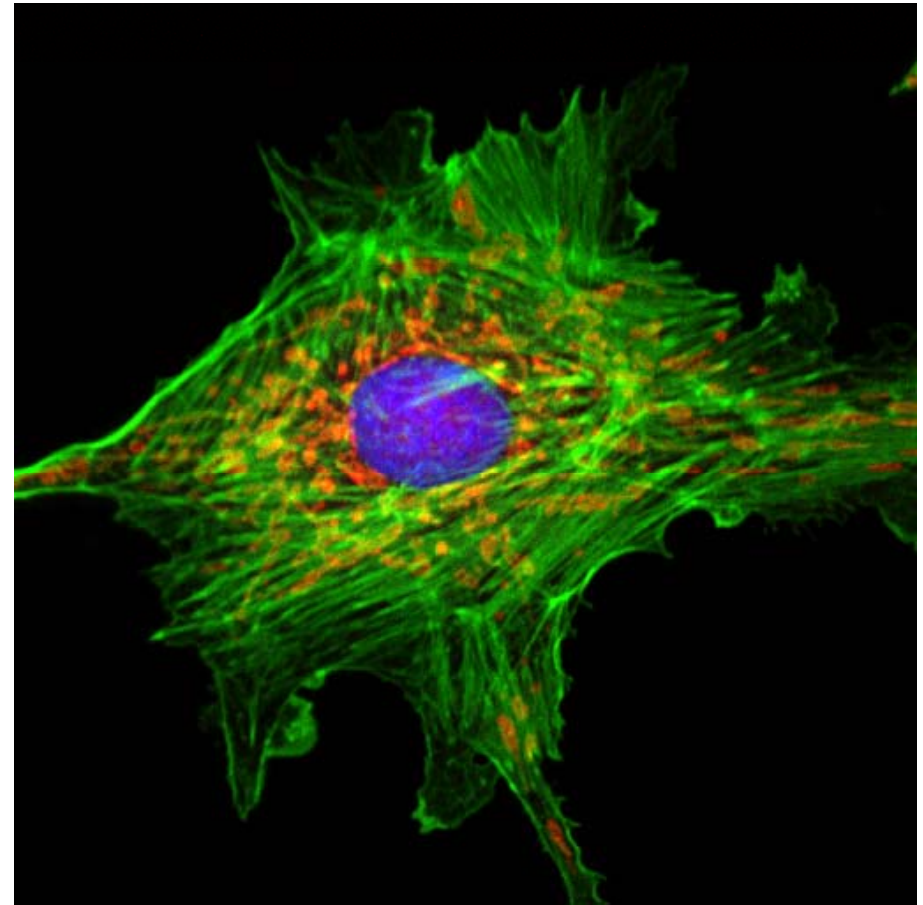
Convolution:	<ol style="list-style-type: none"><li>1) arbitrary filter kernels, non-separable</li><li>2) special cases, separable kernels</li><li>3) special cases, recursive</li></ol>
Diffusion:	<ol style="list-style-type: none"><li>1) nonlinear isotropic</li><li>2) nonlinear anisotropic</li><li>3) reaction-diffusion</li></ol>
Transform-based:	<ol style="list-style-type: none"><li>1) FFT or Hartley</li><li>2) wavelets</li></ol>
Deconvolution:	<ol style="list-style-type: none"><li>1) constrained iterative methods</li><li>2) transform-based methods</li></ol>
Morphological:	<ol style="list-style-type: none"><li>1) grayscale morphological (min, max, etc)</li><li>2) binary morphological (erode, dilate, etc)</li></ol>

# Noise Reduction

Some kinds of image processing are performed during image acquisition  
(e.g. filters that perform denoising using clever forms of averaging)



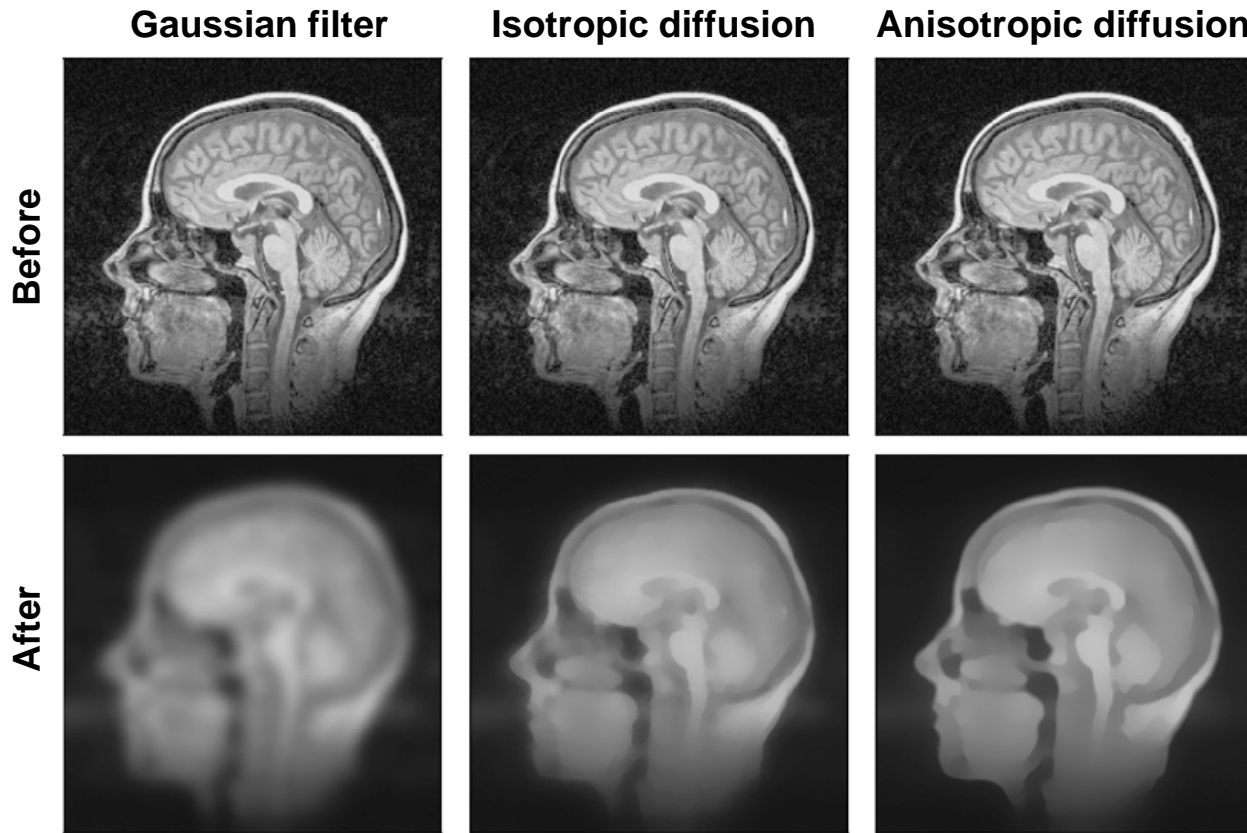
Noisy Image of Cell



Averaged Image

# Smoothing (Averaging) Filters

Iterative smoothing filters based on nonlinear diffusion equations preserve the boundary between different structures better than Gaussian filters



Original work on nonlinear diffusion filters in 2D image processing done by Perona and Malik (1990). Above images by Langer (2004).

# Deconvolution Filters

Some kinds of image processing are performed after image acquisition  
(e.g. deconvolution, crosstalk correction)

**Before Deconvolution**

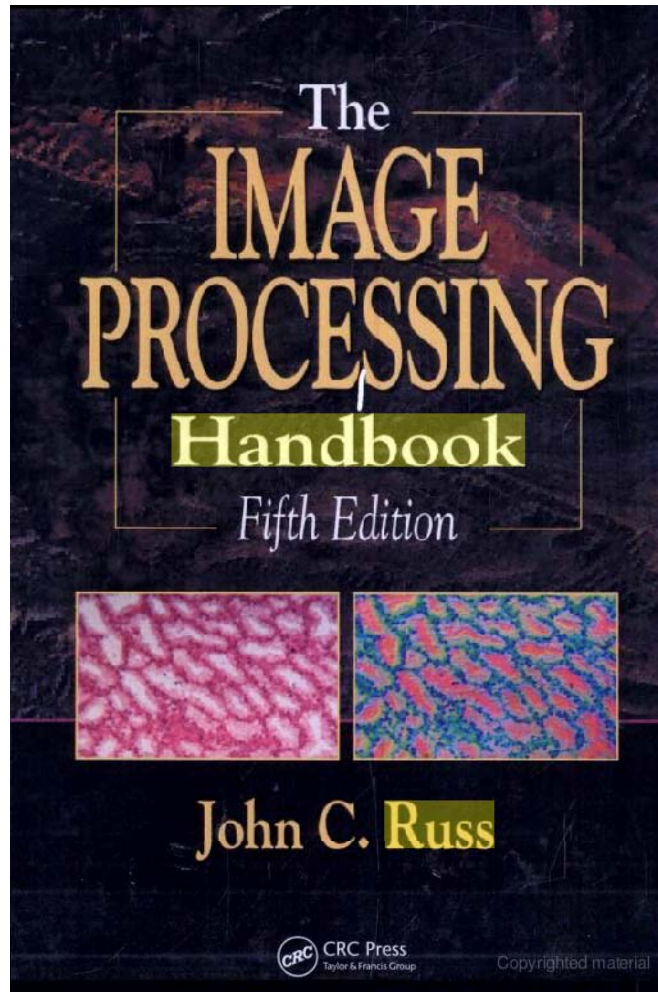


**After Deconvolution**



3D deconvolution software can be used to compensate to some degree for the poor axial resolution of microscope optics. It can also improve brightness and contrast.

# A Good Introduction to Image Processing – with the minimum of math



## **IUPUI main library has**

4<sup>th</sup> edition (TA1637 .R87 2002)

3<sup>rd</sup> edition (TA1637 .R87 1999)

2<sup>nd</sup> edition (TA1637 .R87 1995)

## **Medical school library has**

1<sup>st</sup> edition (TA1632 R958i 1992)

## **Dental school library has**

1<sup>st</sup> edition (WN 160 R958i 1992)

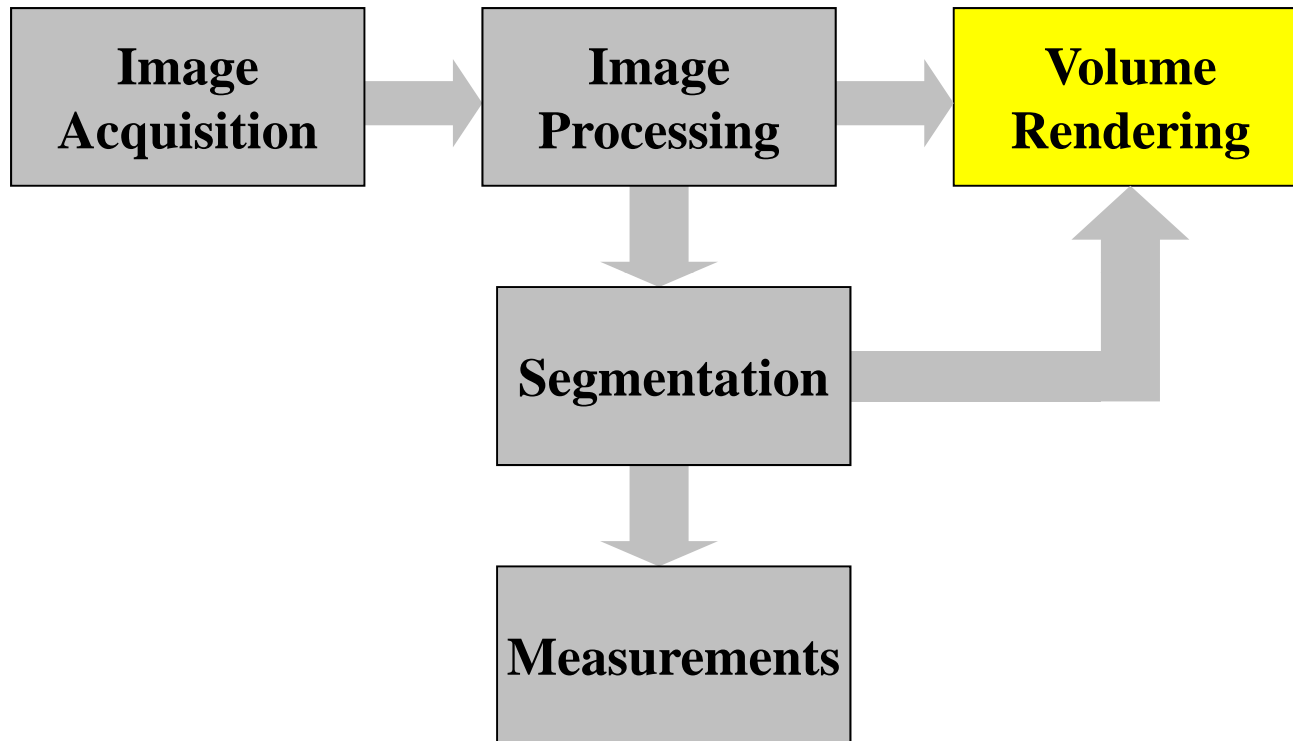
**5th edition (2006) can be purchased for \$130-165 from many booksellers (e.g. amazon.com)**

**ISBN-10: 0849372542**

**ISBN-13: 978-0849372544**

# Volume Rendering

Volume rendering programs can be used to create 2D projection images showing the 3D stacks of cross-sectional images from various points of view.

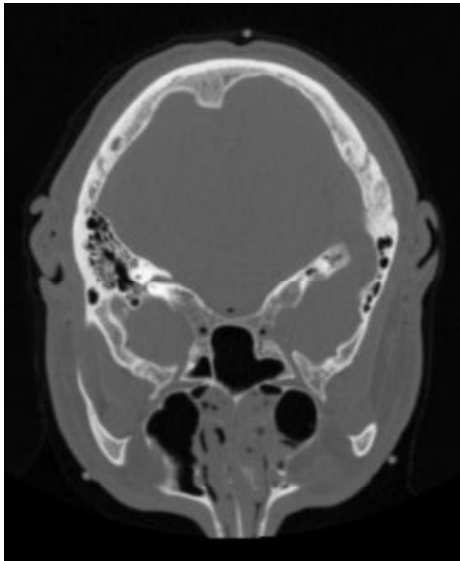


Volume renderers can display individual cross sections, but they can also display many cross sections simultaneously, which makes it easier to study 3D morphology

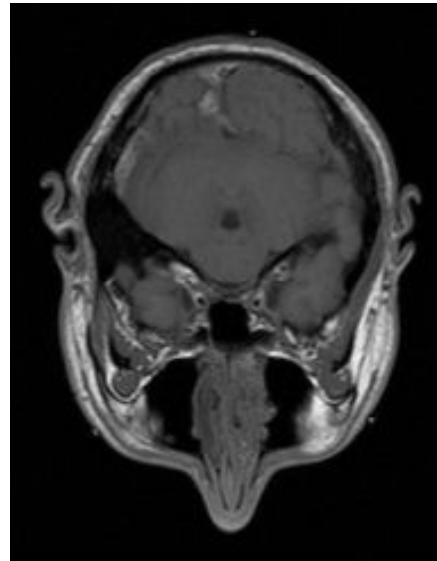
# Cross-Sectional Imaging

Several imaging technologies have been developed over the years that can produce cross-sectional images, such as CT and MR in radiological imaging, and confocal and two-photon techniques in optical microscopy.

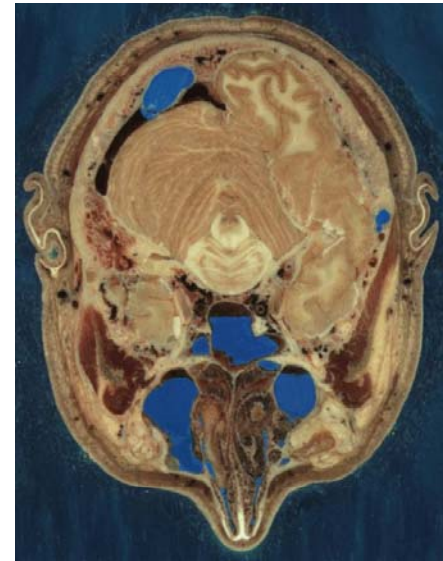
**Computed Tomography**



**Magnetic Resonance**



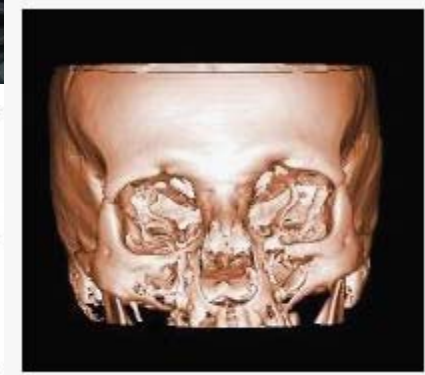
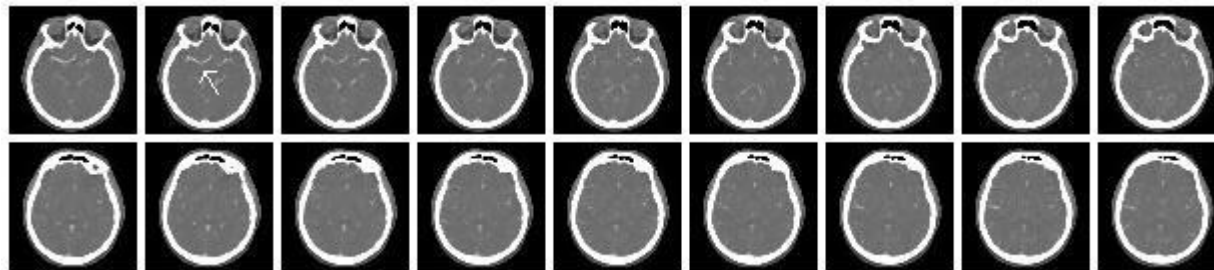
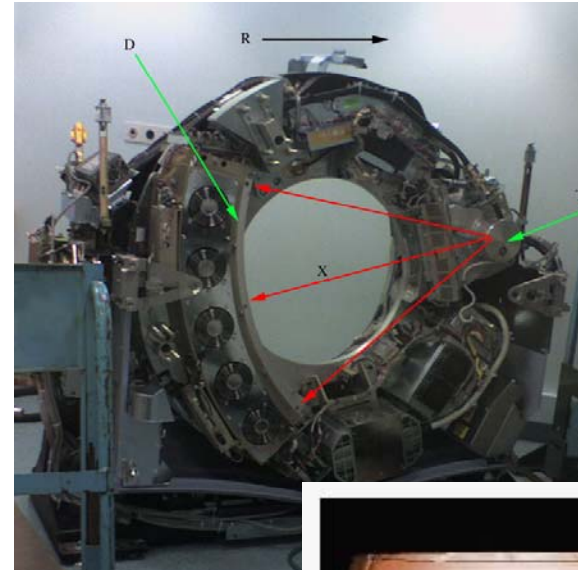
**Blockface**



e.g. Visible Human Project [1994], cross sections of human head.  
[http://www.nlm.nih.gov/research/visible/visible\\_human.html](http://www.nlm.nih.gov/research/visible/visible_human.html)

# CT Image Acquisition

In computed tomography (CT), a rotating gantry carrying x-ray sources and detectors measures x-ray absorption from several directions within a thin slab of tissue. This 1D data is converted into a cross-sectional (2D) image using reconstruction software...

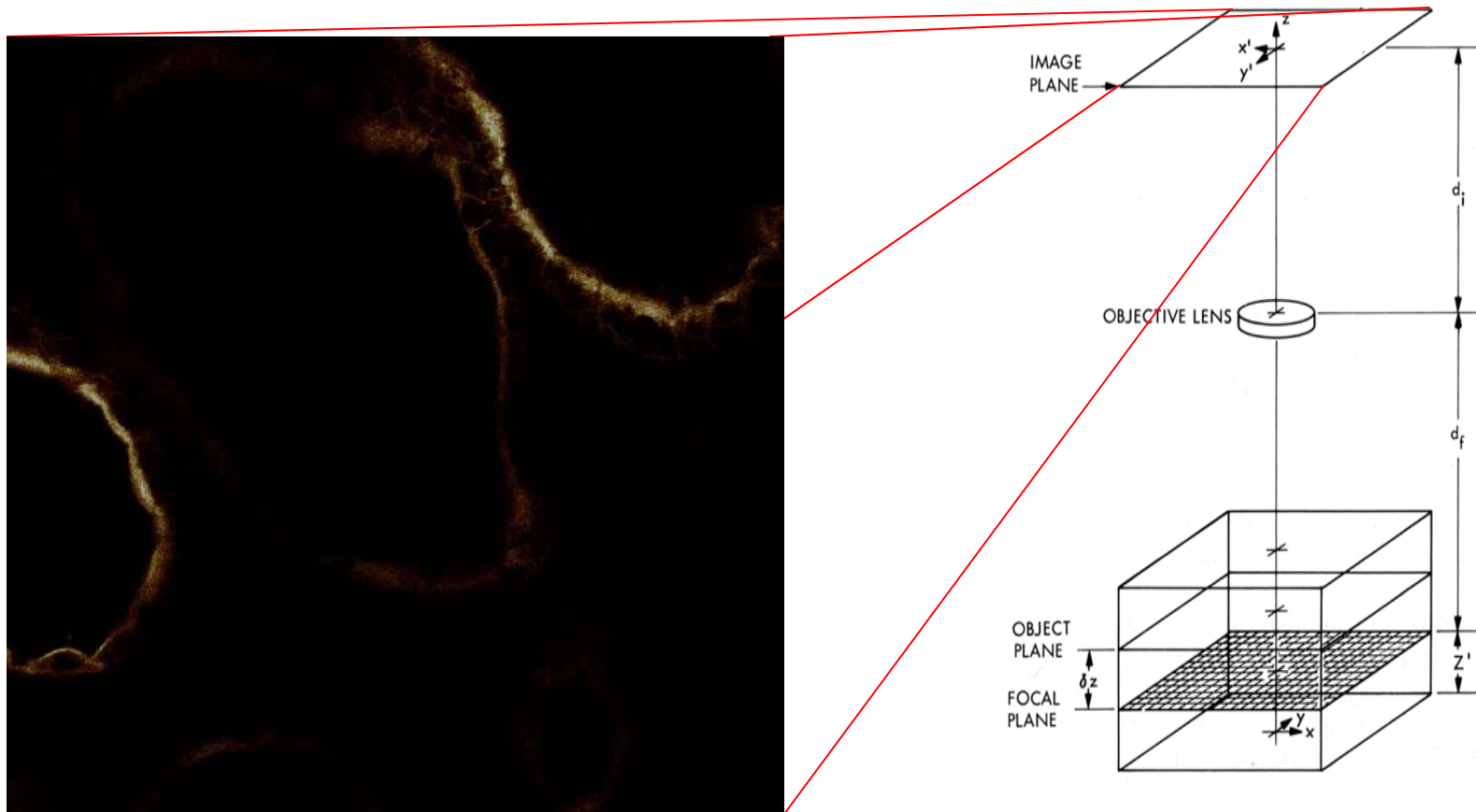


A motorized table then moves the subject to the next position. A set of parallel 2D images is collected by repeating the procedure. Volume rendering software displays them in 3D.



# Image Acquisition using Light Microscopy

Several kinds of optical microscopes use laser scanners and clever optics to collect a set of very thin parallel planar (2D) images, including:  
laser scanning confocal microscopes, spinning disk confocal, two-photon, etc.



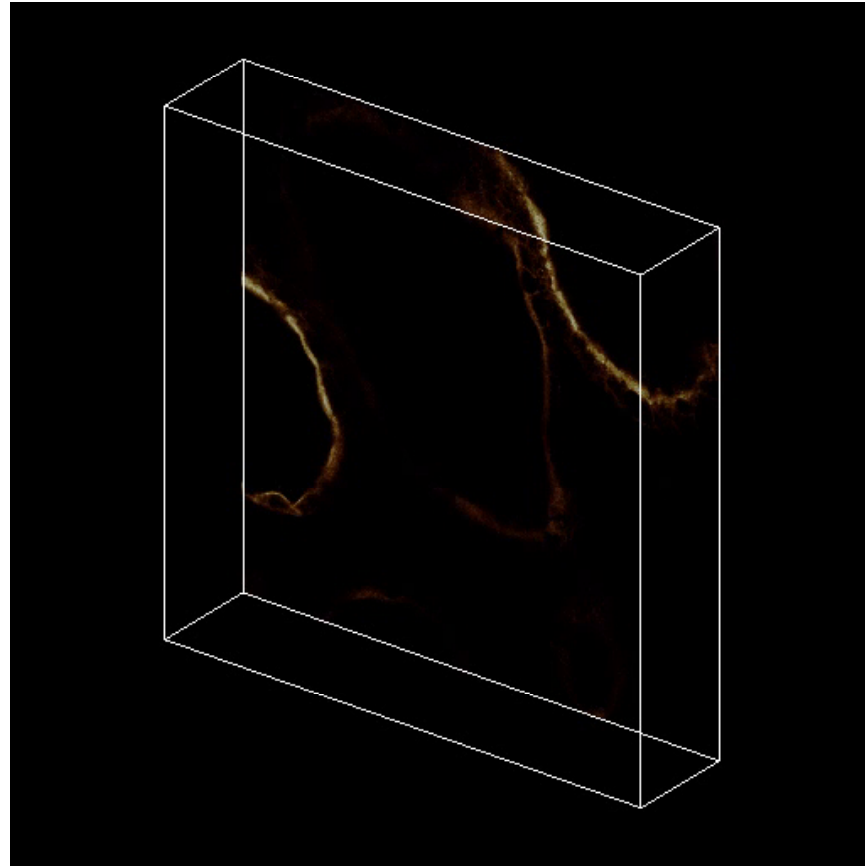
movie

K. R. Castleman,  
Digital Image Processing  
(c) 1979 Prentice Hall

# Volume Rendering: How It Works

The 2D projections of your cross-sectional images are combined as they are rendered in back-to-front order, forming a 2D composite image that is displayed on the monitor

**Laser Scanning Confocal Microscopy**

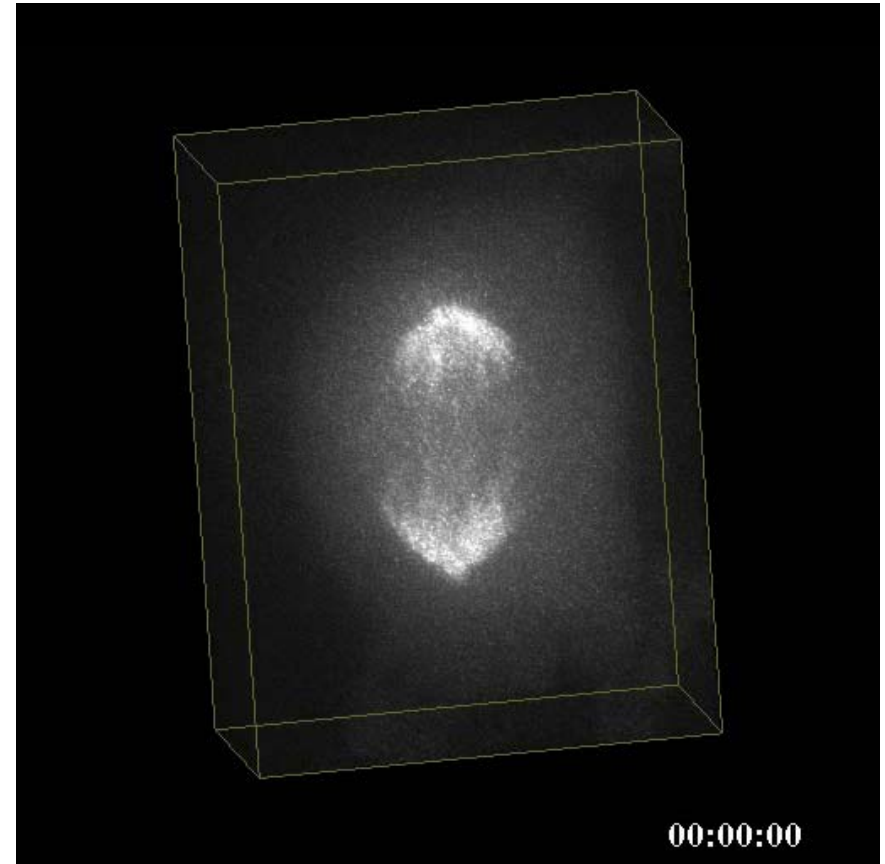
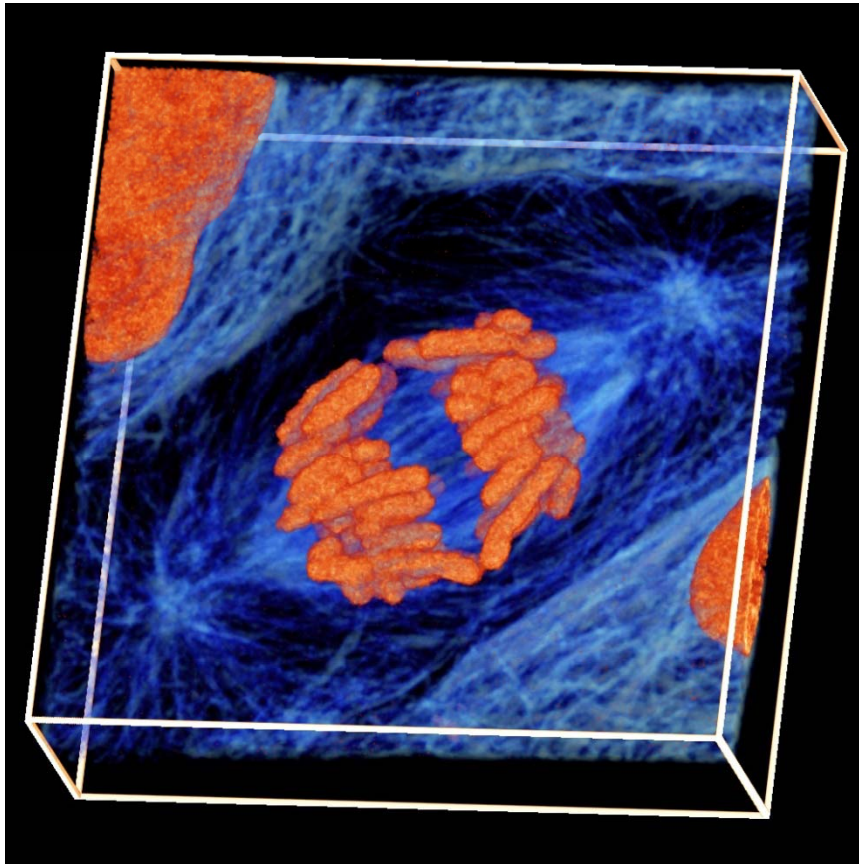


**movie**

You select a function (e.g. max, mean, etc.) that is used to combine the projection images.

# 4D Imaging (3D + Time)

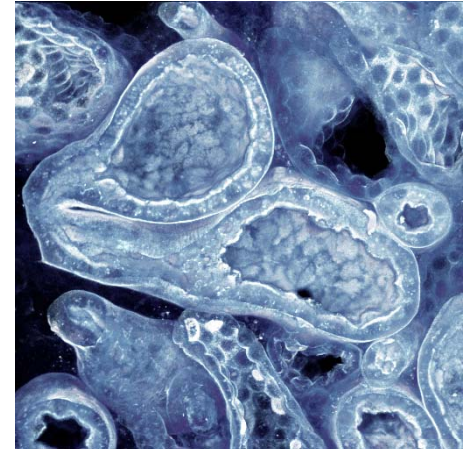
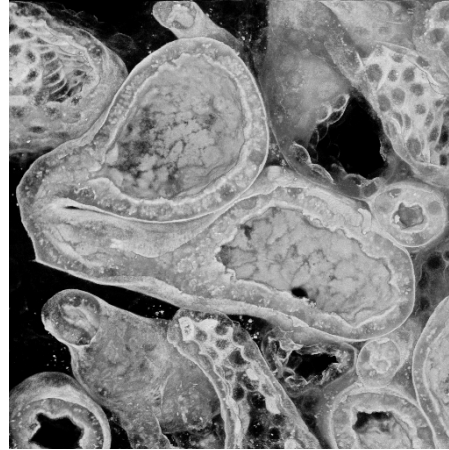
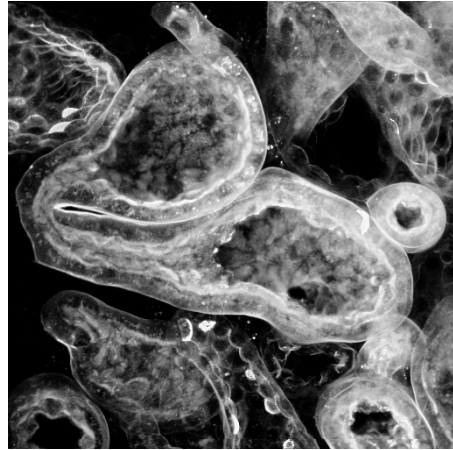
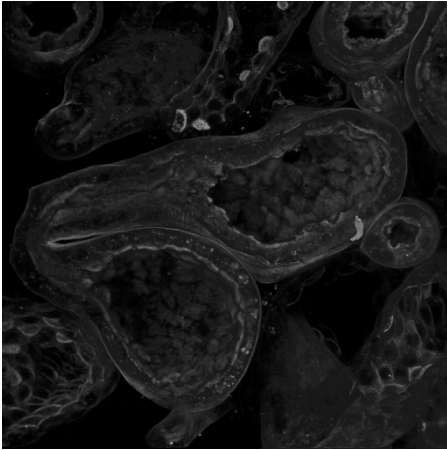
You can collect and display sequences of 3D images, which is useful for developmental studies



e.g. dividing cell (chromosomes and mitotic spindle)

# Volume Rendering: Setup

Volume rendering programs must be carefully configured to produce a high-quality 3D effect. Here is a typical sequence of operations that needs to be performed:



**1) Select blending mode** – alpha, sum, or max.

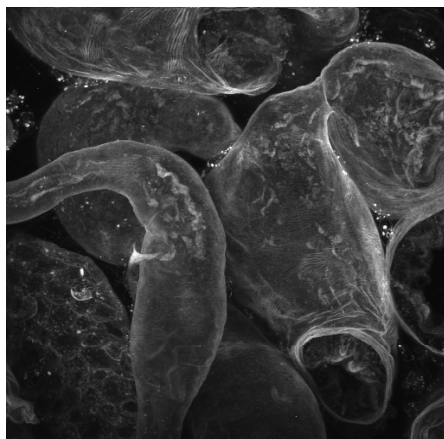
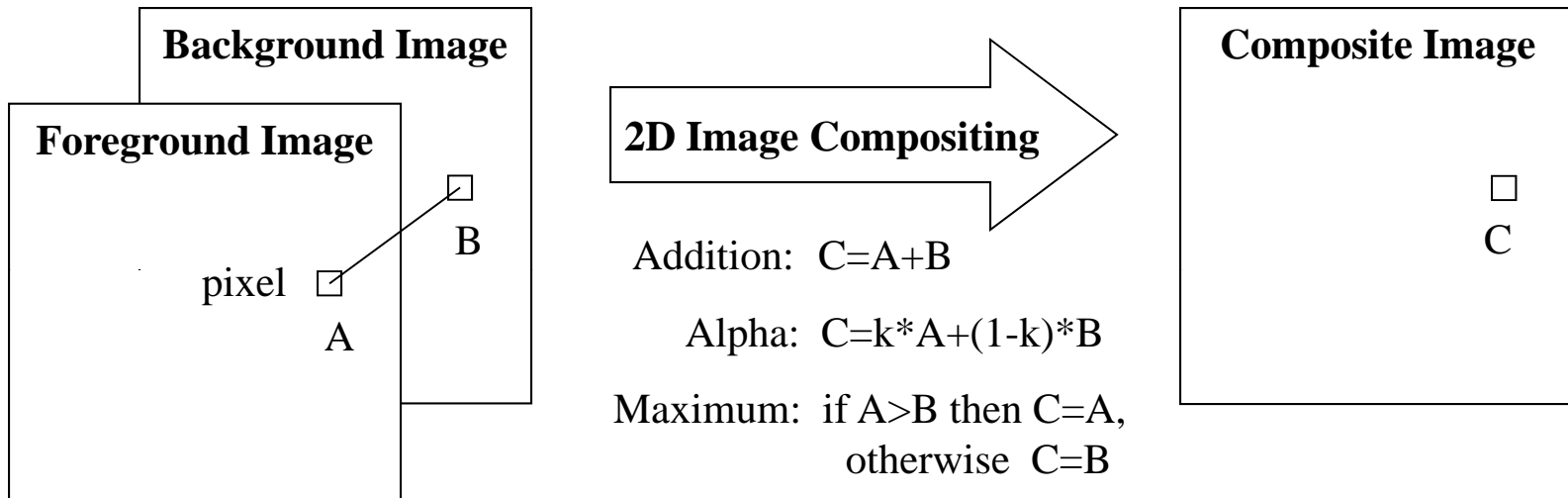
**2) Adjust opacity** – so that you can see into deeper portion of image stack. Only need for alpha blending.

**3) Adjust contrast and brightness** - so that you can see monochrome specimen.

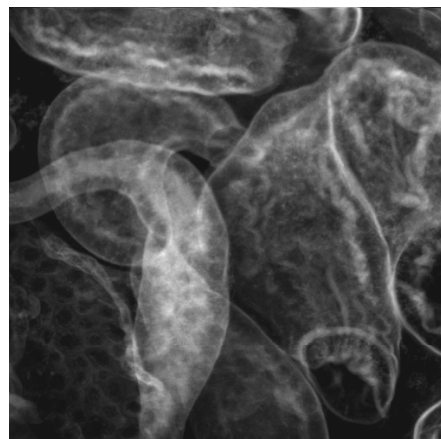
**4) Colorize** – to highlight structures of interest, and/or improve the 3D effect.

# Select Blending Mode (Max, Sum, Alpha)

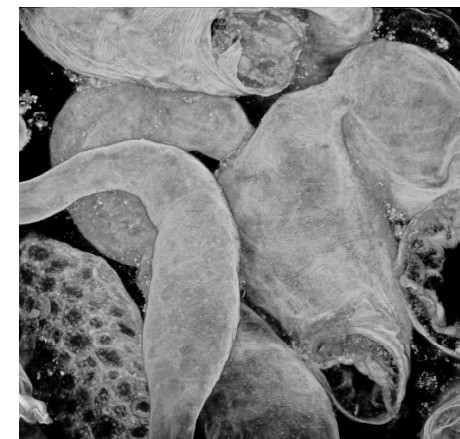
The 2D projections of cross-sectional images are combined as they are rendered in back-to-front order, forming a 2D composite image that is displayed on the monitor



Maximum



Addition

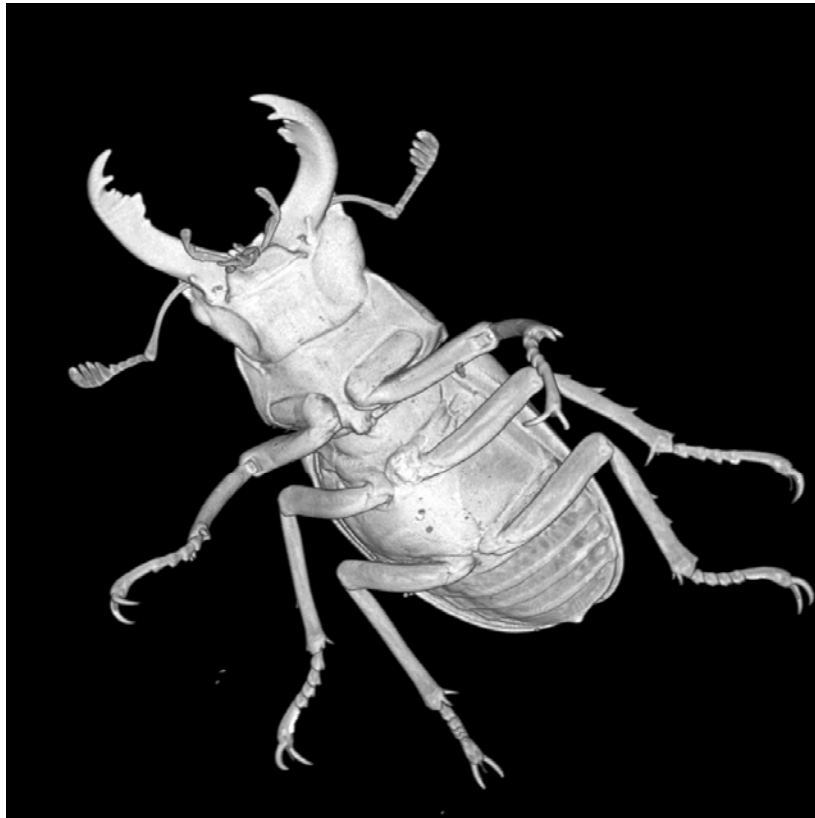


Alpha blending

# Select Blending Mode

The various math operators used to combine projection images can produce very different looking volume renderings, so its important to understand how this works.

MicroCT of stag beetle



Alpha blending

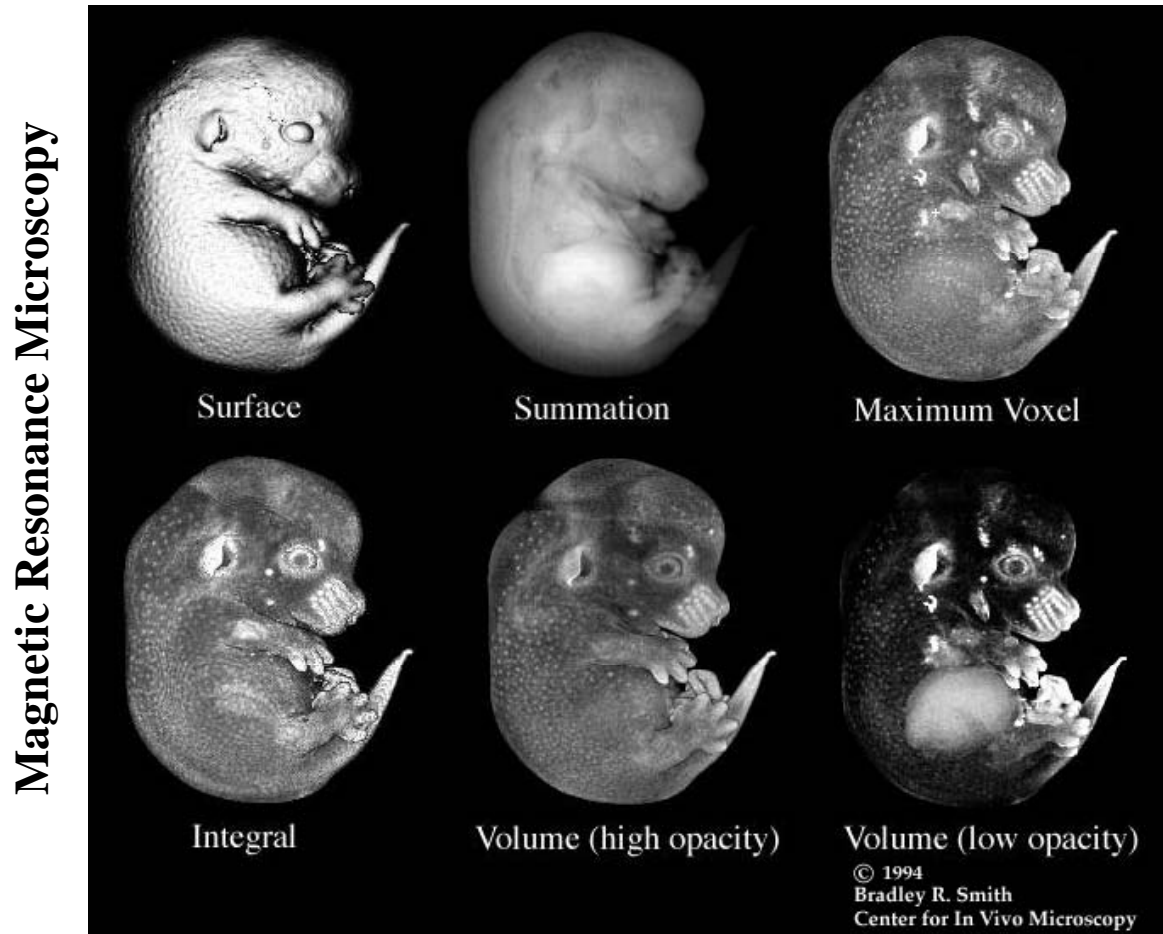


Maximum Intensity Projection

[www.cg.tuwien.ac.at/research/publications/2005/dataset-stagbeetle](http://www.cg.tuwien.ac.at/research/publications/2005/dataset-stagbeetle)

# Select Blending Mode

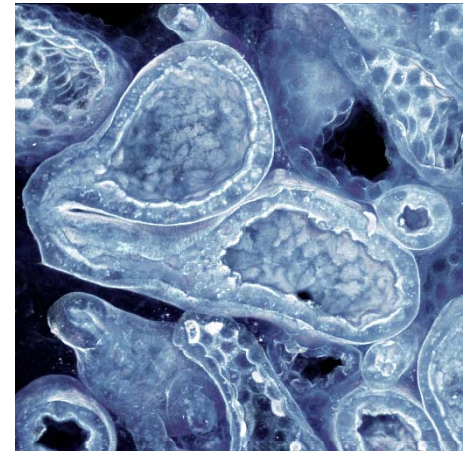
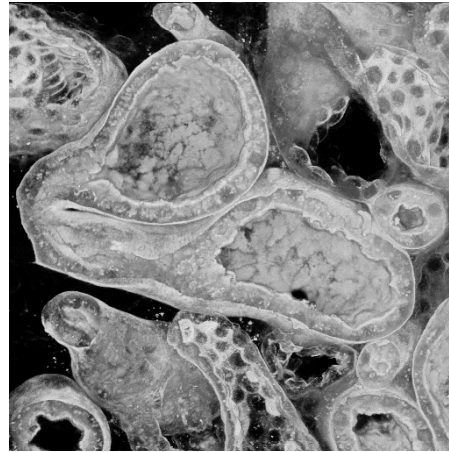
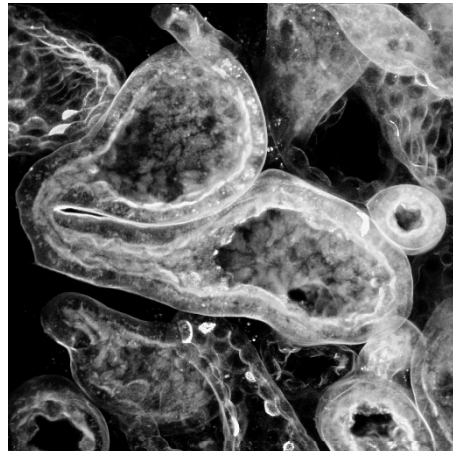
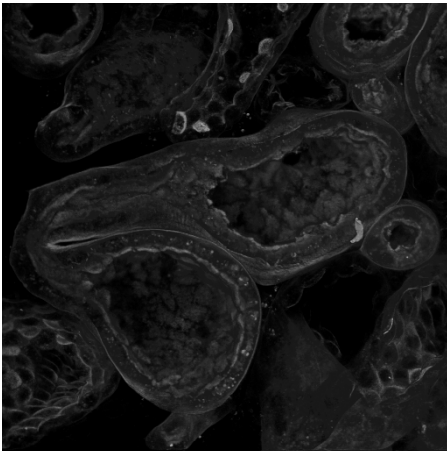
The various math operators used to combine projection images can produce very different looking volume renderings, so its important to understand how this works.



<http://embryo.soad.umich.edu/animal/animalSamples/animalSamples.html>

# Adjust Opacity (only for alpha blending)

Volume rendering programs must be carefully configured to produce a high-quality 3D effect. Here is a typical sequence of operations that needs to be performed:



**1) Select blending mode** – alpha, sum, or max.

**2) Adjust opacity** – so that you can see into deeper portion of image stack. Only need for alpha blending.

**3) Adjust contrast and brightness** - so that you can see monochrome specimen.

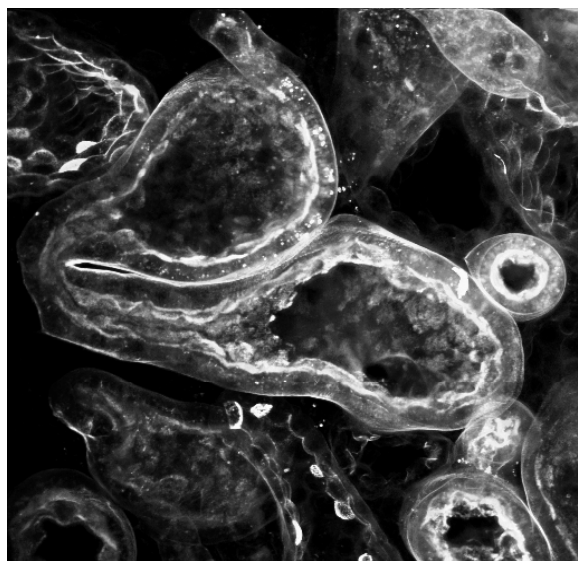
**4) Colorize** – to highlight structures of interest, and/or improve the 3D effect.



# Adjust Opacity

Alpha blending is a weighted averaging procedure used in volume rendering that makes some structures appear more or less opaque

**Small Alpha**



**Medium Alpha**



**Large Alpha**



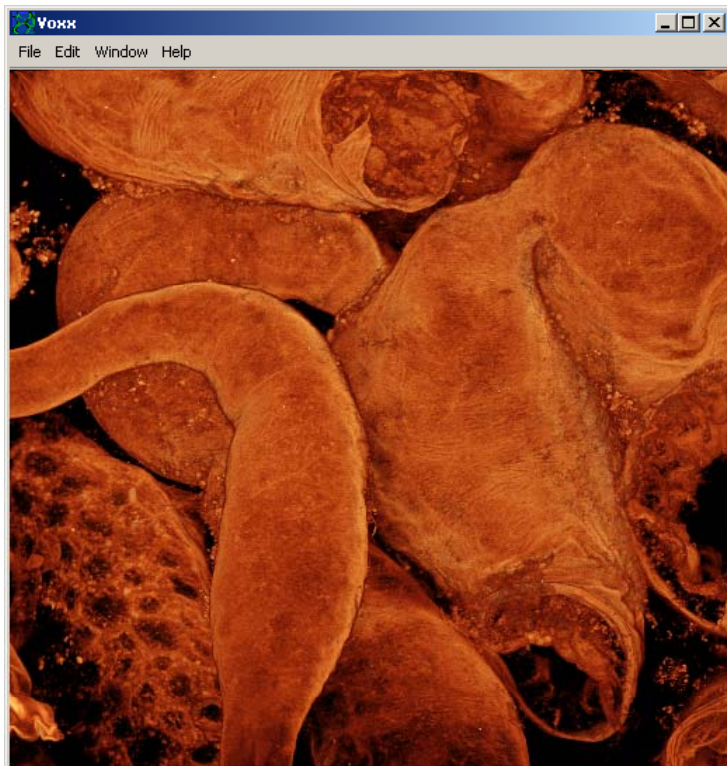
Each pixel in every cross-sectional image has a weight (alpha) associated with it.

Making alpha an increasing function of pixel intensity causes more brightly fluorescing structures to become more visible when the images are averaged.

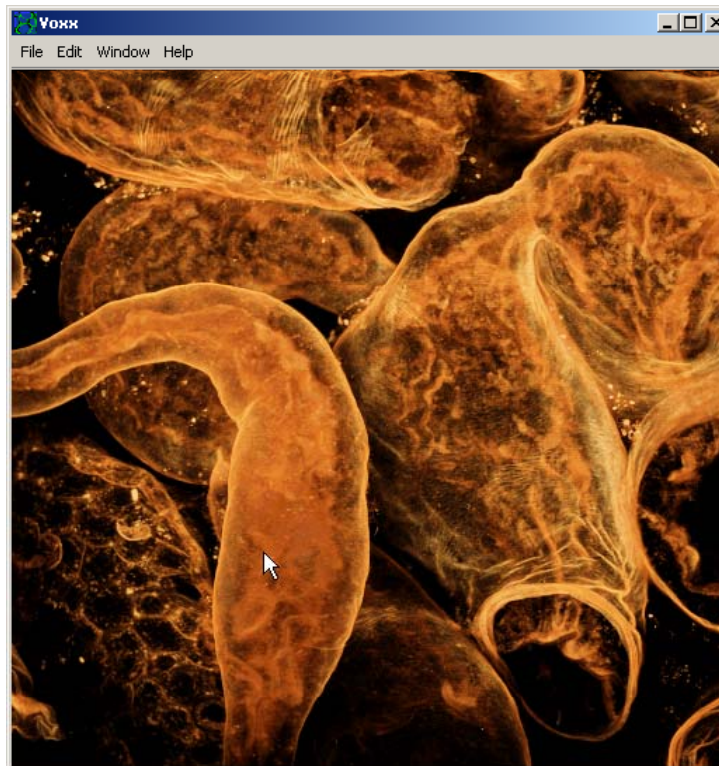
Because images are averaged in back-to-front order, bright foreground structures obscure lower intensity background structures – i.e. brighter objects appear “opaque”.

# Adjust Opacity

Making alpha an increasing function of pixel intensity causes more brightly fluorescing structures to become more visible when the images are averaged



**High Opacity (opaque)**

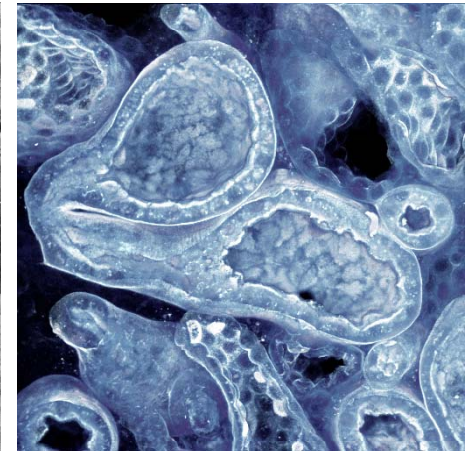
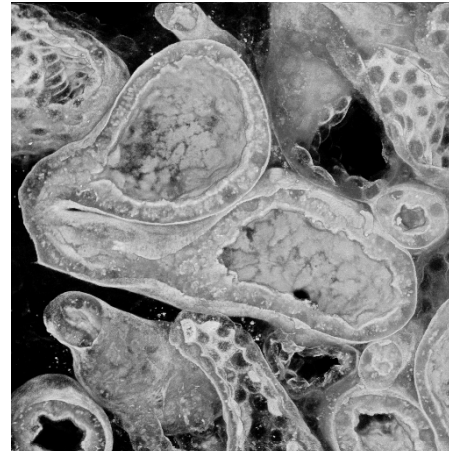
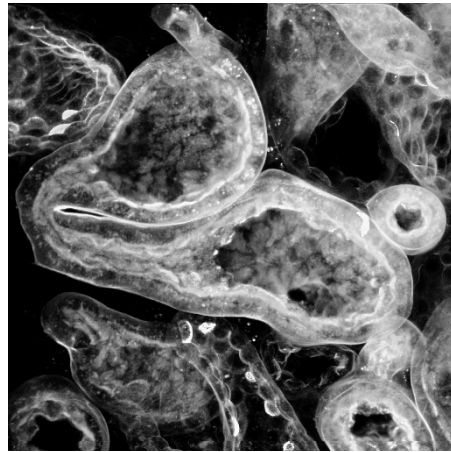
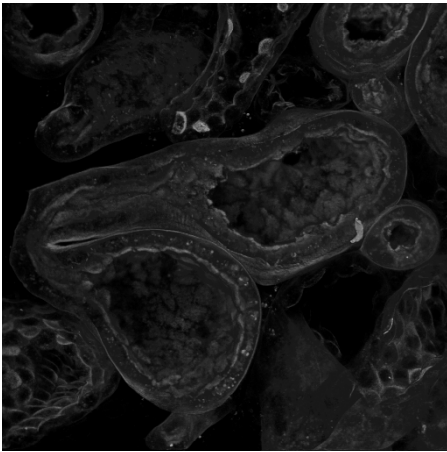


**Low Opacity (translucent)**

e.g. polycystic tubules in which large alpha values allow us to see details on the outer surface, while using smaller alpha values allow us to see the brush border inside the tubules...

# Adjust Brightness and Contrast

Volume rendering programs must be carefully configured to produce a high-quality 3D effect. Here is a typical sequence of operations that needs to be performed:



1) **Select blending mode** – alpha, sum, or max.

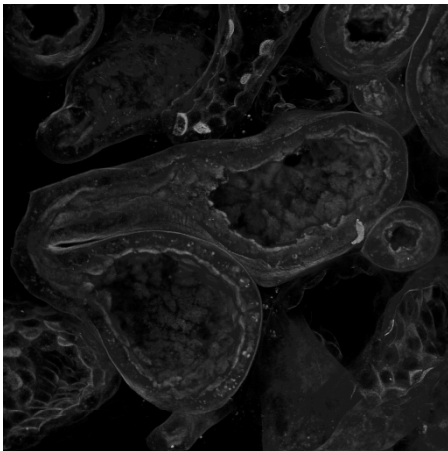
2) **Adjust opacity** – so that you can see into deeper portion of image stack. Only need for alpha blending.

3) **Adjust contrast and brightness** - so that you can see monochrome specimen.

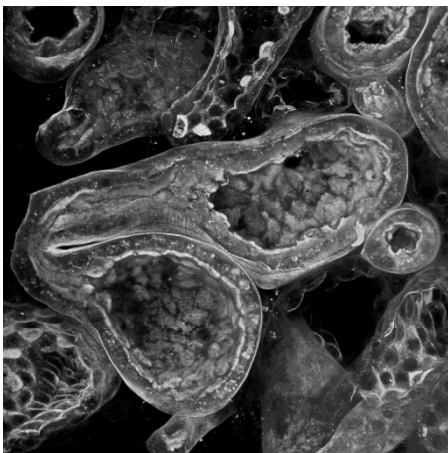
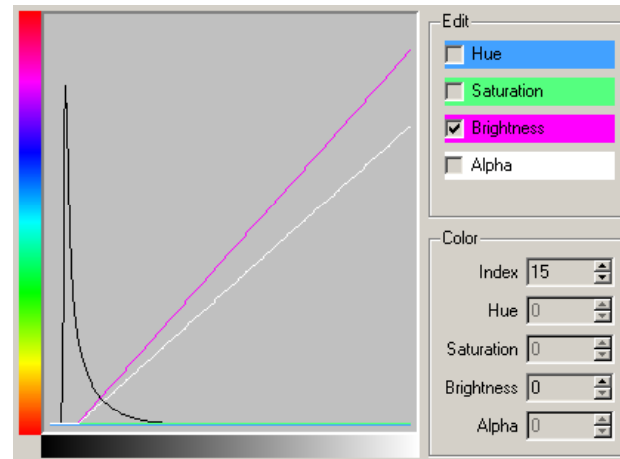
4) **Colorize** – to highlight structures of interest, and/or improve the 3D effect.

# Adjust Brightness and Contrast

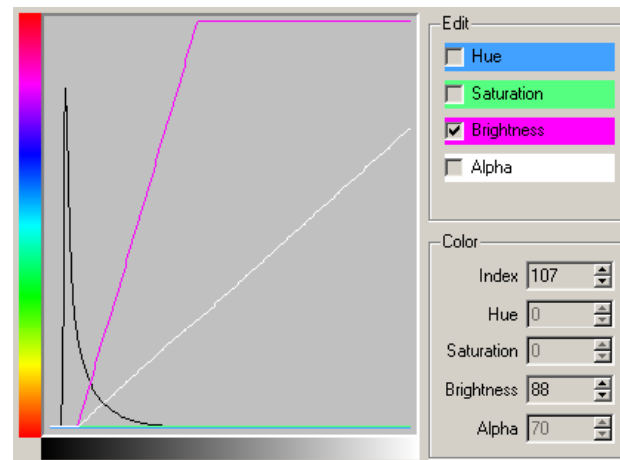
Example of linear transfer functions. Increasing the slope of brightness curve from 1 to 3 increases the brightness and contrast of all pixel values. Also note that x-axis intercept was shifted slightly to the right of the background peak



**Slope=1**

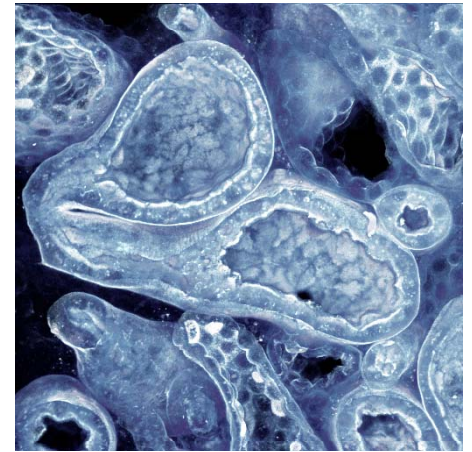
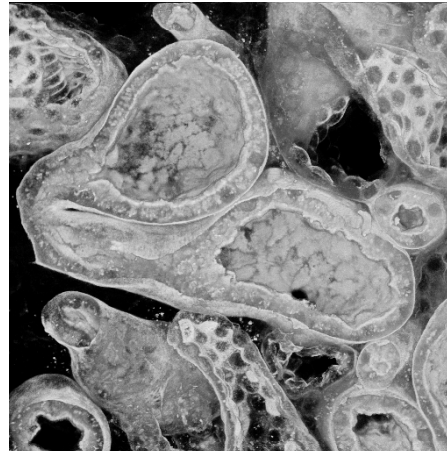
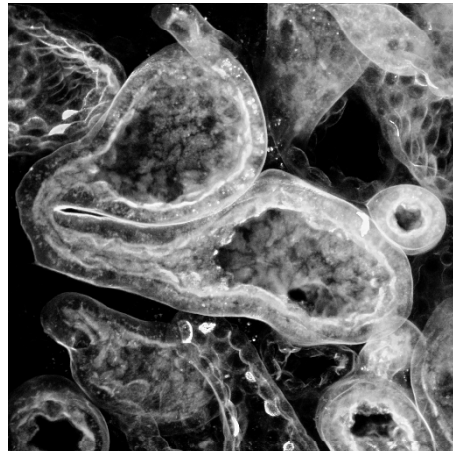
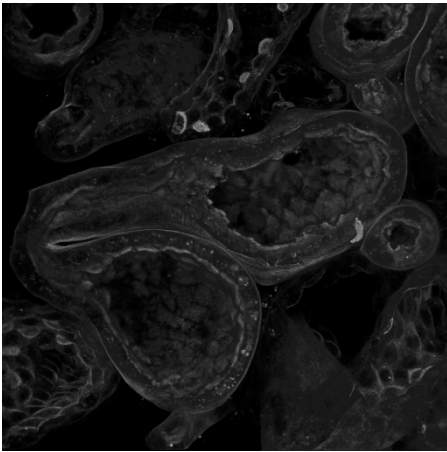


**Slope=3**



# Adjust Color

Volume rendering programs must be carefully configured to produce a high-quality 3D effect. Here is a typical sequence of operations that needs to be performed:



1) **Select blending mode** – alpha, sum, or max.

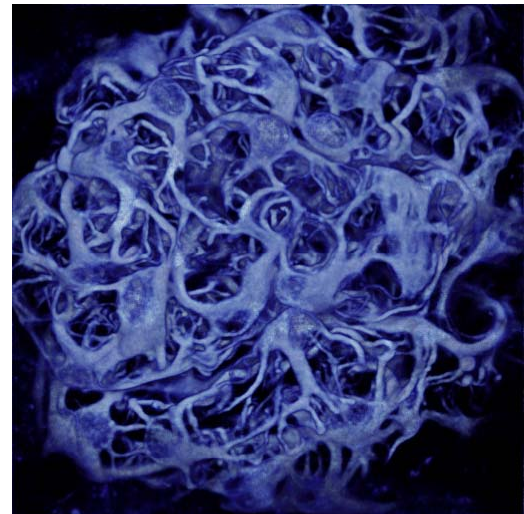
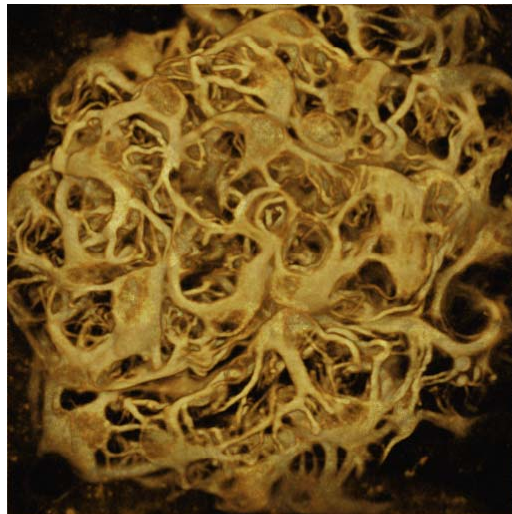
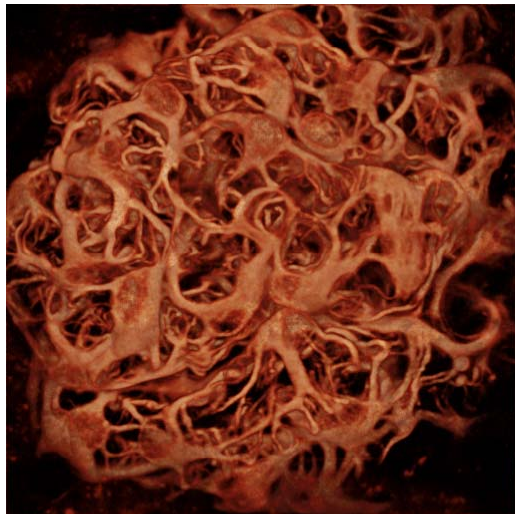
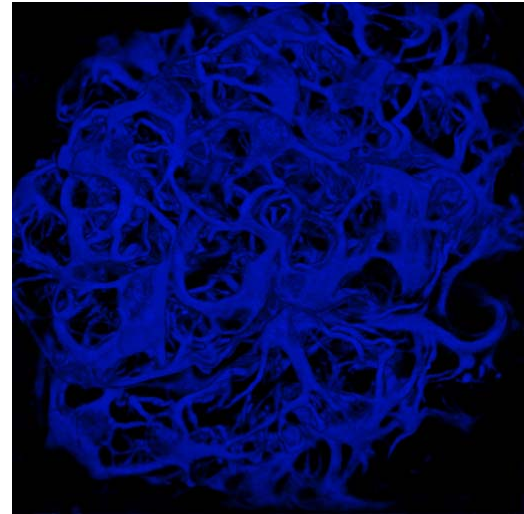
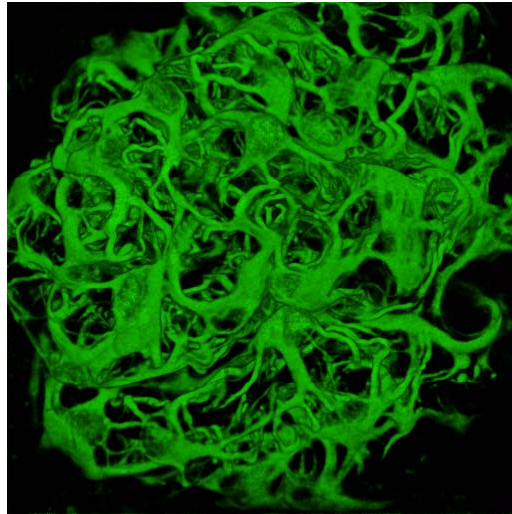
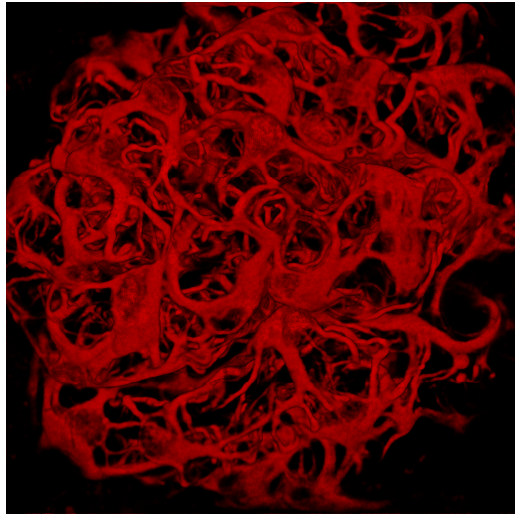
2) **Adjust opacity** – so that you can see into deeper portion of image stack. Only need for alpha blending.

3) **Adjust contrast and brightness** - so that you can see monochrome specimen.

4) **Colorize** – to highlight structures of interest, and/or improve the 3D effect.

# Colorization

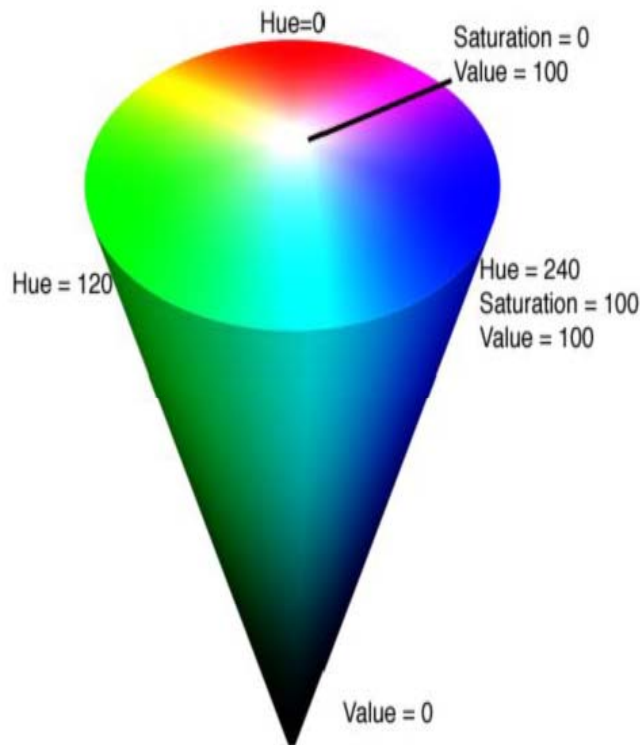
You need to assign each channel a different color when displaying multi-channel images



You must choose appropriate colors for viewing images on monitors and color prints

# Colorization: Adjust Hue and/or Saturation

The RGB representation of color used in monitors and video boards is not easy for people to work with. So other color representations have been developed.

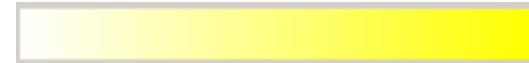


## Hue



determines which spectral color is used. It is often specified as an angle between 0 and 360 degrees, but some programs use 0-1 or 0-255 scales. On a 0-1 scale, red=0, green=120/360, blue=240/360, etc.

## Saturation



determines how much white is added to produce the color.  $S=0$  for a totally desaturated color (i.e. no color),  $S=1$  for a pure color (i.e. no white).

## Value



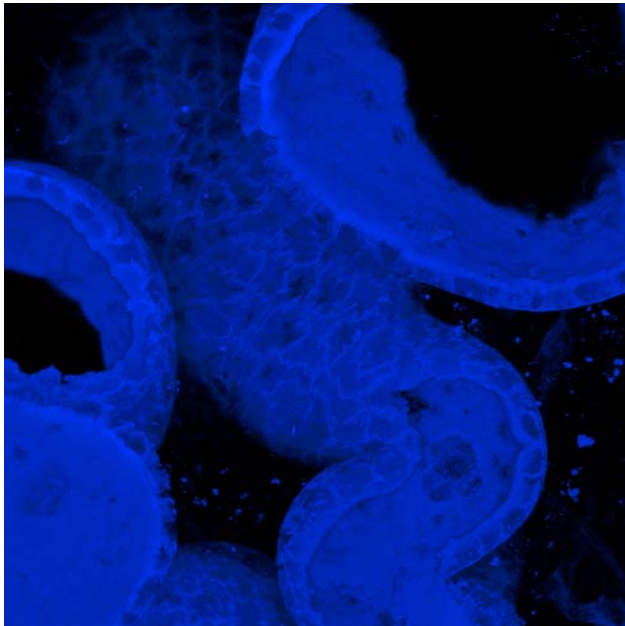
determines the brightness of the color.  $V$  varies between 0 (black) and 1.

Hue, Saturation, and Value (HSV) is commonly used instead of RGB, then converted into RGB by the imaging programs.

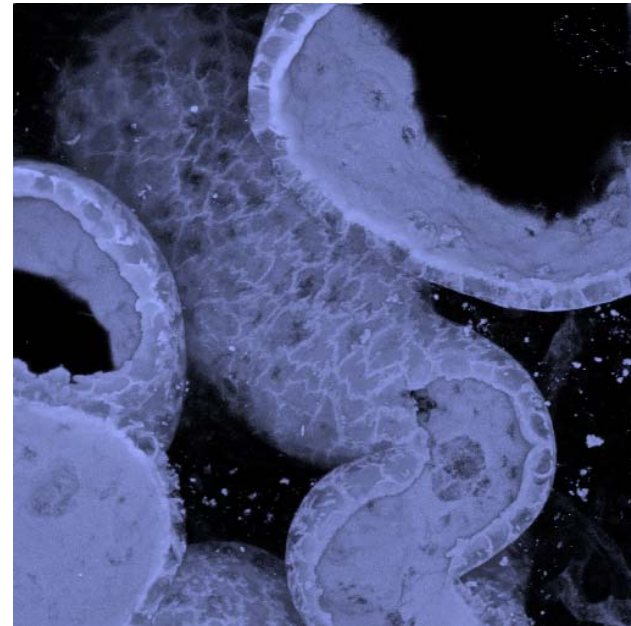
# Colorization: Adjust Hue and/or Saturation

The simplest color choice: pick constant hue and saturation,  
independent of the intensity of the image

**Constant Hue**

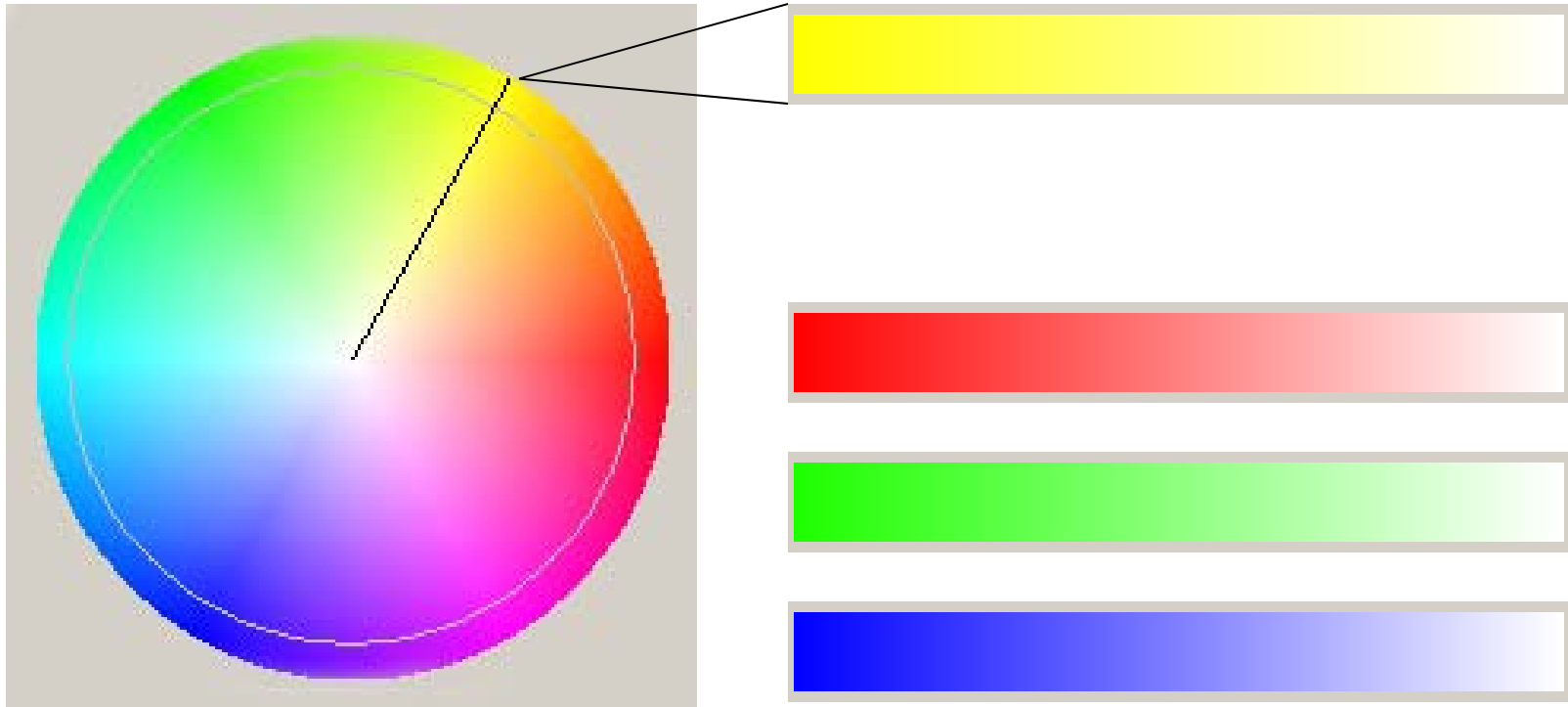


**Constant Saturation**





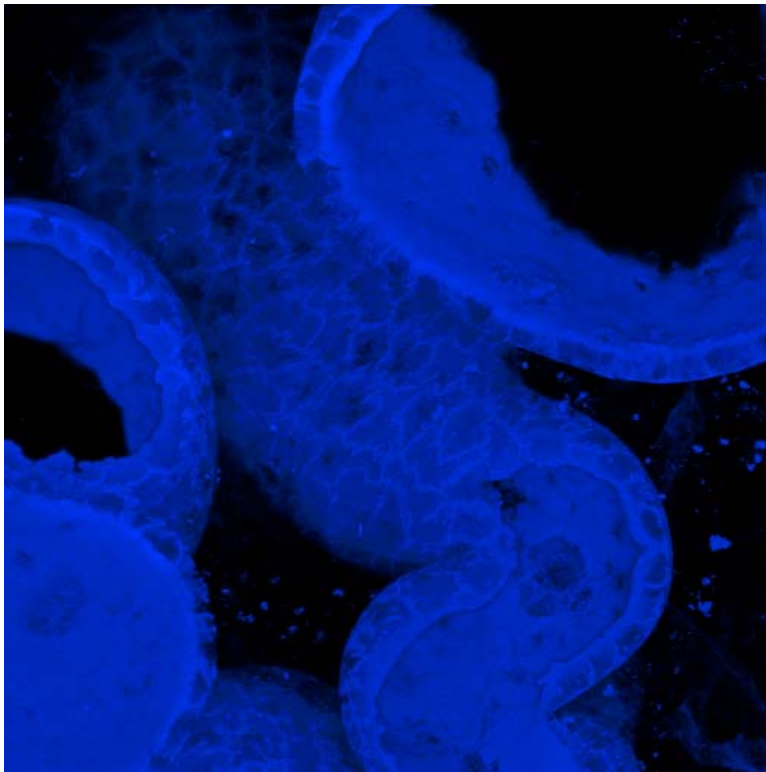
# Colorization: Make Saturation a Function of Intensity



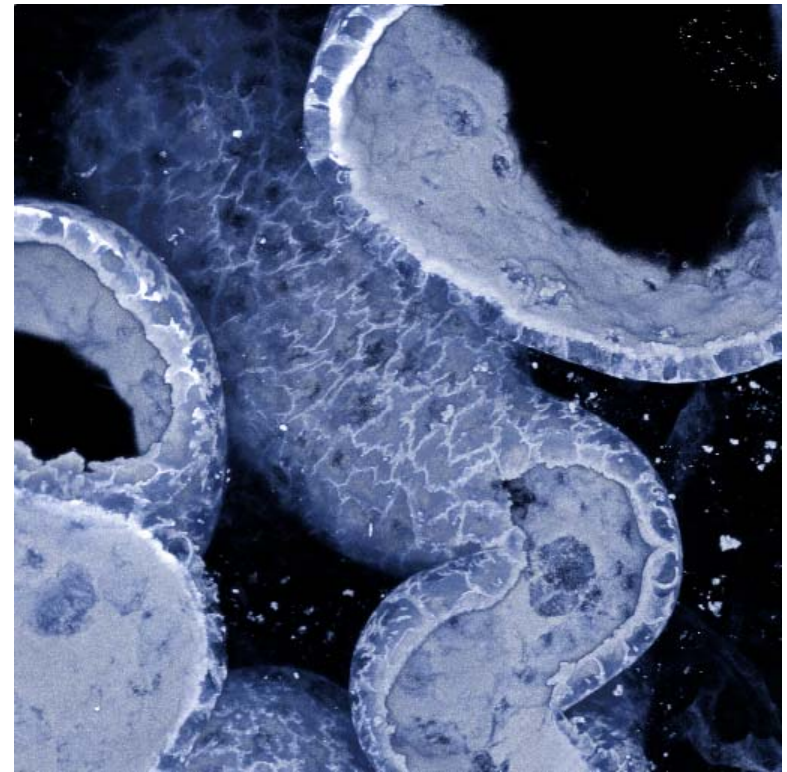
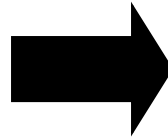
Color scales can provide an additional (weak) depth cue. Darker saturated colors are associated with more distant objects, while lighter less-saturated colors are associated with nearer objects .

# Colorization: Varying the Saturation

Here we improve the visibility of structural details and enhance the 3D effect, by making saturation decrease as the intensity increases in a constant hue image

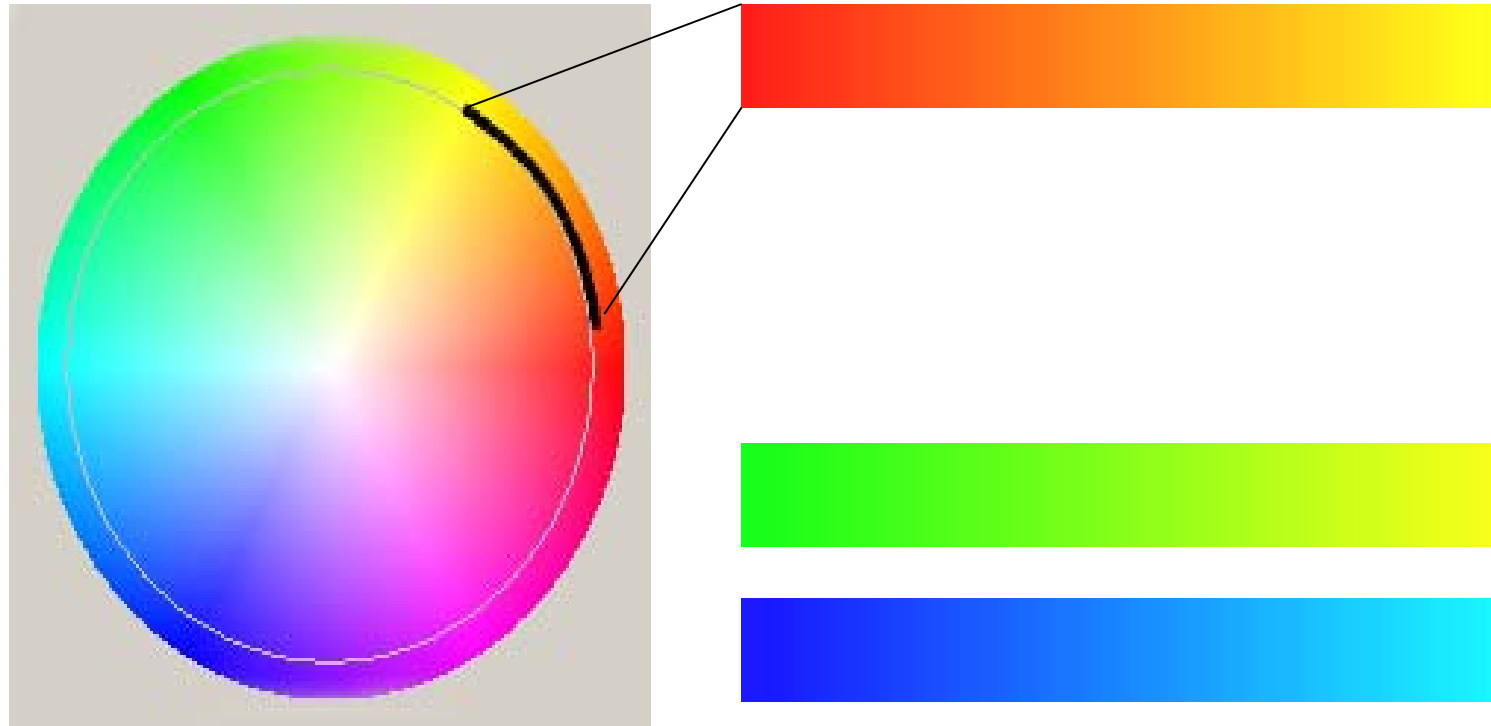


**Both hue and saturation are constant**



**Hue is constant, but saturation varies**

## Colorization: Make Hue a Function of Intensity

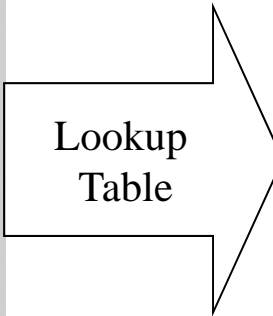


Color scales can provide an additional (weak) depth cue. Darker saturated colors are associated with more distant objects, while lighter less-saturated colors are associated with nearer objects .

# Hue-Based Color Scales

Simultaneously changing the hue and brightness in color scales can produce a better intensity-to-color mapping. Commonly done outside of fluorescence microscopy.

**Crab Nebula (infrared)**



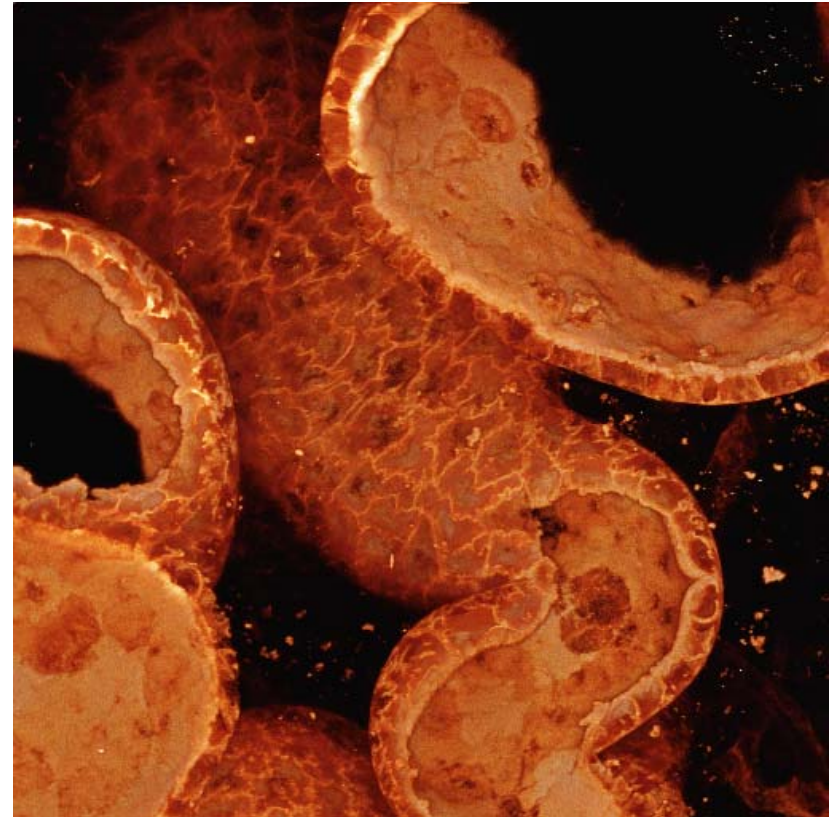
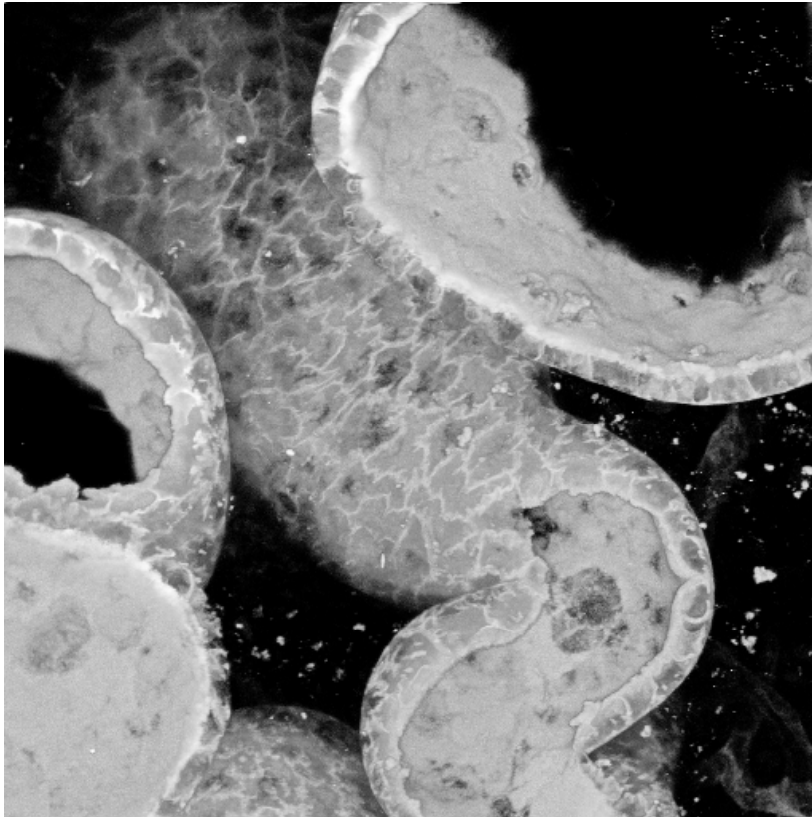
**Colorized Crab Nebula**



© Keck Observatory

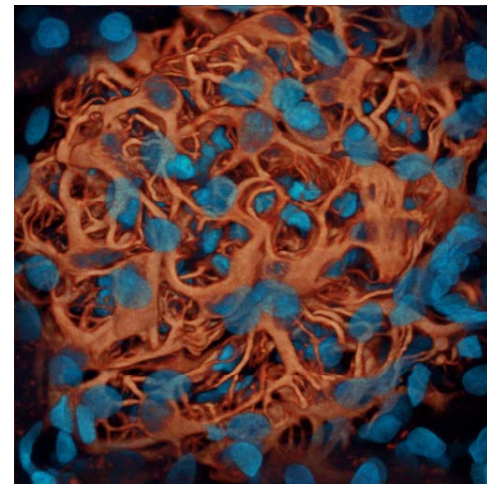
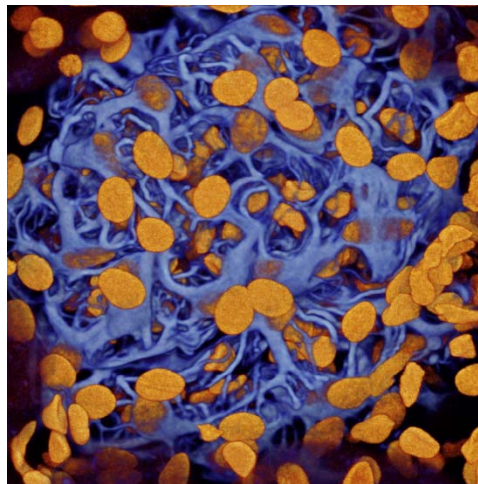
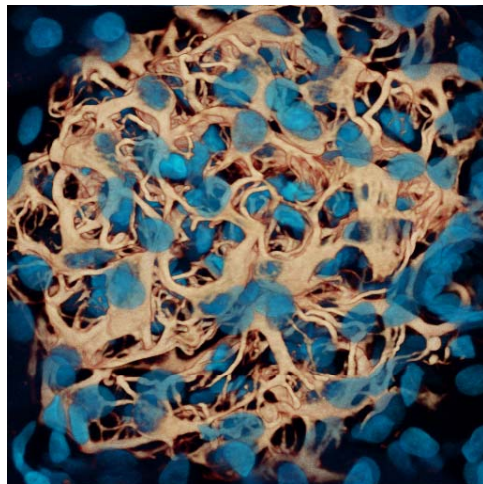
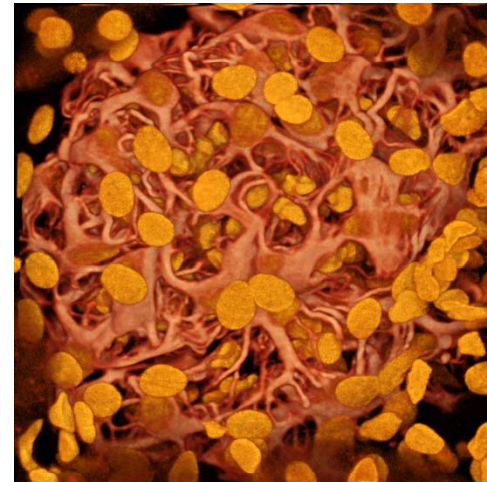
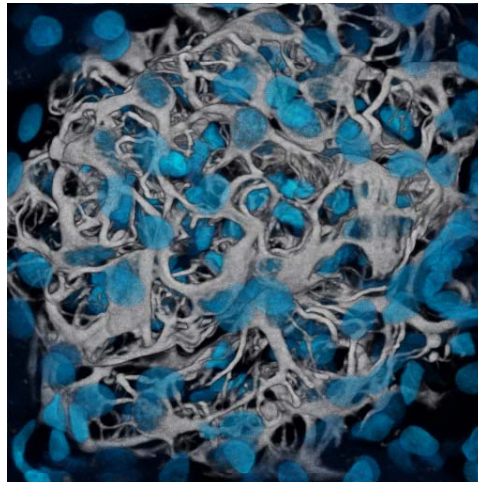
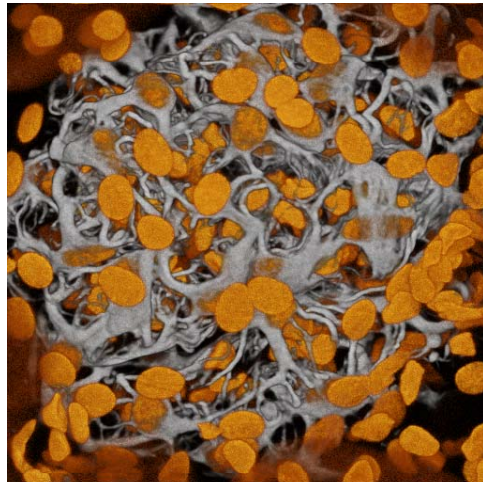
# Modulate Hue and Saturation

Examples of simultaneously changing hue and saturation as a function of brightness, to improve the visibility of structural details and to enhance the 3D effect



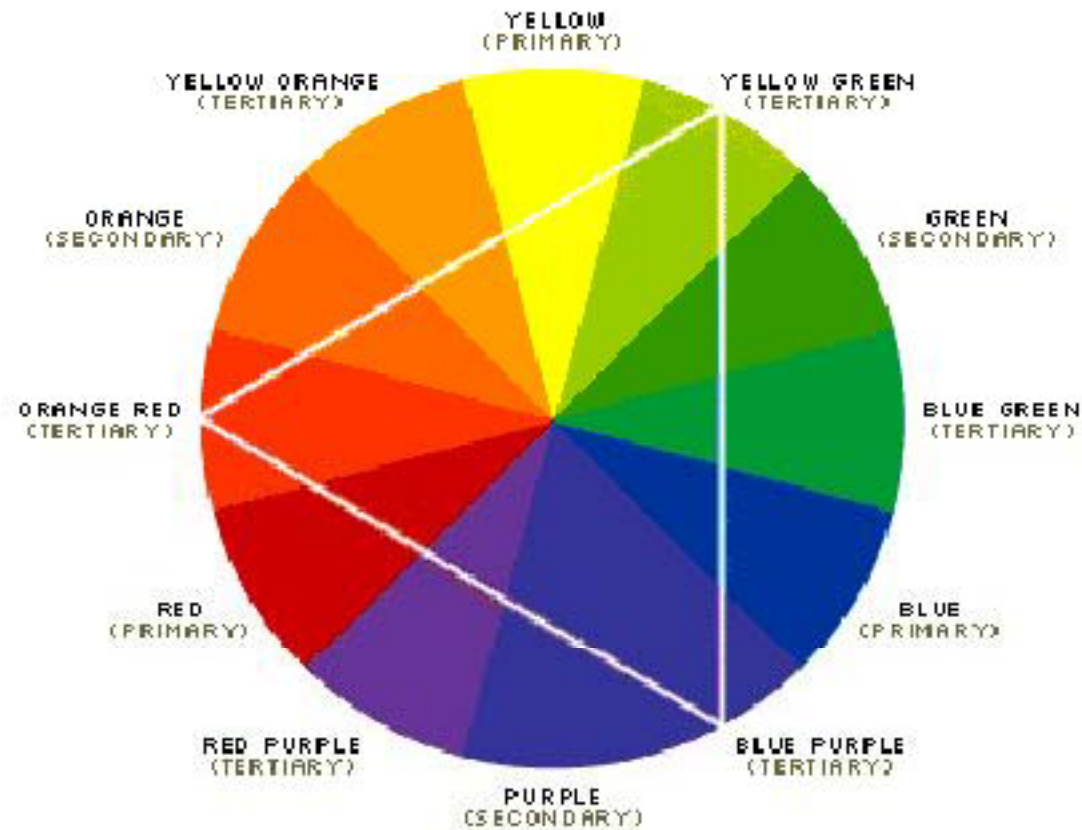
# Colorization: Select Complimentary Colors

One should choose complementary colors when colorizing different structures in images – to improve their visibility, and for aesthetic reasons



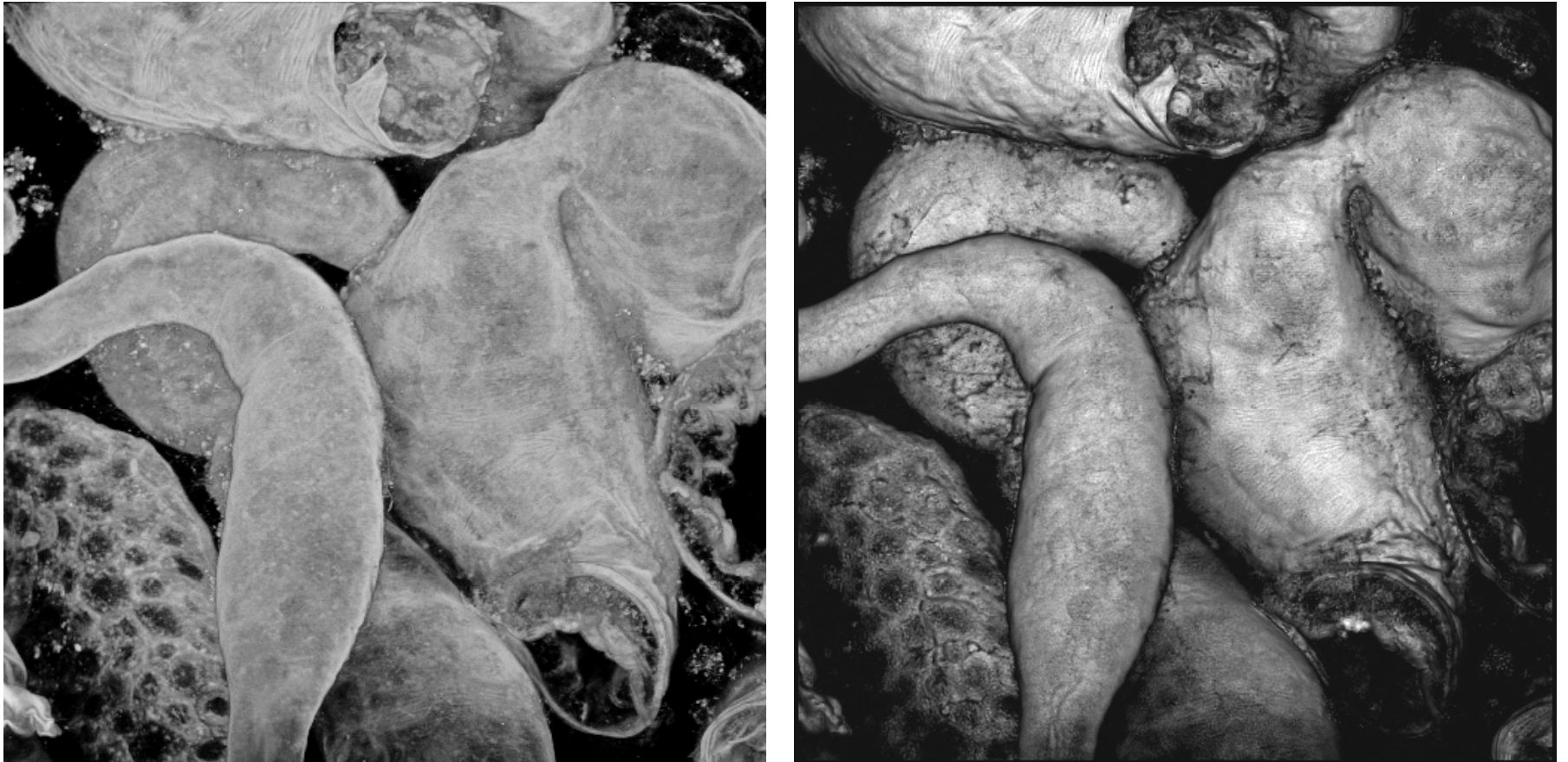
# Colorization: Select Complimentary Colors

One should choose complementary colors when colorizing different structures in images – to improve their visibility, and for aesthetic reasons



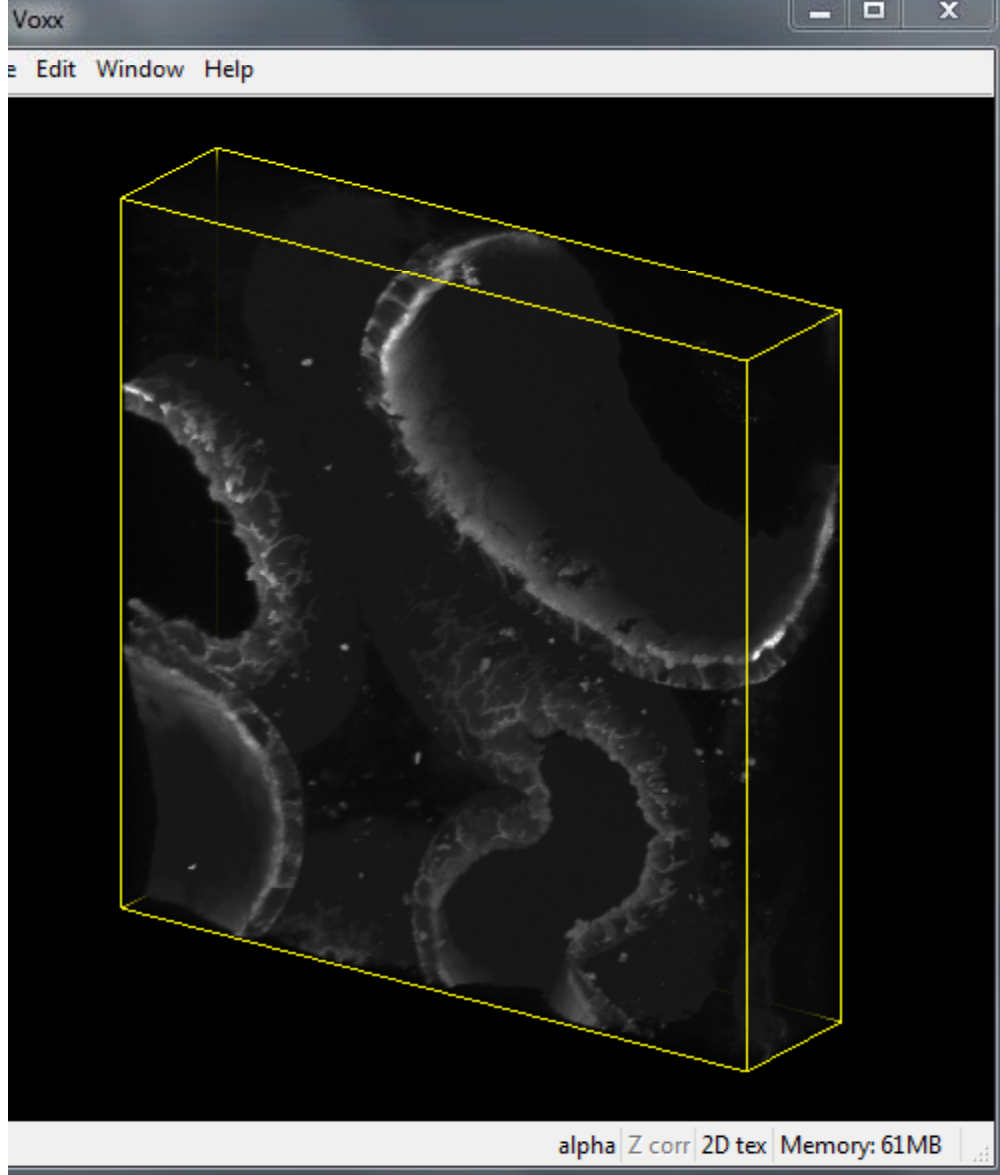
# Lighting

Lighting can produce an improved 3D effect, by providing an additional depth cue and information about the orientation of surfaces of objects



Unlit alpha-blended image of cystic proximal tubules in kidney (left), and simulated illumination using Lambertian model (right).





Palette Editor

Palette 0 - untitled0 HSB ↻

Edit

- Hue
- Saturation
- Brightness
- Alpha

Color

Index 70

Hue 0

Saturation 0

Brightness 94

Alpha 94

Range

From Min 100%

From Max 100%

Auto

Dim dataset

Lazy Redraw

0 255 + -

Axes

Axis X

Scale 1

Power 1

Offset 0

Presets Linear

Curves

Type Power

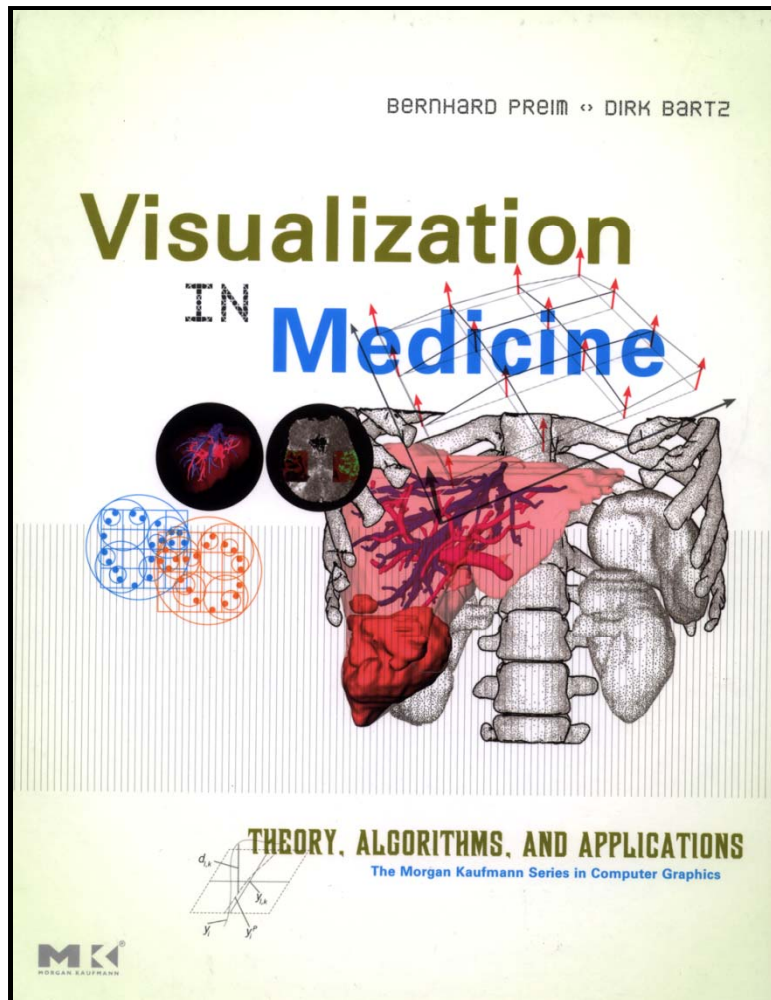
Scale 1

Power 1

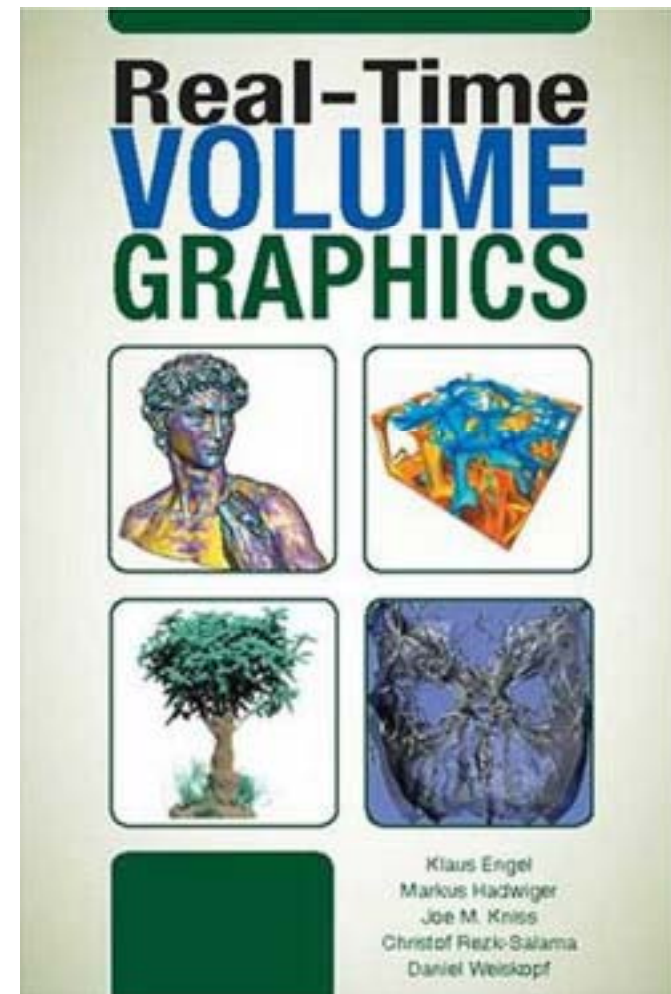
Offset -24

Presets Custom

Are there any good books on this stuff ?



**B. Preim, D. Bartz**  
©2007 Morgan Kaufmann



**K. Engel, M. Hadwiger, et al**  
©2006 A K Peters

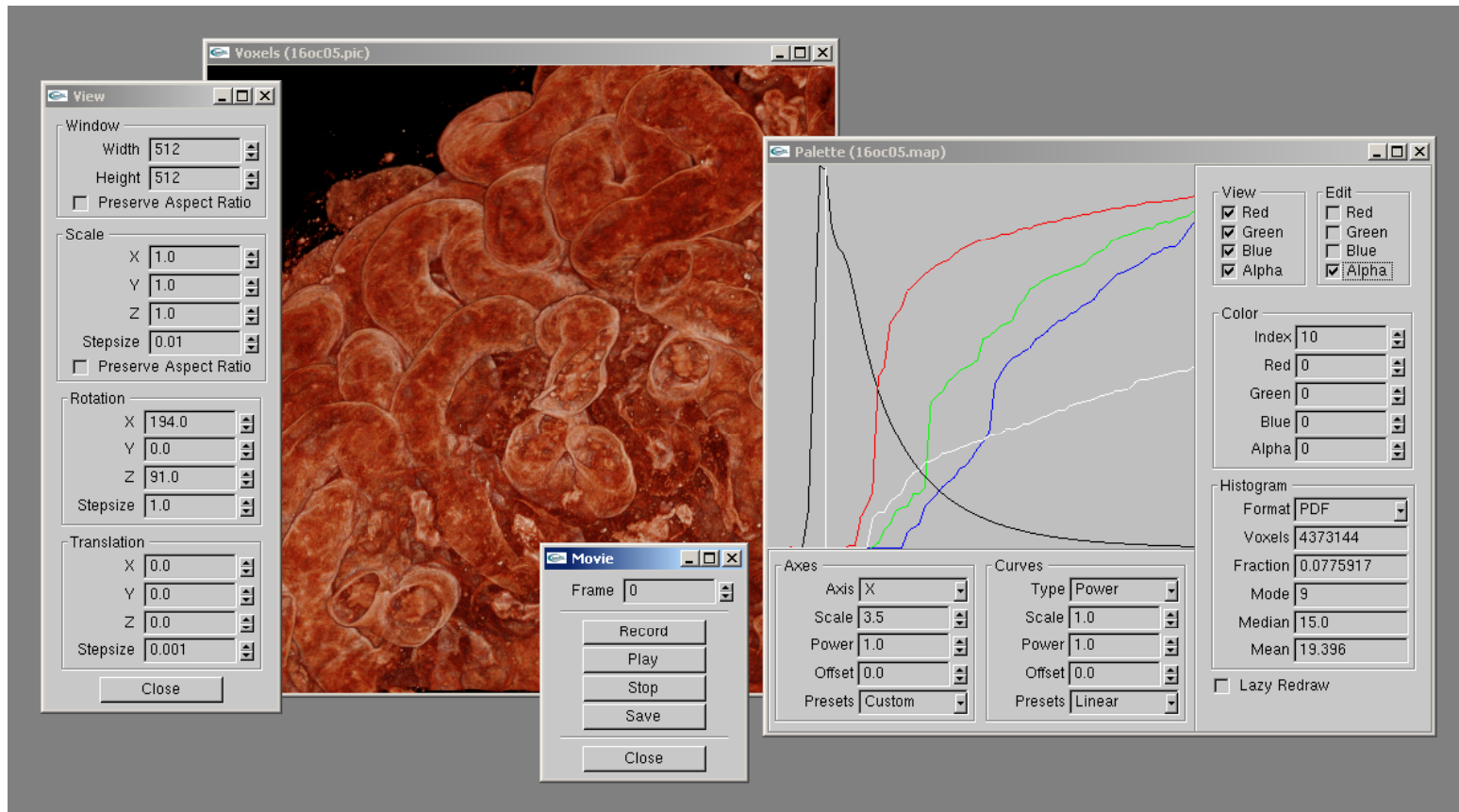
# So what kind of microscopy-oriented 3D imaging software is available ?

There are many 3D image processing programs, but not many can handle the multi-channel 3D and 4D images produced by confocal and 2P microscopes. Here are a few:

<b>Free</b>	<b>BioImageXD</b> <b>Endrov</b> <b>ImageJ</b> <b>VisBio</b> <b>Voxx</b> <b>V3D</b>	<a href="http://www.bioimagexd.net">www.bioimagexd.net</a> <a href="http://www.endrov.net">www.endrov.net</a> <a href="http://rsb.info.nih.gov/ij">http://rsb.info.nih.gov/ij</a> <a href="http://www.loci.wisc.edu/visbio">www.loci.wisc.edu/visbio</a> <a href="http://www.nephrology.iupui.edu/imaging/voxx">www.nephrology.iupui.edu/imaging/voxx</a> <a href="http://penglab.janelia.org/proj/v3d">http://penglab.janelia.org/proj/v3d</a>
<b>Commercial</b>	<b>Amira</b> <b>AutoQuant X</b> <b>Huygens</b> <b>Image-Pro Plus</b> <b>Imaris</b> <b>Metamorph</b> <b>Volocity</b>	<a href="http://www.amiravis.com">www.amiravis.com</a> <a href="http://www.mediacy.com">www.mediacy.com</a> <a href="http://www.svi.nl">www.svi.nl</a> <a href="http://www.mediacy.com">www.mediacy.com</a> <a href="http://www.bitplane.com">www.bitplane.com</a> <a href="http://www.moleculardevices.com">www.moleculardevices.com</a> <a href="http://www.improvision.com/products/volocity">www.improvision.com/products/volocity</a>

# Voxx

If any of you want to experiment with a volume renderer using confocal microscope images - you could download Voxx, along with sample data sets...

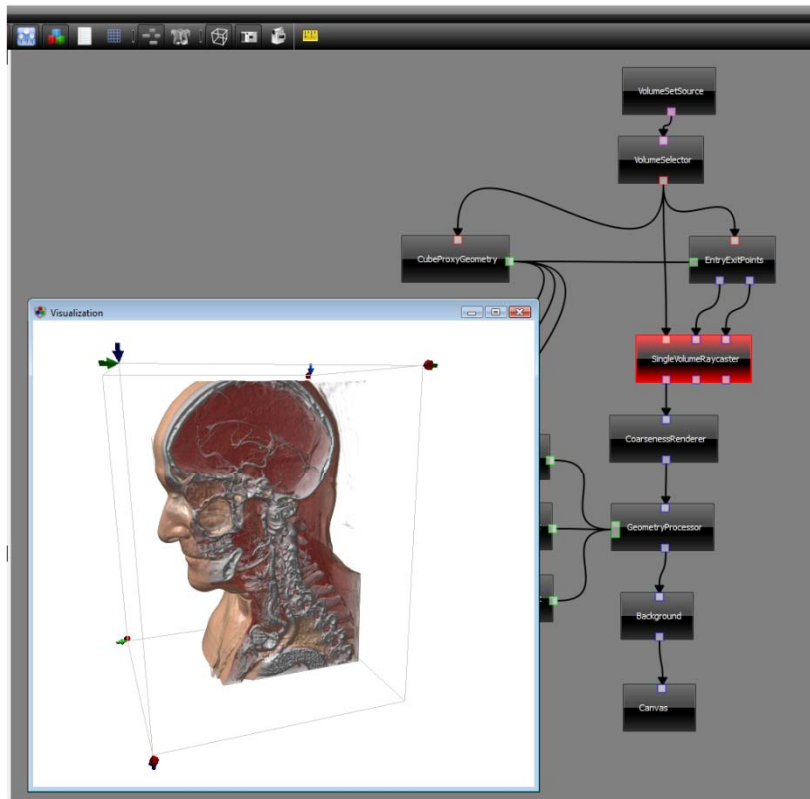


<http://nephrology.iupui.edu/imaging/voxx>

[http://nephrology.iupui.edu/imaging/voxx/sample\\_data](http://nephrology.iupui.edu/imaging/voxx/sample_data)

# Voreen and Slicer

If any of you want to experiment with a volume renderer using CT or MRI images - you can download Voreen or 3D Slicer, along with sample data sets...



[www.slicer.org](http://www.slicer.org)

[www.voreen.org](http://www.voreen.org)

[www.voreen.org/108-Data-Sets.html](http://www.voreen.org/108-Data-Sets.html)

# And what kind of hardware do I need ?



In the early days of high-performance 3D imaging (1990s), volume rendering programs had to be run on very expensive Silicon Graphics workstations if you wanted real-time rendering. Typical prices for SGI's systems ranged from \$30K to \$200K+.

Now, any PC with a low-cost (\$100-400) video board equipped with a GeForce or Radeon graphics processor can do real-time 3D rendering.

# Questions?

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